

# 1 Occurrence, fate and fluxes of plastics and microplastics in 2 terrestrial and freshwater ecosystems

3  
4 Theresa Schell<sup>1</sup>, Andreu Rico<sup>1</sup>, Marco Vighi<sup>1</sup>

5  
6  
7 <sup>1</sup>IMDEA Water Institute, Science and Technology Campus of the University of Alcalá,  
8 Avenida Punto Com 2, 28805, Alcalá de Henares, Madrid, Spain

9  
10  
11 Corresponding author:

12 Theresa Schell

13 IMDEA Water Institute, Avda Punto Com 2, 28805, Alcalá de Henares, Madrid, Spain

14 Email: [theresa.schell@imdea.org](mailto:theresa.schell@imdea.org)

15 Phone: +34 918305962

16  
17  
18 E-mail: [andreu.rico@imdea.org](mailto:andreu.rico@imdea.org), [marco.vighi@imdea.org](mailto:marco.vighi@imdea.org)

## 19 20 21 **Abstract**

22  
23 Plastics and microplastics are nowadays ubiquitously found in the environment. This has  
24 raised concerns on possible adverse effects for human health and the environment. To date,  
25 extensive information exists on their occurrence in the marine environment. However,  
26 information on their different sources and their transport within and across different  
27 freshwater and terrestrial ecosystems is still limited. Therefore, we assessed the current  
28 knowledge regarding the industrial sources of plastics and microplastics, their environmental  
29 pathways and load rates, and their occurrence and fate in different environmental  
30 compartments; thereby highlighting important data gaps which are needed to better describe  
31 their global environmental cycle and exposure. This study shows that the quantitative  
32 assessment of the contribution of the different major sources of plastics, microplastics and  
33 nanoplastics to aquatic and terrestrial ecosystems is challenged by some data limitations.  
34 While the presence of MPs in wastewater and freshwater is relatively well studied, data on  
35 sediments and especially soil ecosystems are too limited. Moreover, the overall occurrence  
36 of large-size plastics, the patterns of microplastic and nanoplastic formation from them, the  
37 presence and deposition of plastic particles from the atmosphere, and the fluxes of all kinds  
38 of plastics from soils towards aquatic environments (e.g. by surface water runoff, soil  
39 infiltration) are still poorly understood. Finally, this study discusses several research areas  
40 that need urgent development in order to better understand the potential ecological risks of  
41 plastic pollution, and provide some recommendations to better manage and control plastic  
42 and microplastic inputs into the environment.

43  
44 **Keywords:** plastics, microplastics, nanoplastics, occurrence, environmental exposure, risk  
45 management, remediation, surface waters, wastewater, sludge, sediment, soil, agriculture,  
46 air, environmental fate, emissions, pollution, water quality, ecotoxicology, impacts, sampling  
47 methods, anthropogenic activity, plastic emission, persistence, synthetic fibres

53	<b>Table of contents</b>
54	
55	<b>1. Introduction</b>
56	<b>2. Environmental sources of plastics</b>
57	<b>3. Pathways of plastic to the environment</b>
58	<b>3.1. Collected solid waste</b>
59	<b>3.2. Wastewater</b>
60	<b>3.3. Sludge and other agricultural amendments</b>
61	<b>4. Occurrence and fluxes of plastics in environmental compartments</b>
62	<b>4.1. Air</b>
63	<b>4.2. Soil</b>
64	<b>4.3. Surface waters</b>
65	<b>4.4. Sediments</b>
66	<b>4.5. Marine</b>
67	<b>5. Discussion</b>
68	<b>5.1. The need for standardization of sampling and analysis</b>
69	<b>5.2. Small size micro-nanoplastics: the largest unknowns</b>
70	<b>5.3. Towards a microplastic mass balance and suitable evaluation of</b>
71	<b>environmental fluxes</b>
72	<b>5.4. Macroplastic breakdown</b>
73	<b>5.5. Complete degradation of MPs</b>
74	<b>5.6. Microplastic concentration in environmental compartments: what</b>
75	<b>does it mean in terms of exposure for living organisms?</b>
76	<b>6. Conclusion</b>
77	<b>6.1. Do MPs represent a threat for human health and for the</b>
78	<b>environment?</b>
79	<b>6.2. How can microplastic inputs in the environment be controlled?</b>
80	<b>7. List of abbreviations</b>
81	<b>8. Acknowledgements</b>
82	<b>9. References</b>
83	
84	
85	
86	
87	
88	
89	
90	
91	
92	
93	
94	
95	
96	
97	
98	
99	
100	
101	
102	
103	
104	
105	

## 1. Introduction

Over the past century, plastic has made the journey from being virtually non-existent to a ubiquitous and integral part of modern life. While plastic has numerous advantages compared to alternative materials, we are facing severe environmental, economic and ethical issues due to the vast plastic waste production and rapid disposal. Up until 2015, the total amount of plastic produced was 8300 million tons, 6300 million tons of which were discarded as waste (Geyer et al. 2017). Much of this waste (79%) is accumulated in landfills or the natural environment, and this amount is expected to increase significantly in the future (up to 12,000 million tons by 2050) if management actions are not immediately taken (Geyer et al. 2017).

Most macroplastics (MaPs) break down due to mechanical and chemical fragmentation into smaller pieces, which are commonly termed microplastics (particles < 5 mm; NPs) or nanoplastics (particles < 1 $\mu$ m; NPs) (Gigault et al. 2018). The breakdown process may take between 50-600 years and usually depends on several factors such as the polymer composition and the environmental condition. MPs that result from the breakdown of MaP are commonly referred to as secondary MPs, while MPs produced in this size range are referred to as primary MPs. Nowadays, MaPs, MPs and NPs can be found floating or in suspension in many water bodies, accumulated in sediments or in terrestrial ecosystems, and even can be transported and deposited in pristine environments due to wind and currents (Dris et al. 2015; Ballent et al. 2016; Dris et al. 2016; Fischer et al. 2016; Hurley and Nizzetto 2018).

The widespread distribution of plastic and its variability in size and shape allow the ingestion by organisms across many trophic levels and habitats (Wright et al. 2013; Kühn et al. 2015). Large plastic debris (MaPs) can cause adverse effects on coastal and marine animals (marine mammals, fish and seabirds) due to ingestion as well as to entanglement which impedes their mobility (Van Franeker et al. 2011; Knowlton et al. 2012; Schuyler et al. 2012; Kühn et al. 2015). Fishing gear, balloons, plastic bags and bottle caps have been identified to be the most harmful type of MaPs to marine organisms (Hardesty et al. 2015). Although most research has focused on the marine environment, freshwater and terrestrial organisms are expected to suffer from the same sort of effects. For example, cattle have been reported to suffocate and die due to the ingestion of plastic bags, which can block airways and stomachs (Ramaswamy and Sharma 2011).

Similar to MaPs, environmental exposure to MPs has raised concerns about their potentially adverse effects in smaller organisms. Ecotoxicological studies with MPs have been primarily conducted using marine organisms (77%), while freshwater organisms have been less researched (23%) (de Sá et al. 2018), and research involving terrestrial organisms is still in its beginnings (Chae and An 2018). MPs may cause physical effects such as

144 internal and external abrasion or blockages of the digestive tract in small invertebrates and  
145 fish (Wright et al. 2013; Karami et al. 2016; Jovanović 2017). Research also shows that MP  
146 ingested by freshwater organisms may reduce their feeding efficiency and lower the energy  
147 uptake, which often results in reduced growth, reproduction and survival (Foley et al. 2018).  
148 In addition MPs may affect the growth, chlorophyll content, photosynthesis activity and  
149 reactive oxygen species of microalgae at high, currently not realistic, concentrations (Prata et  
150 al. 2019).

151 Although several cases evidence deleterious impacts of MaPs on aquatic and  
152 terrestrial organisms under laboratory conditions, the capacity of MPs or even NPs to pose a  
153 real threat for ecosystems and human health is disputable. This is because the majority of  
154 studies showing some impacts of MPs on terrestrial or freshwater organisms have been  
155 performed with very high exposure concentrations, while risk at environmentally relevant  
156 concentrations has yet to be disclosed (Lenz et al. 2016).

157 Despite physical effects, some MaPs and MPs have been reported to induce  
158 endocrine disrupting effects (Rochman et al. 2014) due to the release of plastic additives  
159 such as phthalates, chlorinated paraffins and bisphenols (Stenmarck et al. 2017).  
160 Hydrophobic pollutants (e.g. some pesticides, PCBs, PAHs) can also be adsorbed to plastics  
161 and may be released into the body of the organisms after ingestion, leading to the so-called  
162 Trojan Horse effect (Teuten et al. 2009; Koelmans et al. 2016; Crawford and Quinn 2017;  
163 Bouhroum et al. 2019). → add something on plastic being sink for chemicals upon ingestion  
164 Furthermore, MPs could not only act as carriers for chemicals, but also can transport  
165 bacteria or pathogens attached to them (Keswani et al. 2016; Kirstein et al. 2016) across  
166 different environmental compartments and regions.

167 The continuous emission patterns and the breakdown of plastic litter into smaller  
168 fractions in the environment may contribute to future concentrations that are orders of  
169 magnitude higher than the ones currently monitored (Everaert et al. 2018), thus contributing  
170 to a yet uncertain risk scenario. Policies dedicated to control emissions and manage risks of  
171 MaPs, MPs and NPs in the environment require a proper understanding of the main emission  
172 routes, the current exposure levels and the fluxes among environmental compartments. The  
173 available literature describing the exposure and impacts of plastics in the environment has so  
174 far mainly focused on specific emission routes and local monitoring campaigns, and do not  
175 provide a comparative assessment of the whole occurrence, transport and fate of plastics in  
176 different compartments, which is key to identify suitable management actions.

177  
178 Therefore, this study aimed to assess the state of the knowledge regarding the overall  
179 sources of plastic and its occurrence, fate, fluxes and loads into and in different

180 environmental compartments of terrestrial and freshwater ecosystems. This study identifies  
181 data gaps that need to be addressed in order to understand the life cycle of the different  
182 plastic types in the environment, particularly in the soil-water interface, and provides crucial  
183 information to support research into the accumulation and ecotoxicological characterization  
184 of plastics to living organisms. Ultimately, this study provides guidance information to derive  
185 effective management measures aimed at reducing plastic discharges into the environment  
186 and to attain a more sustainable use and consumption of plastics in the nearby future.

## 187 **2. Environmental sources of plastics**

188 Nowadays, Asia is producing 50% of the world's plastic, followed by Europe and North  
189 America, producing 19% and 18%, respectively (PlasticsEurope 2018). The majority of  
190 plastics can be classified into the two main categories: thermoplastics (pellets that are re-  
191 melted to manufacture the final product), and thermoset plastics (thermally produced into the  
192 commercial shape). Thermoplastics constitute 80% of the total plastic and are the main  
193 source of primary MPs. Thermoplastics are mainly formed by polyethylene (PE),  
194 polypropylene (PP) or polyvinylchloride (PVC), while thermoset plastics are formed, among  
195 others, by Polyester (PES), polyurethane (PUR), Silicone and Polyamide.

196 Sources of plastics can be classified in terms of the life expectancy of the produced  
197 plastics before disposal. Here we classify plastic sources into those with a short-term (single  
198 use or very limited number of times with a useful lifespan up to 1 year), mid-term (up to 10  
199 years), or long-term (more than 10 years) use expectancy.

### 200 *Plastics with short-term use expectancy*

201 Single-use items are mainly formed by packaging material, which is the biggest  
202 plastic sector worldwide (almost 36% in 2015; Fig. 1) and accounts for almost 50% of the  
203 generated plastic waste (Geyer et al. 2017). The vast majority of packaging plastics are PE,  
204 PP and polyethylene terephthalate (PET) (Geyer et al. 2017). Except for refillable PET  
205 bottles used in some countries, packaging is single-use with a life span of less than six  
206 months. Most foods are wrapped in plastic and single-use plastic bags have been widely  
207 used all over the world due to their convenience, availability and low price. Plastic bags are  
208 known to cause severe environmental and health problems, especially in countries without  
209 proper waste management (Adane and Muleta 2009). Thus, many countries have put bans  
210 or levies in force to reduce their use or to encourage voluntary reductions (Xanthos and  
211 Walker 2017). Many African countries, for instance, have banned single-use plastic bags,  
212 while the EU Directive 2015/720 encourages member states to reduce the number of

213 “lightweight” carrier bags by 2025. Those bans and restrictions have already reduced the  
214 plastic bag use drastically in some countries (e.g. Ireland, England, Italy). Moreover, other  
215 single-use items like cutlery, plates, cups and straws are planned to be banned in Europe by  
216 2021 (EC 2019).

217 Another important sector using single-use plastic is agriculture. Plastic films are used  
218 for plastic mulching, for the construction of greenhouses and tunnels, or to wrap silage to  
219 store animal fodder. The global plastic consumption in agricultural production is estimated to  
220 be about 2.5 million tons per year (Hussain and Hanid 2003). A variety of different plastic  
221 types are used in agriculture, including PE, PP, Ethylene-Vinyl Acetate Copolymer (EVA),  
222 PVC and poly-methyl-methacrylate (PMMA) (Scarascia-Mugnozza et al. 2012).

223 MPs added to consumer products (e.g. as a component of personal care, cosmetic  
224 and cleaning products) are especially manufactured to be used once and then washed down  
225 the drain. They are often referred to as microbeads, even though they are mostly irregular in  
226 shape in order to obtain an abrasive effect (Fendall and Sewell 2009; Napper et al. 2015;  
227 Kalčíková et al. 2017). The majority of microbeads in facial and body scrubs are made of PE,  
228 with average concentrations of 4.82 g/100 mL body scrub and 0.74g/100 mL facial scrub  
229 (Kalicova et al. 2017, Gouin et al. 2015). Other plastic polymers used in cosmetic products  
230 include polylactic acid, PET, polyethylene isoterephthalate, nylon-12, nylon-6, PMMA,  
231 polytetrafluoroethylene, and PUR (Leslie 2014; Rochman et al. 2015). Additionally,  
232 microbeads are used in industry as abrasives/scrubbers and sand-blasting media as well as  
233 in anti-slip, anti-blocking applications and for medical applications. It has been calculated that  
234 more than 4000 tons of PE microbeads were used in cosmetic products all over the EU  
235 (including Norway and Switzerland) in 2012 (Gouin et al. 2015), and the US is emitting 263  
236 tons of PE microbeads per year (2.4 mg per person per day; Gouin et al. 2011). A ban of  
237 microplastics intentionally added to products (i.e., microbeads) has been proposed in the EU,  
238 while the US Microbead free waters act of 2015 (US Congress 2015) prohibits the  
239 manufacturing, packaging, and distribution of rinse-off cosmetics containing plastic  
240 microbeads already. This only applies to rinse-off products, while MPs are still permitted as a  
241 component in ‘leave on’ products (e.g. lotions, sunscreens, make-ups and deodorants).

#### 242 *Plastics with mid-term use expectancy*

243 Plastics with a mid-term lifespan are mainly found in the sectors of electronic,  
244 household, tyres and textiles. The production of electrical and electronical products counts to  
245 the fasted growing manufacturing and waste generation sectors (Geyer et al. 2017; Kumar et  
246 al. 2017) and as many textiles are made, entirely or to a certain extent, of synthetic plastic

247 fibres (e.g. PA, PES, Acrylic) also production rates of synthetic plastic fibres have increased  
248 over the last decade. Nowadays, two-thirds of the total fibre production is synthetic plastic  
249 fibres and worldwide 59 tons of plastic textiles were produced in 2015 (Geyer et al. 2017;  
250 Gasperi et al. 2018).

251 Synthetic polymers with rubber-like characteristics are the principal component of  
252 vehicle tyres. They are composed of a mixture of natural and synthetic rubbers (styrene-  
253 butadiene rubber). While driving, tyre and road wear particles are formed which contain  
254 styrene-butadiene rubber in a mix with natural rubber, pavement parts and many other  
255 additives (Unice et al. 2013; Sundt et al. 2014). While tires contain almost 50% of polymers,  
256 tyre wear particles, which are a mix of pavement part and polymers contain only 16-23% of  
257 polymers (Kreider et al. 2010).

### 258 *Plastics with long-term use expectancy*

259 Plastics designed for long-term use belong to the following categories: parts of  
260 transportation (i.e. vehicle, plane and trains parts), building and construction, industrial  
261 machinery, also consumer and institutional products. While plastics for the building and  
262 construction sector account for the second highest plastic consumption, only a small portion  
263 enters the waste stream directly (Fig. 1; Geyer et al. 2017). As these categories do not  
264 belong to the items that are usually littered, they are not expected to contribute significantly  
265 to the plastic load in the environment. However, their breakdown rate into MPs and NPs (due  
266 to exposure to light and weathering), also during their useful lifetime, is not clear.

## 267 **3. Pathways of plastic to the environment**

268 Hereafter plastic waste will refer to all plastic material that is discarded, while litter will  
269 include only those items that are not properly discarded. Packaging material is accounting for  
270 almost 50% of the generated plastic waste, followed by textiles (almost 14%; Geyer et al.  
271 2017). Most plastic waste is generated in Asia, while America, Japan and the European  
272 Union are the world's largest producers of plastic packaging waste per capita.

### 273 **3.1. Collected solid waste**

274 Collected plastic waste is either landfilled, incinerated or recycled. In Europe 27.3% are  
275 landfilled, 30.1% are recycled and 41.6% are incinerated for energy recovery (PlasticsEurope  
276 2018). The percentage of collected plastic waste varies strongly between different countries,  
277 depending on the applied waste management plans and policies. While worldwide the plastic

278 recycling rate is still low, it has increased by almost 79% within the last 10 years in the EU,  
279 including Norway and Switzerland (PlasticsEurope 2018).

280 Large scale industrial plastic production began in the 50s, but plastic recycling was not  
281 established until the 80s. It is estimated that only 9% of the total produced plastic waste up to  
282 2015 has been recycled (Geyer et al. 2017). From this again only a small portion is submitted  
283 to primary recycling in which the recycled plastic is used to replace all or a least a proportion  
284 of the virgin polymer resins (Hopewell et al. 2009). While high-income countries have sorting  
285 and processing facilities, in low income countries plastic recycling is not well established.  
286 Moreover, certain types of plastic are difficult to recycle. For example, thermoset plastics,  
287 including textiles, are usually not recycled.

288 Plastic that is not recycled but still collected is landfilled or incinerated. In eight EU  
289 countries, Norway and Switzerland, a landfill ban for plastic is in force, leading to a very small  
290 percentage of plastic being used for landfill applications (PlasticsEurope 2018). On average  
291 27.3% of the generated plastic waste is landfilled in Europe. In contrast, in low-income  
292 countries, waste is mainly stored in open, poorly managed dumps, from where plastic can be  
293 transported by wind force. In middle-income countries, some controlled landfills are in place,  
294 but open dumping is still common practice. The advantages of combustion of plastic waste  
295 are that it can be used for energy recovery and the incinerated plastic cannot enter the  
296 environment anymore. At the same time, incineration results in the generation of air  
297 pollutants (Verma et al. 2016).

### 298 **3.2. Wastewater**

299 Both MPs as well as MaP enter wastewater either directly if products containing plastic are  
300 flushed down the drain (e.g. fibres detached during laundry of textiles, microbeads in consumer  
301 products, cotton buds or sanitary products), or in combined sewer systems from street dust and  
302 litter. MaP escape wastewater treatment only on rare occasions and mainly enter the  
303 environment with untreated wastewater due to combined sewer overflows e.g. after heavy  
304 rainfall events or snowmelts (Williams and Simmons 1999), or if untreated wastewater enters  
305 the environment because WWTPs are not in place. Although high-income countries treat on  
306 average 70% of the wastewater, yet globally only 20% of the generated wastewater is  
307 treated (Sato et al. 2013). For MPs, the situation is different, due to their small size, they can  
308 escape the treatment and are also released with treated effluents (Ziajahromi et al. 2016).  
309 This pathway for MPs has been increasingly investigated. To date, 21 studies have  
310 measured MPs in wastewater (Tab. S1), from which two do not exclusively assessed MPs  
311 but included other litter items in the micro range (microliter; Michielssen et al. 2016; Talvitie



312 et al. 2017b). Such studies were mainly carried out in (northern and western) Europe (13  
313 studies), followed by north-America (5 studies).

314 The number of MPs in raw wastewater varies greatly between WWTPs, from a few  
315 MPs/L to exceptional maximum values of more than 10,000 MP/L (Fig. 2; Tab. S1).  
316 Especially high concentrations have been observed in raw wastewaters in Denmark  
317 (Vollertsen and Hansen 2017; Simon et al. 2018). The Danish studies assessed MPs in the  
318 smaller size range (i.e., between 10 or 20 and 500  $\mu\text{m}$ ), while other studies assessing MPs  
319 down to 20  $\mu\text{m}$  found much lower MP concentrations (Talvitie et al. 2015; Leslie et al. 2017).

320 WWTPs have in general a large retention potential for MPs, often higher than 95% (Tab.  
321 S1). However, in treated wastewater the number of MPs varies greatly too, from less than 1  
322 MP/L (Browne et al. 2011; Carr et al. 2016; Murphy et al. 2016; Ziajahromi et al. 2017) to  
323 several hundred (Simon et al. 2018), and up to several thousand MP/L (Vollertsen and  
324 Hansen 2017; Fig. 2). Larger MPs are usually better retained during the treatment, so the  
325 most frequently observed MPs in treated wastewater are smaller than 300  $\mu\text{m}$  (Dris et al.  
326 2015; Mintenig et al. 2017; Gündoğdu et al. 2018; Magni et al. 2019; Talvitie et al. 2017a;  
327 Lee and Kim 2018; Wolff et al. 2018; Liu et al. 2019). For example, Magni et al. (2019) found  
328 that 94% of the MPs between 5–1 mm were retained by an Italian WWTP, while only 65% of  
329 the MPs between 0.1–0.01 mm were retained (Magni et al. 2019). Moreover, the number of  
330 MPs seems to be increasing with decreasing particle size. Wolff et al. (2019) reported the  
331 results of small-size MPs measured in treated wastewater and indicated that the 44% of  
332 measured MPs are between 10 and 30  $\mu\text{m}$ , while 51% are between 30 and 100  $\mu\text{m}$ .  
333 Furthermore, current research indicates that the amount of MPs retained by WWTPs is not  
334 only influenced by the size, but also by the particle shape. Usually, fibres are better retained  
335 in WWTPs as compared to microbeads or other irregular particles (Magnusson and Norén  
336 2014; Talvitie et al. 2017b; Gündoğdu et al. 2018). Fibres and fragments are the most  
337 frequently occurring MP types in WWTP effluents (Tab. S1). Regarding polymer composition,  
338 PE particles or PES fibres are the most common plastic types (Tab. S1). Although a huge  
339 amount of tyre debris is suspected to enter WWTPs (Kole et al. 2017), they have not been  
340 frequently reported in treated effluents (Tab. S1). Only Dyachenko et al. (2017) and Lee and  
341 Kim (2018) have reported the presence of black particles possibly being tyre fragments.

342 Interestingly, concentrations of MPs in wastewaters show some seasonal and diurnal  
343 variations related to water consumption rates and human activity (Mintenig et al. 2017;  
344 Talvitie et al. 2017b; Lares et al. 2018). For instance, Talvitie et al. (2017b) reported that  
345 night time concentrations were slightly lower (average concentrations 476.7 and 0.8  
346 microlitter/L in influent and effluent respectively) compared to day time concentrations (584

347 and 1.7 microliter /L in influent and effluent, respectively). Therefore, MPs occurrence seems  
348 to be highly variable and depending on a variety of different environmental (weather, season)  
349 and behavioural variables but also methodological procedures (i.e. sampling method,  
350 including mesh sizes and sample volume), extraction method, and determination method.  
351 Despite the high retention of MPs by WWTPs, considering the large volumes treated daily, it  
352 is considered that more than one million particles can enter the aquatic environment via this  
353 pathway per WWTP (Ziajahromi et al. 2017; Gündoğdu et al. 2018), which constitutes one of  
354 the main sources of MPs into the environment.

### 355 **3.3. Sludge and other agricultural amendments**

356

357 The majority of MPs is already retained by WWTPs during pre- and primary treatment  
358 (mechanical treatment and sludge settling processes) and therefore concentrated in the  
359 grease or sludge phase (Murphy et al. 2016; Leslie et al. 2017; Talvitie et al. 2017b). While  
360 solids intercepted by grids and grease removal steps are disposed on landfills, sludge is  
361 often reused as fertilizers in agriculture. The amount trapped in the sludge roughly  
362 constitutes 50-90% of the MPs present in raw wastewater (Tab. S2; Magnusson and Norén  
363 2014; Carr et al. 2016; Lee and Kim 2018). MP concentrations measured in sludge range  
364 between 650 MPs/Kg dw to more than 240,300 MPs/Kg dw (Fig. 3, Tab. S2). Murphy et al.  
365 (2016) found significant bigger sized MPs in the sludge phase compared to MPs in treated  
366 wastewater, confirming the differential retention potential of WWTPs regarding MPs size.  
367 Furthermore, the sludge treatment process (thickening, digestion, drying, stabilization,  
368 dewatering) may have an effect on the MP size (Mahon et al. 2017). Similar to wastewater,  
369 sludge samples usually show high numbers of fibres, followed by fragments (Tab. S2), and  
370 the main detected polymer is usually PES (particularly when there are many fibres present),  
371 followed by PE and PP.

372  
373 Plastics can end up in compost used as agricultural amendment due to wrong recycling  
374 or separation of waste, e.g. if plastic food packaging is disposed in the organic waste  
375 (Mercier et al. 2017; Weithmann et al. 2018). Weithmann et al. (2018) reported that organic  
376 fertilizers may contain up to 895 MPs/kg, and Fuller and Gautam (2016) found on average  
377 23,000 mg MP/kg in composted waste materials.

378  
379

## 380 **4. Occurrence and fluxes of plastics in environmental compartments**

381

382  
383  
384

### **4.1. Air**

385  
386           Studies assessing the occurrence of airborne plastic particles have identified mainly  
387 fibres (Dris et al. 2015; Abbasi et al. 2019). Atmospheric fallout of fibres in the area of Paris  
388 (France) showed a high variability, with values ranging between 2 and 355 fibres/m<sup>2</sup>/day;  
389 however, half of those were natural (50%; cotton or wool), and only 17% were purely  
390 synthetic (mainly PET; Dris et al. 2016). Based on these samples, the same authors  
391 estimated that the fibre deposition rate in highly populated urban environments can roughly  
392 range between 1.2 and 4 kg/km<sup>2</sup>/year, and concluded that atmospheric fallout might  
393 constitute a relevant pathway of MPs. The limited data on atmospheric MPs deposition rates  
394 makes it is difficult to draw conclusions on the relevance of this pathway for the  
395 environmental distribution of MPs. In the study by Dris et al. (2016) suburban fallout was  
396 found to be only about 50% of that observed in urban areas (53 particles/m<sup>2</sup>/day compared  
397 to 110 particles/m<sup>2</sup>/day), and thus it may be assumed that fibre fallout is even lower in natural  
398 and agricultural environments.

399  
400           In addition to fibres, MPs in street dust are also likely to become airborne (Dall'Osto  
401 et al. 2014; Gasperi et al. 2018). According to Kole et al. (2017), 12% of the generated tyre  
402 dust (1040 tonnes) in the Netherlands ends up in the air. The particles are generated by the  
403 interaction of tires with the road while driving and are generally found along roadside areas  
404 (Kreider et al. 2010). Wind and rainfall might influence the atmospheric transport and fallout  
405 of MPs, while deposited fibres and street dust in urban environments may be transported via  
406 water runoff into sewer systems or directly to terrestrial or aquatic ecosystems, however  
407 studies properly describing such processes are lacking.

408  
409

#### 410           **4.2. Soil**

411  
412           It has been suggested that agricultural soils could constitute larger MP sinks than  
413 marine ecosystems (Hurley and Nizzetto 2018). However, research on the quantification of  
414 plastics in soils (for both MaPs and MPs) is still very limited and mostly contracted to the last  
415 four years. We identified twelve studies reporting plastics in soil, from which three considered  
416 only a limited number of plastic types (Tab. S3). The available studies provide first  
417 indications of the scale of the pollution and suggest the ubiquitous presence of MPs in  
418 terrestrial ecosystems, also beyond agricultural areas. Most studies report plastic quantities  
419 in terms of particles, while some others provide concentrations based on mass  
420 measurements, which hampers to some extent direct comparisons among them. The highest  
421 MP concentration based on mass has been measured in soils from an industrial area in  
422 Australia, which was historically used to produce chlorinated plastic, containing 6700 mg

423 MP/kg dw (Fuller and Gautam 2016). The highest concentration based on the number of MP  
424 particles was provided by Vollertsen and Hansen (2017), who described Danish agricultural  
425 soils containing about 145 000 MPs/kg, in the size range of 20 to 500  $\mu\text{m}$  which was based  
426 on weight however only 12 mg/kg. Also Chinese farmland soils were found to contain a high  
427 MP content, ranging between 70 and 18,760 MPs/kg dw (Fig. 4; Liu et al. 2018; Zhang and  
428 Liu 2018; Zhang et al. 2018). In contrast farmlands in Germany showed a much lower MP  
429 occurrence (0.34 MPs/ kg dw; Piehl et al. 2018). This might be partly related to differences in  
430 the considered MP sizes during the study and due to differences in agricultural practices.  
431 While Piehl et al. (2018) assessed MPs of a size between 1 and 5 mm, the study by  
432 Vollertsen and Hansen (2017) considered MPs between 20  $\mu\text{m}$  and 500  $\mu\text{m}$ . However, the  
433 different ranges in concentrations seem mostly attributed to the presence of different input  
434 sources.

435

436 The application of sewage sludge as agricultural fertilizer (biosolids) is considered to  
437 be a major source of MPs to soils. Nizzetto et al. (2016) estimated that between 63,000-  
438 430,000 and 44,000-300,000 tons of MPs could be yearly added to agricultural land in  
439 Europe and north America, respectively. Corradini et al. (2019a) found that increasing  
440 number of sludge applications were positively correlated to increasing MP concentrations in  
441 soils. Zubris and Richards (2005), report up to 1,210 fibres/kg in soils five years after sewage  
442 sludge application and detected fibres still 15 years after application, which is another  
443 indication for MPs accumulation in soil due to sludge application. On the other hand, almost  
444 twice the concentration of MPs was found in Danish fields not treated with sludge compared  
445 to treated fields (Vollertsen and Hansen 2017). Additional studies investigating the presence  
446 of MPs in soil after application of wastewater sludge are fundamental to better estimate the  
447 importance of this pathway.

448

449 Irrigation with reclaimed wastewater and the usage of plastic material in agriculture  
450 constitute additional sources of plastics in soil ecosystems. Based on studies from China, the  
451 latter one seems to be one of the most important plastic sources for elevated MPs  
452 concentrations in soil in addition to sewage sludge application (Zhang and Liu 2018; Zhang  
453 et al. 2018). In contrast to those concentration hot spots, agricultural areas in Germany  
454 without plastic mulching or use of sewage sludge as fertilizer the MP concentration seems  
455 much lower (i.e. on average 0.34 MP/kg dw soil; Piehl et al. 2018). As the frequency of the  
456 observed MaP polymer types was reflected by the types of MPs, MP particles in this study  
457 most likely come from degradation of (littered) MaP (Piehl et al. 2018). The breakdown of  
458 MaP into MPs in terrestrial ecosystems may be dependent on their whereabouts in the soil  
459 and on soil cultivation. Williams and Simmons (1996) assessed Low density PE degradation

460 over a period of four months in different environments (river beach, in trees at the river bench  
461 and buried by soil). They found that MaPs on the soil surface degrade faster as compared to  
462 buried plastics, and assumed light to be the main influencing driver (although rainfall and  
463 other weathering processes may have affected degradation).

464  
465 Littering, drift from landfills or spills from industry can also become important sources  
466 of plastics into soils. As described above, deposition from of MPs from the air can  
467 additionally add MPs to soils, this seems however more relevant close to urban areas and  
468 streets with heavy traffic. Finally, during flood events plastics from the aquatic environment  
469 can be deposited in the shores of rivers (Scheurer and Bigalke 2018). Therefore, based on  
470 the data that is available up to now, the main inputs of MPs into soil seem to come from  
471 agricultural practices (sewage sludge, plastic mulching) and the fragmentation of plastic litter.

472  
473 The most common polymer types reported in soils are PE and PP (Tab. S3). MaP  
474 reported in terrestrial systems are PE films and bottles (Ramos et al. 2015; Huerta Lwanga et  
475 al. 2017b; Piehl et al. 2018). In a more remote place (desert in southern Arizona) plastic that  
476 is more mobile due to transportation by wind like plastic bags and balloons have been  
477 reported (Zylstra 2013).

478  
479 The fate of MPs within the soil is not completely clear yet. MPs in soils may be  
480 transported along with water runoff and soil erosion into adjacent streams and rivers. So far,  
481 there is no knowledge on the importance of this pathway as it has not been experimentally  
482 proven. Translocation into deeper soil layers can occur through soil cultivation (Hurley and  
483 Nizzetto 2018) or transport by soil organisms. Earthworms and collembola have been shown  
484 to ingest and transport MPs from the soil surface into deeper soil layers (Huerta Lwanga et  
485 al. 2017a; Maaß et al. 2017; Rillig et al. 2017). Also other animals e.g. birds or domestic  
486 animals, which have been shown to take up MPs (Zhao et al. 2016; Huerta Lwanga et al.  
487 2017b) can transport MPs over longer distances. To date, it is yet unclear whether low sized  
488 MPs can be transported through soil pores into ground water, but low concentrations of MPs  
489 (0 to 7 MPs/m<sup>3</sup>) have been reported in raw drinking waters from groundwater wells (Mintenig  
490 et al. 2019). Uptake of plastics by plants is another potential source of mobilization of plastics  
491 from soil ecosystems, particularly for NPs, however no studies have investigated this using  
492 whole plants (Ng et al. 2018). The only study available in this respect is the one provided by  
493 Bandmann et al. (2012), who demonstrated uptake of 20 and 40 nm PS beads by tobacco  
494 BY-2 cells in cell culture via endocytosis, while 100 nm beads were excluded.

495  
496  
497

### **4.3 Surface waters**

498 Plastic pollution along rivers has been already observed and assessed in the 1990s  
499 (Williams and Simmons 1996, 1999). Nevertheless, few studies have reported plastic  
500 pollution in freshwaters until the whole environmental movement was initiated few years ago.  
501 Some studies assessing litter in rivers have not exclusively focused on plastic, but also  
502 included other litter items like glass, paper and wood. Those studies show that about 80% of  
503 the litter items are plastics, but do not provide concentrations or mass estimates (Crosti et al.  
504 2018; González-Fernández et al. 2018; Castro-Jiménez et al. 2019).

505 Studies focusing on providing concentrations of MaPs in the environment are very  
506 limited (Tab S4). MaPs concentrations have been reported for example for the Los Angeles  
507 river, in California (819 MaPs/m<sup>3</sup>; Moore et al. 2011), the Yangtze river in China (8.74x10<sup>3</sup>  
508 MaPs/km<sup>2</sup>; Xiong et al. 2019), and in Lakes (1,800 MaPs/km<sup>2</sup>) and Rivers (0.012 MaPs/m<sup>3</sup>) in  
509 Switzerland (Faure et al. 2015). It has been estimated that in the river Seine in France,  
510 28,000 kg of floating plastic are trapped annually by floating debris retention booms (Gasperi  
511 et al. 2014) and floating MaP in the Saigon river in Vietnam were estimated to range between  
512 7,500 – 13,700 tons per year (van Emmerik et al. 2018). As only buoyant plastics were  
513 considered in those studies, the total loads may be underestimated as plastic is also  
514 transported by sub-surface transport (Morritt et al. 2014). The most common MaPs reported  
515 in freshwater environments are plastic bottles, food packaging items, plastic bags and  
516 sewage-related plastic like handles from buds of cotton wool and sanitary towels (Tab. S4).  
517 Regarding polymer composition, PP and PE are the plastic types that were omnipresent, and  
518 to a lesser extent PS and PET have been reported (Table S4).

519  
520 MPs in water have been reported in different units (i.e. particles per water volume, or  
521 particles per area). To be able to compare the results of the different studies, we choose 37  
522 studies which either reported the number of MPs per water volume or gave sufficient  
523 information to transform the reported unit. Like in other environmental compartments the  
524 concentrations varied greatly among studies (Fig. 5, Tab S5). Most studies in Europe found  
525 average concentrations of less than 1 to less than 100 MP/m<sup>3</sup>, while the highest average  
526 concentration of 100,000 MPs/ m<sup>3</sup> (with a maximum concentration of 187,000 MPs/ m<sup>3</sup>) was  
527 measured in the Amsterdam Canals (Leslie et al. 2017). Furthermore, Lui et al. (2019a)  
528 reported up to 22,849 MPs/m<sup>3</sup> (average: 1,409 MPs/m<sup>3</sup>) in storm water ponds receiving  
529 urban runoff in Denmark. The highest peak concentration from all studies was found in the  
530 Snake River in North America and was as high as 5,405,000 MPs/m<sup>3</sup> (average: 91 MPs/m<sup>2</sup>)  
531 (Kapp and Yeatman 2018). The second highest peak concentration was reported by Lahens  
532 et al. (2018), and corresponds to 519,223 MPs/m<sup>3</sup> (minimum 17,210 MPs/m<sup>3</sup>) monitored in  
533 the Saigon River (Vietnam). Overall, reported concentrations of MPs appear to be higher in  
534 Asia, as compared to Europe and North America (Fig. 5). However, most of the studies

535 carried out in Asia were performed in China and focused on assessing lower size classes  
536 that those studied in Europe. The only two studies conducted in Europe that considered a  
537 very low size (MPs below 20  $\mu\text{m}$ ), were the ones by Leslie et al. (2017a) and Lui et al.  
538 (2019), who observed by far the highest concentrations. Current research shows that smaller  
539 particles ( $<0.5$  mm) are usually the most frequent ones (e.g. Leslie et al. 2017; Yan et al.  
540 2019). Therefore, the higher concentrations found in Asia may be not exclusively related to a  
541 higher pollution but also to the sampling methods used.

542

543 Studies assessing the concentration of MPs using different net sizes at the same  
544 sampling sites found substantial differences in the number of particles intercepted by  
545 plankton nets vs trawling nets (Dris et al. 2015; Xiong et al. 2019). Kapp and Yeatman (2018)  
546 used both sampling methods to assess the occurrence of particles larger than 100  $\mu\text{m}$  and  
547 found that on average there were higher concentrations in grab samples (glass containers  
548 were filled with water from the surface) as compared to net samples (Tab. S5). Also, other  
549 differences in study design such as sample volume, sample depth, or sample location in the  
550 river could influence the measured MPs concentration. For example, Vermaire et al. (2017)  
551 found higher concentrations in grab samples close to the river shore, which were  
552 subsequently filtered through a 100  $\mu\text{m}$  net compared to open water samples taken using a  
553 100  $\mu\text{m}$  manta trawl.

554

555

556 Fig. 2 Overview on most common sampling methods used for freshwater MPs sampling

557

558

559 Although MPs have been found in remote locations and rural areas, there is evidence  
560 that MPs concentration increases with proximity to cities (Wang et al. 2017b; Di and Wang  
561 2018; Tibbetts et al. 2018). A modelling study identified the Yangtze River catchment as the  
562 catchment transporting the highest plastic loads into the ocean (Schmidt et al. 2017a). The  
563 four case studies looking at MPs concentrations in the Yangtze river found highly variable  
564 concentrations, but were also amongst the highest observed (Zhang et al. 2015; Wang et al.  
565 2017b; Di and Wang 2018; Xiong et al. 2019). However, concentrations in the same order of  
566 magnitude were also monitored in other rivers in China such as the Pearl river, which was  
567 also ranked under the top ten catchments transporting plastic into the ocean (Schmidt et al.  
568 2017a).

569

570 Not only spatial hot spots but temporal hot spots based on weather condition may  
571 exist in freshwater ecosystems. Storms and rainfall can increase plastic concentration in

572 waters from both lateral (land-based) and sewage effluent discharge points (Fischer et al.  
573 2016), and MPs that had been deposited on river beds can re-enter the water phase again  
574 after flood events (Hurley et al. 2018a).

575

576 Fragments and fibres formed by PE and PP are the most frequently observed  
577 particles across all studies evaluating MP pollution in freshwater ecosystems; whereas  
578 pellets or beads are only rarely reported as the main occurring plastic types (Tab. S5). The  
579 latter are mainly found in studies along the rivers Rhine and Danube, in the proximity to  
580 plastic processing plants and are thus assumed to be pre-production pellets (Lechner et al.  
581 2014; Lechner and Ramler 2015; Mani et al. 2016). The prevalence of secondary MPs  
582 (fragments and fibres) suggests wastewater and runoff as sources for plastic pollution in  
583 freshwater ecosystems (Tab.S5). Several studies confirmed that by demonstrating that MP  
584 concentrations are higher downstream of WWTP as compared to sampling sites in upstream  
585 areas (McCormick et al. 2014; Estahbanati and Fahrenfeld 2016; Vermaire et al. 2017; Kay  
586 et al. 2018). For example, in the Ottawa River (Canada), 0.71 particles/m<sup>3</sup> were found  
587 upstream of a WWTPs compared to 1.99 MPs/m<sup>3</sup> downstream. In the Raritan River and the  
588 North Shore Channel (USA) 24 MPs/m<sup>3</sup> and 1.94 MPs/m<sup>3</sup> were found upstream the WWTP,  
589 and 71.7 particles and 17.93 MPs/m<sup>3</sup> were detected downstream, respectively (McCormick et  
590 al. 2014; Estahbanati and Fahrenfeld 2016; Vermaire et al. 2017). As mentioned above, the  
591 majority of MPs in wastewater is smaller than 300 µm, thus it may be presumed that larger  
592 MPs enter via different pathway like surface runoff, or steam from the breakdown of MaPs  
593 directly in the aquatic environment. However, with untreated wastewater, for instance during  
594 sewage overflows, MaPs can enter river ecosystems. For example, Morritt et al. (2014)  
595 identified pollution hotspots in the vicinity of WWTPs that were mainly constituted of sanitary  
596 products. MPs hotspots were also detected in areas with low population density but high  
597 agricultural use, pointing also to agricultural runoff as an important source (Kapp and  
598 Yeatman 2018).\_Finally, poor waste management likely increases plastic input into aquatic  
599 ecosystems (Lahens et al. 2018), where they can break down into smaller particles. Xiong et  
600 al. (2019), for example, found that the abundance of microplastic is positively related to the  
601 presence of MaPs.

602

603

#### 604 **4.4. Sediments**

605 Similar to MaP in surface waters also MaPs in sediments are only rarely assessed  
606 and the way MaP occurrence is reported is highly variable and difficult to compare (Tab. S6).  
607 MaPs along river banks have been observed while assessing buoyant litter in general  
608 (Williams and Simmons 1999; Rech et al. 2014), and river beach sediments in Switzerland



609 contained on average 90 MaPs/m<sup>2</sup> (Faure et al. 2015). Across different lake shores, MaPs  
610 concentrations have been shown to vary notably (Imhof et al. 2013; Fischer et al. 2016).  
611 While high MaPs concentrations have been observed at the south shore of Lake Garda  
612 (Italy; with an average concentration of 483 MaP/m<sup>2</sup>), the occurrence at the north shore was  
613 significantly lower (i.e. 0-8.3 MaP/m<sup>2</sup>; Imhof et al. 2013). Food packaging is among the most  
614 frequently observed MaPs but also bottles, bags and ropes are described by several studies.  
615 Regarding the polymer composition, PE and PP as well as Styrofoam (PS) are reported (Tab  
616 S6).

617  
618 As for MaPs and the other compartments, the concentration of MPs in freshwater  
619 sediments has not been reported in consistent units across all studies. Therefore, we  
620 focused on studies that have reported the concentration in MPs/kg sediment. However,  
621 studies reporting MPs per sediment area, which gave sufficient information to estimate the  
622 concentration in MPs/kg, were also included. Therefore, from the 33 studies that were found  
623 during the literature search, 30 were chosen for comparisons (Fig.6, Tab. S7). The highest  
624 sediment concentration of 2,071 MPs/kg dw has been found in the urban canals of  
625 Amsterdam, where also the highest water concentrations were observed (Leslie et al. 2017).  
626 MP concentrations in river bed sediments seem, in general, higher than in river beach and  
627 shore sediments (Fig. 6; Tab S7). Most studies on MPs in river bed sediments report  
628 concentrations between 100 MP/kg and a few thousands. Studies from Asia were exclusively  
629 carried out in China, and reported similar concentration ranges as those described in Europe.  
630 Interestingly, the study on the Yangtze River (China), which has been estimated to be the  
631 highest contributor of plastic to the sea (Schmidt et al. 2017a) and amongst the highest MPs  
632 concentrations reported in water (Fig. 5, Tab. S5), had a comparably low sediment  
633 concentration 7-66 MP/kg. The only study carried out in Africa assessing the concentration of  
634 MPs in river sediment reports notable differences between concentrations in summer (1-  
635 14.61 MP/kg dw) and winter (13.3 - 563.8 MP/kg dw; Nel et al. 2018), which were related to a  
636 reduced flow condition in winter. Subsequently, the hydrological variation shown by many  
637 rivers seems to be one of the main factors contributing to MPs deposition and re-mobilization  
638 from river beds. This was also demonstrated by Hurley et al. (2018a), who report that about  
639 70% of the MPs in the sediments of the upper Mersey and Irwell catchments (UK) were  
640 exported after a flooding event. Several studies show that, after transportation with the river  
641 flows, MPs tend to (re-)deposit in low energy environments, such as meanders, deltas,  
642 dams, harbours and coastal lagoons (Claessens et al. 2011; Vianello et al. 2013; Shruti et al.  
643 2019). The deposition of low-density polymers in sediment environments is also related to a  
644 density increase by biofouling (e.g. Ye and Andrady 1991; Andrady 2011; Zettler et al. 2013;  
645 McCormick et al. 2014).

646

647 For lakes, mainly beach and shore sediment concentrations have been reported. In  
648 Europe average concentrations for beach and shore sediments ranged between 0.94 and 44  
649 MP/kg, while beach and shore sediments from Lake Ontario (Canada) contained much  
650 higher concentrations (20-27,830 MPs/kg; Fig. 6, Tab. S7). Several studies have noted that  
651 plastic concentrations differ strongly between different areas of the same lake (Zbyszewski  
652 and Corcoran 2011; Imhof et al. 2013; Zbyszewski et al. 2014; Zhang et al. 2016),  
653 suggesting that accumulation is patchy and form contamination hotspots influenced by  
654 winds, waves and/or beach morphology (Imhof et al. 2016, 2018). Similar observations were  
655 made at Lake Huron (Canada), in which 94% of all monitored pellets were found to  
656 accumulate in one single beach (Zbyszewski and Corcoran 2011). In the Taihu Lake (China),  
657 MPs concentrations ranged from 11 to 235 MP/kg in different bed areas, and the average  
658 MPs abundance in sediments in the northwest area was approximately six times higher than  
659 that of the southeast area (Su et al. 2016).

660 Fibres followed by fragments were usually the most common particle types monitored (Tab.  
661 S7). Spheres/beads or pellets were, in rare occasions, reported to be dominant, and mostly  
662 in the vicinity to plastic industries (Zbyszewski and Corcoran 2011; Zbyszewski et al. 2014;  
663 Corcoran et al. 2015; Hurley et al. 2018a; Peng et al. 2018). Based on polymer type, PE and  
664 PP were the most common, despite their buoyant properties, as well as PS (Tab. S7).

665

666

667

#### 4.5. *Marine*

668

669 Rivers are estimated to be the main pathways for plastics entering the oceans.  
670 Estimations on the amount of plastic waste entering the ocean through this pathway range  
671 between 0.41 and  $4 \times 10^6$  tons per year (Lebreton et al. 2017; Schmidt et al. 2017b). From  
672 the top ten river catchment that transport 88-95% of the global plastic load into the oceans,  
673 eight are located in Asia (Schmidt et al. 2017b). Oceans have been assumed to be the final  
674 sink for MaPs and MPs. As this review is focused on terrestrial and freshwater ecosystems,  
675 this compartment will not be discussed in detail. A number of articles and reviews have been  
676 published on the topic within the last few years which describe plastic occurrence in the  
677 oceans and its effects on marine life (see Barboza and Gimenez 2015; Jambeck et al. 2015;  
678 Auta et al. 2017).

679

680

## 681 **5. Discussion**

682

683 We full agree with the statement provided by the SAPEA (2018) report: “*The number*  
684 *of papers is growing exponentially in this field, but knowledge is not growing at the same rate*  
685 *— there is some redundancy and marginality in the papers*”. Furthermore, many papers on  
686 plastic pollution do not assess and describe important plastic sources and flows. This review  
687 paper made an attempt to describe the available information regarding global environmental  
688 loads and the plastic life cycle, and to show that further research studies are needed to fully  
689 understand specific plastic sources and pathways. This section describes the areas that  
690 need further research commitment and development to improve exposure assessments and  
691 to evaluate the long-term risks of plastics to terrestrial and freshwater ecosystems.

692

### 693 **5.1. The need for standardization of sampling and analysis**

694

695 As indicated in several parts of this review, the sampling methods reported in the  
696 literature are extremely variable and, in many cases, difficult to compare. In water, the most  
697 commonly used method for sampling is the so-called manta trawl, a device similar to a large  
698 plankton net with a mesh size usually larger than 300  $\mu\text{m}$ . The same device is generally used  
699 in rivers, lakes and in marine monitoring studies. Using a manta trawl allows to sample a thin  
700 layer of surface water and, therefore, the results are generally reported as MPs (number or  
701 weight) per surface area ( $\text{m}^2$  or  $\text{km}^2$ ). When grab water samples were taken or water was  
702 pumped through a net or a sieve, the results are expressed as MPs per volume unit (e.g. L or  
703  $\text{m}^3$ ) and different size fractions are considered, sometimes down to 20  $\mu\text{m}$ . The results from  
704 studies considering the two aforementioned sampling methods are hardly comparable. Data  
705 for surface units may be converted into data for unit volume, by calculating the mouth surface  
706 area of the manta trawl. However, this is a rough approximation because the trawl is not  
707 always fully immersed. Moreover, with the manta trawl, all particles below 300  $\mu\text{m}$  are lost.  
708 This is shown by studies using both sampling methods (Kapp and Yeatman 2018; Lahens et  
709 al. 2018; Xiong et al. 2019) Small particles generally represent the largest share of the total  
710 amount of particles present in natural waters. Therefore, the manta trawl method largely  
711 underestimates the actual MP concentrations, at least in terms of particle numbers.

712 The available data on soil and sediments is relatively scarce. This may be partly  
713 related to the complex and time-consuming procedure required to extract MPs from these  
714 matrices (Hurley et al. 2018b). Some studies report MP concentrations as number of  
715 particles per kg, while others provide the weight of MPs per kg. In other cases, data is  
716 reported as MP number or weight per surface unit (e.g.  $\text{mg}/\text{m}^2$ ). Therefore, the comparison of  
717 literature data is not straightforward.

718 Besides this, existing methods for the identification and counting of MPs are quite  
719 variable. Until recently, it was common practice to solely rely on visual detection (using a  
720 microscope), which may lead to false positive or false negatives. In more recent studies,  
721 visual examination is usually combined with FTIR (Fourier Transform Infrared) or Raman  
722 Spectroscopy, which allows polymer Identification. This is, however, time-consuming and  
723 thus frequently only a sub-sample is subjected to spectroscopic methods. Other studies use  
724 different methods like SEM (Scanning Electron Microscopy), XRF (X-Ray Fluorescence),  
725 Pyr-GC/MS (Pyrolysis interfaced with gas chromatography/mass spectrometry). It has been  
726 observed that MP abundance often varies with the methods used (Song et al. 2015; Mai et  
727 al. 2018; Picò and Barcelo 2019), so analytical results may be difficult to compare across  
728 studies.

729 There is an urgent need for a harmonisation of methods for sampling in different  
730 environmental compartments, sample processing, MP extraction, identification, and counting,  
731 as well as for the units to be used for reporting data. A recent report from GESAMP (Group  
732 of Experts on the Scientific Aspects of Marine Protection) describes and compares methods  
733 for sampling and analysing MaPs and MPs, with particular focus on the marine environment  
734 (GESAMP 2019). Although many problems remain unsolved (e.g. the need for sampling  
735 small size MPs and NPs), the report may represent a valuable starting point for the  
736 development of protocols for large scale monitoring of plastic litter in the environment.

737

## 738 **5.2. Small size micro-nanoplastics: the largest unknowns**

739 Most procedures commonly applied to date allow sampling, processing and  
740 measuring particles down to a minimum size of 20  $\mu\text{m}$ . Only very few studies measured  
741 smaller particles, down to 10  $\mu\text{m}$  (e.g. (Leslie et al. 2017; Simon et al. 2018). In theory, very  
742 small particles and, especially NPs, should be more abundant in the environment, and their  
743 concentrations are expected to increase. Moreover, from a toxicological point of view, NPs  
744 are particularly interesting because it is possible that below a given size (still unknown) they  
745 cross cellular membranes and enter into the cells, with possible interactions in the cellular  
746 content and structure. This represents a substantial difference in comparison to MaPs or  
747 MPs. Indeed MPs cannot be accumulated in biological organs and tissues and may produce  
748 mainly physical stress on living organisms, although the consequences of that may result in  
749 physiological and metabolic alterations. The development of methods for the evaluation and  
750 quantification of small-size MPs and NPs is one of the major research needs to assess the  
751 potential risks for human and environmental health. In particular, detection technologies to  
752 identify nano-sized plastic particles are still lacking (Mai et al. 2018). A promising approach,  
753 at least to quantify the mass and the composition (if not the number of particles), could be

754 the use of Pyr-GC/MS (Hendrickson et al. 2018, Mintenig et al. 2018) coupled with methods  
755 of small size particle separation based on ultrafiltration membrane technologies (Mulder  
756 1998; Judd and Jefferson 2003).

757

### 758 **5.3. Towards a microplastic mass balance and suitable evaluation of environmental** 759 **fluxes**

760 The difficulties to get reliable and comparable results for the concentrations of MPs in  
761 the different environmental compartments, and the limited information regarding some fluxes  
762 among compartments makes the evaluation of a regional and global mass balance of  
763 plastics challenging. However, some first estimates can be made on the basis of the  
764 available data, at least to give an approximate order of magnitude of the contribution of  
765 different sources to surface waters.

766 From the data reported in Fig. 2 and Table S1, it can be concluded that the range of  
767 particles in effluents from WWTPs that include secondary and tertiary treatments spans from  
768 1 to 5,800 MPs/L, with a geometric mean around 29 MPs/L. In non-treated wastewaters the  
769 concentrations range from few particles/L up to more than 100,000, with a geometric mean of  
770 about 242 MPs/L. These data are in reasonable agreement with the percentage of retention  
771 by WWTPs reported by several authors, which ranges from 80% to 99% of the inflowing  
772 particles number (see Section 4.3).

773 The approximated per capita consumption of water in Europe is 140 L per day  
774 (EUROSTAT, 2015). Although with some regional differences, it may be estimated that about  
775 85% of the EU population (525 millions in the EU plus Norway and Switzerland) is  
776 connected to WWTP with secondary or tertiary treatment, while the rest (15%) is connected  
777 to a WWTP with only primary treatment or not connected at all (Table 1).

778

779 From these data, it can be estimated that the daily input of MPs (in the range 20 to  
780 5000  $\mu\text{m}$ ) via wastewater into European surface waters is:

- 781 • from treated wastewater: an average value of 1,800E+9 particles per day (possible  
782 range from 9E+9 to 130E+12 particles/day)
- 783 • from untreated wastewater: an average value of 2,700E+9 particles per day (possible  
784 range from 27E+9 to 1,400E+12 particles/day).

785

786 Transforming these data on a weight basis is not easy because, in general, only  
787 numbers of MPs are reported, while size/weight conversion factors are not readily available.  
788 Combined data on numbers and weight are reported in a Danish report (Vollertsen and

789 Hansen 2017) assessing MPs occurrence in ten different WWTP, and in the study by Simon  
790 et al. (2018). However, both studies took only MPs between 10 or 20 and 500  $\mu\text{m}$  into  
791 account. Therefore, estimating the load on a weight basis from the particle numbers is not  
792 possible.

793

794 Despite their wide range of variability, these estimates give a first approximation of  
795 the load of MPs in surface waters from urban wastewater and allow the following  
796 observations. First, the load that may be attributed to the relatively small percentage of  
797 European untreated wastewaters is much higher than the load deriving from treated  
798 wastewater, which points towards a definite need of implementing secondary and tertiary  
799 WWTPs in areas that are still not connected to reduce total MPs emission. Taking into  
800 account that untreated wastewater is concentrated in south-eastern Europe, it may be  
801 hypothesized that some watersheds (e.g. lower Danube) are subject to higher contamination  
802 than those located in other European regions (Lechner et al. 2014). Unfortunately, data on  
803 MP concentrations in surface waters of south-eastern Europe are not available. Due to the  
804 scarcity of data of water consumption and WWTP implementation, a comparable evaluation  
805 cannot be done for other continents. However, it may be hypothesized that the percentage of  
806 treated wastewater in Asia and Africa is much lower than in Europe or North America.

807 The problem is also complicated by the fact that only a relatively small part of the  
808 population is connected to sewerage systems. Data from the WHO/UNICEF Joint Monitoring  
809 Programme (JMP), referred to 2015, indicate that in Eastern, South-eastern and Central  
810 Asia, with a population of more than four billion inhabitants, only 25% of the population is  
811 connected with sewerage systems; and in Sub-Saharan Africa the percentage is lower than  
812 6% (WHO/UNICEF 2019). The high concentrations of MPs in surface waters of Asia (mostly  
813 in China), as compared to those measured in Europe (Fig. 5), supports the hypothesis  
814 regarding the large influence of WWTP on surface water emissions.

815

816 The total values calculated in this study seem relatively low, particularly for treated  
817 waters, if one considers that they represent emissions at the continental level. Nevertheless,  
818 the low concentrations of MPs in surface waters of Europe, as compared to Asia for  
819 example, seem to be directly related to their own values estimated from wastewater  
820 concentrations may justify the relatively low values measured in surface waters in Europe  
821 (Fig 5 and Tab. 5), all referred to north, central and south European water bodies. On the  
822 other hand, the high values measured in Asia (mostly in China), supports the hypothesis  
823 regarding the large influence of WWTP on surface water emissions.

824 The lowest average weight of MPs in effluents compared to influents (almost 50%)  
825 indicates that the removal efficiency is higher on bigger particles. The dominant shape in  
826 WWTP effluents were fibres, followed by fragments. Only in one case a minor amount  
827 (<10%) of pellets that may be assimilated to primary microbeads was observed.

828 Obviously, wastewater represents only one of the possible pathways of MPs into  
829 surface waters, and as discussed in this study, there is no doubt that surface runoff from  
830 agricultural and urban soils may also represent a major source. Unfortunately, a comparable  
831 estimate of MPs emissions from soils due to water runoff is not possible due to field data  
832 limitations. On the other hand, this review shows that MP concentrations in WWTP sludge  
833 (mainly from Europe) range between  $10E+3$  and  $10E+5$  particles/kg dw. Nizzetto et al.  
834 (2016) estimated that the total yearly input of MPs from sewage sludge to farmland is about  
835 63,000-430,000 tons in Europe, and 44,000-300,000 tons in North America. Data on MP  
836 concentrations in soil are scarce and scattered (Fig. 4 and Tab S3). The majority of data on  
837 agricultural soils refer to China and indicate a reduced range of variability (from about 60 to  
838 200 particles/kg dw), except for a couple of higher values (more than 10,000 particles/kg dw)  
839 from soils sampled in a greenhouse. Overall this study shows that soil could be considered  
840 as a sink as well as a source of MPs to surface water. Therefore, further research is urgently  
841 required to assess fluxes of MPs from soils into surface water ecosystems and to assess the  
842 fate of MPs in the soil ecosystems, investigating its retention potential and the capacity of  
843 MPs to reach groundwater ecosystems. An additional source of MPs to soil and surface  
844 water may be atmospheric fall-out (Dris et al. 2016). However, the information available to  
845 date does not yet allow a quantitative estimate (Wetherbee et al. 2019).

846

847 MaP fragmentation in the different compartments is reasonably one of the major  
848 sources of MPs in the environment. However, the patterns of MaPs fragmentation, their  
849 characterization and quantification in terms of amount produced and time to produce them  
850 are still largely unknown. The only fragmentation pattern that is sufficiently documented and  
851 quantified is the production of fibers during laundry of synthetic fabrics (Browne et al. 2011;  
852 Eerkes-Medrano et al. 2015). Although the amount of fibres may vary depending on the type  
853 of clothes (e.g. polymer composition, weave type, age), the type of washing machine, and  
854 the washing condition, it has been estimated that several thousand fibres are generated per  
855 washing cycle (Hartline et al. 2016; Napper and Thompson 2016; Pirc et al. 2016; Carney  
856 Almroth et al. 2018).

857 For any other type of plastic breakdown process, reliable quantitative information is  
858 not yet available. Plastic fragmentation in the environment may be extremely variable in  
859 function of factors like light intensity, temperature, erosion and other physical impacts. The

860 number and weight of MPs and NPs that may be produced by a MaP item (e.g. a bag or a  
861 bottle) in a given time under environmental conditions is still largely unknown. This is an  
862 important knowledge gap that must be investigated in depth, and that may be somewhat  
863 inferred based on the amount and type of polymers of MaP litter in the environment and their  
864 documented half-lives.

865  
866 It has been known for a long time that, although plastic polymers are persistent  
867 compounds, some polymers can undergo biodegradation (Albertsson et al. 1987). Scientific  
868 evidence of biodegradation through bacterial activity and invertebrate digestion mechanisms  
869 has increased recently (Briassoulis et al. 2015; Yoshida et al. 2016; Yang et al. 2018).  
870 Compared to MaPs, MPs and NPs may be more readily attacked by this bacterial and  
871 invertebrate activity. Therefore, a real possibility of their complete disappearance exists.  
872 Nevertheless, to date, the extent of these degradation processes in environmental  
873 compartments, their time scale as well as the patterns and the end-products are fully  
874 unknown (SAPEA 2018). Although plastic polymers are practically inert molecules, with low  
875 biological and toxicological activity, many monomers, that can be formed during the  
876 degradation of plastic, are not. Monovinylchloride (the monomer of PVC), for instance, is a  
877 recognised carcinogenic compound (Brandt-Rauf et al. 2012).

878  
879

#### 880 **5.4. Microplastics in environmental compartments: what does it mean in** 881 **terms of exposure for living organisms?**

882

883 As discussed above, information on the presence of MPs in environmental  
884 compartments is often biased by the inconsistency of units (e.g. n/L, n/m<sup>2</sup>, mg/L, n/kg,  
885 mg/kg), by the variability in size classes sampled and measured, and by the complexity in  
886 shape and composition that are often not clearly reported. These inconsistencies make the  
887 assessment of their possible impact on living organisms rather complex, so the actual  
888 environmental risks of different plastics and their associated chemicals remain largely  
889 unknown (Koelmans et al. 2017). It is important to highlight that the effects of MPs on living  
890 organisms cannot be quantified by a simple concentration-response relationship of the  
891 whole mass of MPs of certain type found in environmental samples, as for most chemical  
892 contaminants. Their impacts on aquatic organisms depend on a number of factors such as:

- 893 • the shape: the physical effect determined by long and thin fibres may be completely  
894 different from those determined by microspheres or by irregular fragments (Au et al.  
895 2015; Lambert et al. 2017);



- 896 • the size range: the definition of MPs in term of size is extremely wide (from 5 mm to 1  
897  $\mu\text{m}$ ) and the living organisms that may be affected by MPs are also extremely variable in  
898 size. For example, in the aquatic environment, from fish to zooplankton; for any type and  
899 size of organism, different MP size classes may be ingested and thus effective, including  
900 small sizes (below 20  $\mu\text{m}$ ) and NPs, that are practically never measured;
- 901 • the composition: for most MP polymers, being the effects mainly physical, it may be  
902 hypothesised that the response is not related to the polymer composition; however, for  
903 some particular MP particles, such as for tyre debris, the composition is much more  
904 complex and the effects may also be determined by the leaching of non-polymeric  
905 chemicals.

906

907 It follows that the available information on the presence of MPs in the environmental  
908 compartments does not allow, to perform an ecological risk assessment based on a  
909 comparison between an environmental exposure (e.g. a PEC: predicted environmental  
910 concentration) and an effect level (e.g. a PNEC: predicted no effect concentration). So far  
911 only An ecological risk assessment of MPs would require much more detailed information on  
912 MP exposure with a precise assessment of number (or weight) of particles per size classes,  
913 shape and composition. Considering that current methods for the analysis of MPs are  
914 complex, expensive and time consuming, this level of detail is, to date, difficult to be  
915 achieved. Moreover, ecotoxicological tests have been frequently carried out using PE  
916 microspheres, while other polymers and especially other shapes like fragments and fibres  
917 are expected to be more abundant in the environment (de Sá et al. 2018). Further research  
918 must be devoted to both areas, to refine exposure assessments and to perform effect  
919 assessments taking into account ecologically relevant combinations of organisms and MPs  
920 sizes, shapes and types. It is most likely that future risk assessments need to necessarily  
921 consider MP particle mixtures taking into account different polymer type, shape and size, and  
922 that exposure and risk indicators are derived taking all these variables into account.

923 Regarding the effect assessment, the major unknown issues are related to small and  
924 very small particles (Koelmans 2019). As mentioned above, the size threshold below which  
925 these particles may enter in the cells is still unknown. Moreover, once they enter in the cells,  
926 the possible interactions of these, theoretically chemically inert polymeric molecules, with cell  
927 structure and functioning are also unknown. Recent studies on NPs performed with reference  
928 materials painted with fluorescent dye demonstrate their capacity to be taken up, enter  
929 tissues, and accumulate in small organisms (Cui et al. 2017; Lee et al. 2019). However,  
930 some authors discuss that this can be an artefact created by the leaching of those dye  
931 paints, which can be taken up into cells or due to the autofluorescence of the evaluated  
932 biological tissues (Catarino et al. 2019; Schür et al. 2019).

### 933 **5.5 How can MP inputs in the environment be controlled?**

934 From all the considerations mentioned above, it is evident that the precautionary  
935 principles strongly push towards the control of MPs and NPs. From the available literature on  
936 MP presence in the environment, it appears that primary MPs represent a relatively small  
937 amount of the total bulk of MPs detected, being secondary MPs (i.e., textile fibres, fragments  
938 from MaP breakdown, tyre debris, etc.) the largest majority. It is difficult to quantify the  
939 percentage of primary MPs in the environment precisely. However, in general, it seems to be  
940 never higher than 10%, and in most cases the percentage is much lower, sometimes almost  
941 negligible. For example, in urban wastewater, the majority of MPs is represented by textile  
942 fibres (see for example Dris et al., 2015; Vollertsen and Hansen 2017; Wang et al. 2017)  
943 while in runoff water the most abundant particles are fragments from MaP breakdown (see  
944 for example Liu et al. 2019a). Therefore, the recent proposal of ECHA (2019) for a ban or  
945 restriction of primary MPs may have a limited relevance and effectiveness for the reduction  
946 of the presence of MPs in the environment.

947 Regarding the information available to date, the most plausible solution for reducing  
948 the environmental emission and exposure to MPs seems to be the control of MaPs. The  
949 restrictions on single use plastic items that will be active in Europe starting from 2021 (EC  
950 2019) seem to be a very good starting point. Comparable restrictions should be applied in  
951 the short-term on food and other kinds of packaging, which represent the largest amount of  
952 plastic wastes. In addition to restrictions, a more efficient recycling strategy and improvement  
953 of circular economy related to plastic products would be beneficial (Barra and Leonard 2018).  
954 However, in some cases, different types of measures should be developed. As shown above,  
955 fibres represent the most abundant type of MPs present in wastewater. Since it is almost  
956 impossible to ban synthetic fabrics that today make up the majority of our clothing, the  
957 solution should be sought in another direction (e.g. by means of retaining fibres in washing  
958 machines, water treatment procedures, etc.).

959 Finally, the substitution of traditional plastic polymers, based on the petrochemical  
960 industry, with new generation polymers, based on biological resources (e.g. PLA: polylactic  
961 acid; PHA: polyhydroxyalkanoates) is often proposed as a suitable solution. However,  
962 present knowledge on the toxicological properties of these new compounds and of their  
963 degradation products must be improved (Lambert and Wagner 2017; Picó and Barceló  
964 2019). Understanding possible biodegradation patterns of traditional and emerging plastic  
965 polymers is important for future management and remediation of plastics in the environment.

966

967

## 968 **6. Conclusions**

969

970  
971 In this study we have described the state of the knowledge regarding the occurrence  
972 of MaPs and MPs in different environmental compartments. It has been highlighted that  
973 some data gaps still exist in order to better understand their life cycle, to develop a precise  
974 mass balance and to quantitatively assess the contribution of the different main sources of  
975 MaPs, MPs and NPs in the environment. The emission of MPs from WWTPs into aquatic  
976 ecosystems is the environmental pathway that has been most researched. However there  
977 are other pathways that may have similar or even larger contributions, and that require  
978 further investigation. For example, the fluxes of plastics from landfills and agricultural soils  
979 towards surface and groundwater ecosystems by water runoff or deep-horizon infiltration, or  
980 the transport and deposition of plastic particles from the atmosphere. Moreover, quantitative  
981 evaluations of the occurrence of large-size plastics in natural environments need to be  
982 performed, and their breakdown rates into MPs and NPs still need to be assessed under  
983 different environmental conditions (i.e. temperature and light intensities, water currents).

984 There is enough experimental evidence demonstrating that the presence of MaPs in aquatic  
985 ecosystems represent an environmental risk, particularly for large animals. Regarding MPs, a  
986 risk for human and environmental health has not been demonstrated (EC SAM 2019;  
987 GESAMP 2019). All available toxicological evidence indicates that some effects on aquatic  
988 and terrestrial organisms, vertebrates and invertebrates, have been observed only at  
989 concentrations that are orders of magnitude higher than the maximum levels measured in the  
990 environment (Lenz et al. 2016; Redondo Hasselerharm et al. 2018). Other possible effects,  
991 such as a potential increase in the bioaccumulation of chemicals due to their transport into  
992 the organisms adsorbed on MPs (the “Trojan horse effect”) seems to be context dependent,  
993 and negligible in comparison to direct accumulation from the surrounding environment (e.g.  
994 from water) or from food (Koelmans et al. 2013, 2014; Lohmann 2017; Mohamed Nor and  
995 Koelmans 2019). However, research is still needed to demonstrate this experimentally.

996 Current knowledge gaps regarding environmental fluxes and breakdown of MPs and  
997 NPs are still large in order to assess future risks for man and for the environment.  
998 Furthermore, the bias on sampling and analysis makes a precise quantification challenging.  
999 This is particularly difficult for small MPs and NPs, which are probably the more concerning  
1000 particles from a toxicological point of view. Moreover, although present exposure seems to  
1001 be far away from levels of concern, it is difficult to predict future emission patterns since they  
1002 will be closely related to plastic use and management policies. This review shows that the  
1003 construction of waste-water treatment facilities and the proper management of sludge  
1004 applications in agriculture are efficient means to reduce MPs emissions. Moreover, the ban  
1005 of single-use plastics, the substitution of some plastic polymers with biodegradable  
1006 compounds, and the reduction of MPs emission at a source are key to control plastic

1007 pollution. From now onwards, we expect technological solutions to be developed and  
1008 implemented in this direction. There is no doubt that plastics changed our life in the middle of  
1009 last century, and the control of plastics will again change our life in the near future.

1010

## 1011 **7. List of abbreviations**

1012

1013	ATR	Attenuated Total Reflectance
1014	ECHA	European Chemical Agency
1015	EEA	European Environmental Agency
1016	ERA	Ecological Risk Assessment
1017	EVA	Ethylene-vinyl-acetate
1018	FTIR	Fourier Transform Infrared
1019	MaP	Macroplastic
1020	MP	Microplastic
1021	NP	Nanoplastic
1022	PAH	Polycyclic aromatic hydrocarbons
1023	PC	Polycarbonate
1024	PCB	Polychlorinated biphenyl
1025	PE	Polyethylene
1026	PEC	Predicted Environmental Concentration
1027	PES	Polyester
1028	PET	Polyethylene-terephthalate
1029	PMMA	Poly-methyl-metacrylate
1030	PNEC	Predicted No Effect Concentration
1031	PP	Polypropylene
1032	PS	Polystyrene
1033	PUR	Polyurethane
1034	PVC	Polyvinylchloride
1035	Pyr-GC/MS	Pyrolysis-Gas Chromatography/Mass Spectrometry
1036	SAPEA	Science Advice for Policy by European Academies
1037	SEM	Scanning Electron Microscopy
1038	WWTP	Waste Water Treatment Plant
1039	XRF	X-Ray Fluorescence

1040

1041

1042

## 1043 **8. Acknowledgements**

1044

1045 The study has been conducted as part of the EU JPI-Water initiative IMPASSE project  
1046 (Impacts of MicroPlastics in AgroSystems and Stream Environments, PCIN-2017-016). A.  
1047 Rico is supported by a postdoctoral grant provided by the Spanish Ministry of Science,  
1048 Innovation and University (IJCI-2017-33465).

1049

1050

1051 On behalf of all authors, the corresponding author states that there is no conflict of interest.

1052

1053

## 1054 **9. References**

1055

- 1056 Abbasi S, Keshavarzi B, Moore F, et al (2019) Distribution and potential health impacts of  
1057 microplastics and microrubbers in air and street dusts from Asaluyeh County, Iran. *Environ Pollut*  
1058 244:153–164. doi: 10.1016/j.envpol.2018.10.039  
1059 Adane L, Muleta D (2009) Survey on the usage of plastic bags, their disposal and adverse impacts on

1060 environment: A case study in Jimma City, Southwestern Ethiopia. *J Toxicol Environ Heal Sci*  
1061 3:234–248

1062 Albertsson A-C, Andersson SO, Karlsson S (1987) The mechanism of biodegradation of polyethylene.  
1063 *Polym Degrad Stab* 18:73–87. doi: 10.1016/0141-3910(87)90084-X

1064 Anderson PJ, Warrack S, Langen V, et al (2017) Microplastic contamination in Lake Winnipeg,  
1065 Canada. *Environ Pollut* 225:223–231. doi: 10.1016/j.envpol.2017.02.072

1066 Andrady AL (2011) Microplastics in the marine environment. *Mar Pollut Bull* 62:1596–1605. doi:  
1067 10.1016/j.marpolbul.2011.05.030

1068 Au SY, Bruce TF, Bridges WC, Klaine SJ (2015) Responses of *Hyalella azteca* to acute and chronic  
1069 microplastic exposures. *Environ Toxicol Chem.* 34:2564-2572.

1070 Auta HS, Emenike CU, Fauziah SH (2017) Distribution and importance of microplastics in the marine  
1071 environment: A review of the sources, fate, effects, and potential solutions. *Environ Int* 102:165–  
1072 176. doi: 10.1016/j.envint.2017.02.013

1073 Baldwin AK, Corsi SR, Mason SA (2016) Plastic Debris in 29 Great Lakes Tributaries: Relations to  
1074 Watershed Attributes and Hydrology. *Environ Sci Technol* 50:10377–10385. doi:  
1075 10.1021/acs.est.6b02917

1076 Ballent A, Corcoran PL, Madden O, et al (2016) Sources and sinks of microplastics in Canadian Lake  
1077 Ontario nearshore, tributary and beach sediments. *Mar Pollut Bull* 110:383–395. doi:  
1078 10.1016/j.marpolbul.2016.06.037

1079 Bandmann V, Müller JD, Köhler T, Homann U (2012) Uptake of fluorescent nano beads into BY2-cells  
1080 involves clathrin-dependent and clathrin-independent endocytosis. *FEBS Lett* 586:3626–3632.  
1081 doi: 10.1016/j.febslet.2012.08.008

1082 Barboza LGA, Gimenez BCG (2015) Microplastics in the marine environment: Current trends and  
1083 future perspectives. *Mar Pollut Bull* 97:5–12. doi: 10.1016/j.marpolbul.2015.06.008

1084 Barra R, Leonard SA (2018) Plastics and the circular economy.  
1085 [https://www.thegef.org/sites/default/files/publications/PLASTICS\\_for\\_posting.pdf](https://www.thegef.org/sites/default/files/publications/PLASTICS_for_posting.pdf) .

1086 Barrows APW, Christiansen KS, Bode ET, Hoellein TJ (2018) A watershed-scale, citizen science  
1087 approach to quantifying microplastic concentration in a mixed land-use river. *Water Res*  
1088 147:382–392. doi: 10.1016/j.watres.2018.10.013

1089 Blettler MCM, Ulla MA, Rabuffetti AP, Garelo N (2017) Plastic pollution in freshwater ecosystems:  
1090 macro-, meso-, and microplastic debris in a floodplain lake. *Environ Monit Assess* 189:. doi:  
1091 10.1007/s10661-017-6305-8

1092 Bouhroum R, Boulkamh A, Asia L, et al (2019) Concentrations and fingerprints of PAHs and PCBs  
1093 adsorbed onto marine plastic debris from the Indonesian Cilacap coast and the North Atlantic  
1094 gyre. *Reg Stud Mar Sci* 29:100611. doi: 10.1016/J.RSMA.2019.100611

1095 Brandt-Rauf P, Long C, Kovvali G, et al (2012) Plastics and carcinogenesis: The example of vinyl  
1096 chloride. *J Carcinog* 11:5. doi: 10.4103/1477-3163.93700

1097 Briassoulis D, Babou E, Hiskakis M, Kyrikou I (2015) Analysis of long-term degradation behaviour of  
1098 polyethylene mulching films with pro-oxidants under real cultivation and soil burial conditions.  
1099 *Environ Sci Pollut Res* 22:2584–2598. doi: 10.1007/s11356-014-3464-9

1100 Browne MA, Crump P, Niven SJ, et al (2011) Accumulations of microplastic on shorelines worldwide:  
1101 sources and sinks. *Environ Sci Technol* 9175–9179. doi: 10.1021/es201811s

1102 Carney Almroth BM, Åström L, Roslund S, et al (2018) Quantifying shedding of synthetic fibers from  
1103 textiles; a source of microplastics released into the environment. *Environ Sci Pollut Res*  
1104 25:1191–1199. doi: 10.1007/s11356-017-0528-7

1105 Carr SA, Liu J, Tesoro AG (2016) Transport and fate of microplastic particles in wastewater treatment  
1106 plants. *Water Res* 91:174–182. doi: 10.1016/j.watres.2016.01.002

1107 Castañeda RA, Avlijas S, Simard MAA, et al (2014) Microplastic pollution in St. Lawrence River  
1108 sediments. *Can J Fish Aquat Sci* 71:1767–1771. doi: 10.1139/cjfas-2014-0281

1109 Castro-Jiménez J, González-Fernández D, Fornier M, et al (2019) Macro-litter in surface waters from  
1110 the Rhone River: Plastic pollution and flows to the NW Mediterranean Sea. *Mar Pollut Bull*  
1111 146:60–66. doi: 10.1016/j.marpolbul.2019.05.067

1112 Catarino AI, Frutos A, Henry TB (2019) Use of fluorescent-labelled nanoplastics (NPs) to demonstrate  
1113 NP absorption is inconclusive without adequate controls. *Sci Total Environ* 670:915–920. doi:  
1114 10.1016/j.scitotenv.2019.03.194

1115 Chae Y, An Y-J (2018) Current research trends on plastic pollution and ecological impacts on the soil  
1116 ecosystem: A review. *Environ Pollut* 240:387–395. doi: 10.1016/j.envpol.2018.05.008

1117 Claessens M, Meester S De, Landuyt L Van, et al (2011) Occurrence and distribution of microplastics  
1118 in marine sediments along the Belgian coast. *Mar Pollut Bull* 62:2199–2204. doi:  
1119 10.1016/j.marpolbul.2011.06.030

1120 Corcoran PL, Norris T, Ceccanese T, et al (2015) Hidden plastics of Lake Ontario, Canada and their

1121 potential preservation in the sediment record. *Environ Pollut* 204:17–25. doi:  
1122 10.1016/j.envpol.2015.04.009

1123 Corradini F, Bartholomeus H, Lwanga EH, et al (2019a) Predicting soil microplastic concentration  
1124 using vis-NIR spectroscopy. *Sci Total Environ* 650:922–932. doi:  
1125 <https://doi.org/10.1016/j.scitotenv.2018.09.101>

1126 Corradini F, Meza P, Eguiluz R, et al (2019b) Evidence of microplastic accumulation in agricultural  
1127 soils from sewage sludge disposal. *Sci Total Environ* 671:411–420. doi:  
1128 10.1016/j.scitotenv.2019.03.368

1129 Crawford CB, Quinn B (2017) The interactions of microplastics and chemical pollutants. *Microplastic*  
1130 *Pollut* 131–157. doi: 10.1016/B978-0-12-809406-8.00006-2

1131 Crosti R, Arcangeli A, Campana I, et al (2018) ‘Down to the river’: amount, composition, and economic  
1132 sector of litter entering the marine compartment, through the Tiber river in the Western  
1133 Mediterranean Sea. *Rend Lincei* 29:859–866. doi: 10.1007/s12210-018-0747-y

1134 Cui R, Kim SW, An YJ (2017) Polystyrene nanoplastics inhibit reproduction and induce abnormal  
1135 embryonic development in the freshwater crustacean *Daphnia galeata*. *Sci Rep* 7:1–10. doi:  
1136 10.1038/s41598-017-12299-2

1137 Dall’Osto M, Beddows DCS, Gietl JK, et al (2014) Characteristics of tyre dust in polluted air: Studies  
1138 by single particle mass spectrometry (ATOFMS). *Atmos Environ* 94:224–230. doi:  
1139 10.1016/j.atmosenv.2014.05.026

1140 de Sá LC, Oliveira M, Ribeiro F, et al (2018) Studies of the effects of microplastics on aquatic  
1141 organisms: What do we know and where should we focus our efforts in the future? *Sci Total*  
1142 *Environ* 645:1029–1039. doi: 10.1016/j.scitotenv.2018.07.207

1143 Di M, Wang J (2018) Microplastics in surface waters and sediments of the Three Gorges Reservoir,  
1144 China. *Sci Total Environ* 616–617:1620–1627. doi: 10.1016/j.scitotenv.2017.10.150

1145 Dris R, Gasperi J, Rocher V, et al (2015) Microplastic contamination in an urban area: A case study in  
1146 Greater Paris. *Environ Chem* 12:592–599. doi: 10.1071/EN14167

1147 Dris R, Gasperi J, Saad M, et al (2016) Synthetic fibers in atmospheric fallout: A source of  
1148 microplastics in the environment? *Mar Pollut Bull* 104:290–293. doi:  
1149 10.1016/j.marpolbul.2016.01.006

1150 Dyachenko A, Mitchell J, Arsem N (2017) Extraction and identification of microplastic particles from  
1151 secondary wastewater treatment plant (WWTP) effluent. *Anal Methods* 9:1412–1418. doi:  
1152 10.1039/c6ay02397e

1153 EC (European Commission) (2019) Press Release Database. [http://europa.eu/rapid/press-](http://europa.eu/rapid/press-release_STATEMENT-19-1873_en.htm)  
1154 [release\\_STATEMENT-19-1873\\_en.htm](http://europa.eu/rapid/press-release_STATEMENT-19-1873_en.htm)

1155 ECHA (2019) Proposal for a restriction on intentionally added microplastics. Version n. 1.1. 20 March  
1156 2019, European Chemicals Agency (ECHA), Helsinki, Finland

1157 Eerkes-Medrano D, Thompson RC, Aldridge DC (2015) Microplastics in freshwater systems: A review  
1158 of the emerging threats, identification of knowledge gaps and prioritisation of research needs.  
1159 *Water Res* 75:63–82. doi: 10.1016/j.watres.2015.02.012

1160 Eriksen M, Mason S, Wilson S, et al (2013) Microplastic pollution in the surface waters of the  
1161 Laurentian Great Lakes. *Mar Pollut Bull* 77:177–182. doi: 10.1016/j.marpolbul.2013.10.007

1162 Estahbanati S, Fahrenfeld NL (2016) Influence of wastewater treatment plant discharges on  
1163 microplastic concentrations in surface water. *Chemosphere* 162:277–284. doi:  
1164 10.1016/j.chemosphere.2016.07.083

1165 Everaert G, Van Cauwenberghe L, De Rijcke M, et al (2018) Risk assessment of microplastics in the  
1166 ocean: Modelling approach and first conclusions. *Environ Pollut* 242:1930–1938. doi:  
1167 10.1016/j.envpol.2018.07.069

1168 Faure F, Corbaz M, Baecher H, De Alencastro LF (2012) Pollution due to plastics and microplastics in  
1169 lake Geneva and in the Mediterranean Sea. *Arch des Sci* 65:157–164. doi: 10.1071/EN14218

1170 Faure F, Demars C, Wieser O, et al (2015) Plastic pollution in Swiss surface waters: Nature and  
1171 concentrations, interaction with pollutants. *Environ Chem* 12:582–591. doi: 10.1071/EN14218

1172 Fendall LS, Sewell MA (2009) Contributing to marine pollution by washing your face: Microplastics in  
1173 facial cleansers. *Mar Pollut Bull* 58:1225–1228. doi: 10.1016/j.marpolbul.2009.04.025

1174 Fischer EK, Paglialonga L, Czech E, Tamminga M (2016) Microplastic pollution in lakes and lake  
1175 shoreline sediments - A case study on Lake Bolsena and Lake Chiusi (central Italy). *Environ*  
1176 *Pollut* 213:648–657. doi: 10.1016/j.envpol.2016.03.012

1177 Foley CJ, Feiner ZS, Malinich TD, Höök TO (2018) A meta-analysis of the effects of exposure to  
1178 microplastics on fish and aquatic invertebrates. *Sci Total Environ* 631–632:550–559. doi:  
1179 10.1016/j.scitotenv.2018.03.046

1180 Free CM, Jensen OP, Mason SA, et al (2014) High-levels of microplastic pollution in a large, remote,  
1181 mountain lake. *Mar Pollut Bull* 85:156–163. doi: 10.1016/j.marpolbul.2014.06.001

1182 Fuller S, Gautam A (2016) A Procedure for Measuring Microplastics using Pressurized Fluid  
1183 Extraction. *Environ Sci Technol* 50:5774–5780. doi: 10.1021/acs.est.6b00816  
1184 Gasperi J, Dris R, Bonin T, et al (2014) Assessment of floating plastic debris in surface water along  
1185 the Seine River. *Environ Pollut* 195:163–166. doi: 10.1016/j.envpol.2014.09.001  
1186 Gasperi J, Wright SL, Dris R, et al (2018) Microplastics in air: Are we breathing it in? *Curr Opin*  
1187 *Environ Sci Heal* 1:1–5. doi: 10.1016/j.coesh.2017.10.002  
1188 GESAMP (2019). Guidelines on the monitoring and assessment of plastic litter and microplastics in the  
1189 ocean (Kershaw P.J., Turra A. and Galgani F. editors), (IMO/FAO/UNESCO-  
1190 IOC/UNIDO/WMO/IAEA/UN/UNEP/UNDP/ISA Joint Group of Experts on the Scientific Aspects  
1191 of Marine Environmental Protection). Rep. Stud. GESAMP No. 99, 130p.  
1192 [https://environmentlive.unep.org/media/docs/marine\\_plastics/une\\_science\\_dvision\\_gesamp\\_re](https://environmentlive.unep.org/media/docs/marine_plastics/une_science_dvision_gesamp_reports.pdf)  
1193 [ports.pdf](https://environmentlive.unep.org/media/docs/marine_plastics/une_science_dvision_gesamp_reports.pdf). Accessed 06.06.2019  
1194 Geyer R, Jambeck JR, Law KL (2017) Production, use, and fate of all plastics ever made. *Sci Adv*  
1195 3:e1700782. doi: 10.1126/sciadv.1700782  
1196 Gies EA, Lenoble JL, Noël M, et al (2018) Retention of microplastics in a major secondary wastewater  
1197 treatment plant in Vancouver, Canada. *Mar Pollut Bull* 133:553–561. doi:  
1198 10.1016/j.marpolbul.2018.06.006  
1199 Gigault J, Halle A ter, Baudrimont M, et al (2018) Current opinion: What is a nanoplastic? *Environ*  
1200 *Pollut* 235:1030–1034. doi: 10.1016/j.envpol.2018.01.024  
1201 González-Fernández D, Hanke G, Network and the R (2018) Floating Macro Litter in European Rivers  
1202 - Top Items. *Publ Off Eur Union EUR* 29383: doi: 10.2760/316058  
1203 Gouin T, Avalos J, Brunning I, et al (2015) Use of Micro-Plastic Beads in Cosmetic Products in Europe  
1204 and Their Estimated Emissions to the North Sea Environment. *SOFW J* 141:40–46.  
1205 Gouin T, Roche N, Lohmann R, Hodges G (2011) A Thermodynamic Approach for Assessing the  
1206 Environmental Exposure of Chemicals Absorbed to Microplastic. *Environ Sci Technol* 45:1466–  
1207 1472. doi: 10.1021/es1032025  
1208 Gündoğdu S, Çevik C, Güzel E, Kilercioğlu S (2018) Microplastics in municipal wastewater treatment  
1209 plants in Turkey: a comparison of the influent and secondary effluent concentrations. *Environ*  
1210 *Monit Assess* 190:. doi: 10.1007/s10661-018-7010-y  
1211 Hardesty BD, Good TP, Wilcox C (2015) Novel methods, new results and science-based solutions to  
1212 tackle marine debris impacts on wildlife. *Ocean Coast Manag* 115:4–9. doi:  
1213 10.1016/j.ocecoaman.2015.04.004  
1214 Hartline NL, Bruce NJ, Karba SN, et al (2016) Microfiber Masses Recovered from Conventional  
1215 Machine Washing of New or Aged Garments. *Environ Sci Technol* 50:11532–11538. doi:  
1216 10.1021/acs.est.6b03045  
1217 HELCOM (2014) Preliminary study on synthetic microfibers and particles at a municipal waste water  
1218 treatment plant. *Balt Mar Environ Prot Comm HELCOM* 14 p.  
1219 [http://www.helcom.fi/Lists/Publications/Microplastics%20at%20a%20municipal%20waste%20wat](http://www.helcom.fi/Lists/Publications/Microplastics%20at%20a%20municipal%20waste%20water%20treatment%20plant.pdf)  
1220 [er%20treatment%20plant.pdf](http://www.helcom.fi/Lists/Publications/Microplastics%20at%20a%20municipal%20waste%20water%20treatment%20plant.pdf). Accessed 20.04.2019  
1221 Hendrickson E, Minor EC, Schreiner K (2018) Microplastic Abundance and Composition in Western  
1222 Lake Superior As Determined via Microscopy, Pyr-GC/MS, and FTIR. *Environ Sci Technol*  
1223 52:1787–1796. doi: 10.1021/acs.est.7b05829  
1224 Hopewell J, Dvorak R, Kosior E (2009) Plastics recycling: challenges and opportunities. *Philos Trans*  
1225 *R Soc B Biol Sci* 364:2115–2126. doi: 10.1098/rstb.2008.0311  
1226 Horton AA, Svendsen C, Williams RJ, et al (2017) Large microplastic particles in sediments of  
1227 tributaries of the River Thames, UK – Abundance, sources and methods for effective  
1228 quantification. *Mar Pollut Bull* 114:218–226. doi: 10.1016/j.marpolbul.2016.09.004  
1229 Hu L, Chernick M, Hinton DE, Shi H (2018) Microplastics in Small Waterbodies and Tadpoles from  
1230 Yangtze River Delta, China. *Environ Sci Technol* 52:8885–8893. doi: 10.1021/acs.est.8b02279  
1231 Huerta Lwanga E, Gertsen H, Gooren H, et al (2017a) Incorporation of microplastics from litter into  
1232 burrows of *Lumbricus terrestris*. *Environ Pollut* 220:523–531. doi: 10.1016/j.envpol.2016.09.096  
1233 Huerta Lwanga E, Vega JM, Quej VK, et al (2017b) Field evidence for transfer of plastic debris along a  
1234 terrestrial food chain. 1–7. doi: 10.1038/s41598-017-14588-2  
1235 Hurley R, Woodward J, Rothwell JJ (2018a) Microplastic contamination of river beds significantly  
1236 reduced by catchment-wide flooding. *Nat Geosci* 11:251–257. doi: 10.1038/s41561-018-0080-1  
1237 Hurley RR, Lusher AL, Olsen M, Nizzetto L (2018b) Validation of a method for extracting microplastics  
1238 from complex ., doi: 10.1021/acs.est.8b01517  
1239 Hurley RR, Nizzetto L (2018) Fate and occurrence of micro(nano)plastics in soils: Knowledge gaps  
1240 and possible risks. *Curr Opin Environ Sci Heal* 1:6–11. doi: 10.1016/j.coesh.2017.10.006  
1241 Imhof AHK, Ivleva NP, Schmid J, et al (2013) Contamination of beach sediments of a subalpine lake  
1242 with microplastic particles. *Curr Biol* 23:867–868. doi: 10.1016/j.cub.2013.09.001

1243 Imhof HK, Laforsch C, Wiesheu AC, et al (2016) Pigments and plastic in limnetic ecosystems: A  
1244 qualitative and quantitative study on microparticles of different size classes. *Water Res* 98:64–  
1245 74. doi: 10.1016/j.watres.2016.03.015

1246 Imhof HK, Wiesheu AC, Anger PM, et al (2018) Variation in plastic abundance at different lake beach  
1247 zones - A case study. *Sci Total Environ* 613–614:530–537. doi: 10.1016/j.scitotenv.2017.08.300

1248 Jambeck JR, Geyer R, Wilcox C, et al (2015) Plastic waste inputs from land into the ocean. *Science*  
1249 (80- ) 347 (6223):768–771. doi: 10.1126/science.1260352

1250 Jovanović B (2017) Ingestion of microplastics by fish and its potential consequences from a physical  
1251 perspective. *Integr Environ Assess Manag* 13:510–515. doi: 10.1002/ieam.1913

1252 Judd S, Jefferson B (2003) *Membrane for Industrial Wastewater Recovery and Re-Use*. Oxford, UK,  
1253 Elsevier advanced technology.

1254 Kalčíková G, Alič B, Skalar T, et al (2017) Wastewater treatment plant effluents as source of cosmetic  
1255 polyethylene microbeads to freshwater. *Chemosphere* 188:25–31. doi:  
1256 10.1016/j.chemosphere.2017.08.131

1257 Kapp KJ, Yeatman E (2018) Microplastic hotspots in the Snake and Lower Columbia rivers: A journey  
1258 from the Greater Yellowstone Ecosystem to the Pacific Ocean. *Environ Pollut* 241:1082–1090.  
1259 doi: 10.1016/j.envpol.2018.06.033

1260 Karami A, Romano N, Galloway T, Hamzah H (2016) Virgin microplastics cause toxicity and modulate  
1261 the impacts of phenanthrene on biomarker responses in African catfish (*Clarias gariepinus*).  
1262 *Environ Res* 151:58–70. doi: 10.1016/j.envres.2016.07.024

1263 Kataoka T, Nihei Y, Kudou K, Hinata H (2019) Assessment of the sources and inflow processes of  
1264 microplastics in the river environments of Japan. *Environ Pollut* 244:958–965. doi:  
1265 10.1016/j.envpol.2018.10.111

1266 Kay P, Hiscoe R, Moberley I, et al (2018) Wastewater treatment plants as a source of microplastics in  
1267 river catchments. *Environ Sci Pollut Res* 25:20264–20267. doi: 10.1007/s11356-018-2070-7

1268 Keswani A, Oliver DM, Gutierrez T, Quilliam RS (2016) Microbial hitchhikers on marine plastic debris:  
1269 Human exposure risks at bathing waters and beach environments. *Mar Environ Res* 118:10–9.  
1270 doi: 10.1016/j.marenvres.2016.04.006

1271 Kirstein I V., Kirmizi S, Wichels A, et al (2016) Dangerous hitchhikers? Evidence for potentially  
1272 pathogenic *Vibrio* spp. on microplastic particles. *Mar Environ Res* 120:1–8. doi:  
1273 10.1016/j.marenvres.2016.07.004

1274 Klein S, Worch E, Knepper TP (2015) Occurrence and spatial distribution of microplastics in river  
1275 shore sediments of the rhine-main area in Germany. *Environ Sci Technol* 49:6070–6076. doi:  
1276 10.1021/acs.est.5b00492

1277 Knowlton AR, Hamilton PK, Marx MK, et al (2012) Monitoring North Atlantic right whale *Eubalaena*  
1278 *glacialis* entanglement rates: A 30 yr retrospective. *Mar Ecol Prog Ser* 466:293–302. doi:  
1279 10.3354/meps09923

1280 Koelmans AA (2019) Proxies for nanoplastic. *Nat Nanotechnol* 14:307–308. doi: 10.1038/s41565-019-  
1281 0416-z

1282 Koelmans AA, Bakir A, Burton GA, Janssen CR (2016) Microplastic as a Vector for Chemicals in the  
1283 Aquatic Environment: Critical Review and Model-Supported Reinterpretation of Empirical  
1284 Studies. *Environ Sci Technol* 50:3315–3326. doi: 10.1021/acs.est.5b06069

1285 Koelmans AA, Besseling E, Foekema E, et al (2017) Risks of Plastic Debris: Unravelling Fact,  
1286 Opinion, Perception, and Belief. *Environ Sci Technol* 51:11513–11519. doi:  
1287 10.1021/acs.est.7b02219

1288 Koelmans AA, Besseling E, Foekema EM (2014) Leaching of plastic additives to marine organisms.  
1289 *Environ Pollut* 187:49–54. doi: 10.1016/j.envpol.2013.12.013

1290 Koelmans AA, Besseling E, Wegner A, Foekema EM (2013) Plastic as a carrier of POPs to aquatic  
1291 organisms: A model analysis. *Environ Sci Technol* 47:7812–7820. doi: 10.1021/es401169n

1292 Kole PJ, Löhr AJ, Van Belleghem FGJ, Ragas AMJ (2017) Wear and tear of tyres: A stealthy source  
1293 of microplastics in the environment. *Int J Environ Res Public Health* 14:. doi:  
1294 10.3390/ijerph14101265

1295 Kreider ML, Panko JM, McAtee BL, et al (2010) Physical and chemical characterization of tire-related  
1296 particles: Comparison of particles generated using different methodologies. *Sci Total Environ*  
1297 408:652–659. doi: https://doi.org/10.1016/j.scitotenv.2009.10.016

1298 Kühn S, Bravo Rebolledo EL, van Franeker JA (2015) Deleterious Effects of Litter on Marine Life. In:  
1299 Bergmann M, Gutow L, Klages M (eds) *Marine Anthropogenic Litter*. Springer International  
1300 Publishing, Cham, pp 75–116

1301 Kumar A, Holuszko M, Espinosa DCR (2017) E-waste: An overview on generation, collection,  
1302 legislation and recycling practices. *Resour Conserv Recycl* 122:32–42. doi:  
1303 10.1016/j.resconrec.2017.01.018



1304 Lahens L, Strady E, Kieu-Le TC, et al (2018) Macroplastic and microplastic contamination assessment  
1305 of a tropical river (Saigon River, Vietnam) transversed by a developing megacity. *Environ Pollut*  
1306 236:661–671. doi: 10.1016/j.envpol.2018.02.005

1307 Lambert S, Scherer C, Wagner M (2017) Ecotoxicity testing of microplastics: Considering the  
1308 heterogeneity of physicochemical properties. *Integr Environ Assess Manag* 13:470–475. doi:  
1309 10.1002/ieam.1901

1310 Lambert S, Wagner M (2017) Environmental performance of bio-based and biodegradable plastics:  
1311 the road ahead. *Chem Soc Rev* 46:6855–6871. doi: 10.1039/C7CS00149E

1312 Lares M, Ncibi MC, Sillanpää M, Sillanpää M (2018) Occurrence, identification and removal of  
1313 microplastic particles and fibers in conventional activated sludge process and advanced MBR  
1314 technology. *Water Res* 133:236–246. doi: 10.1016/j.watres.2018.01.049

1315 Lebreton LCM, Van Der Zwet J, Damsteeg JW, et al (2017) River plastic emissions to the world's  
1316 oceans. *Nat Commun* 8:1–10. doi: 10.1038/ncomms15611

1317 Lechner A, Keckeis H, Lumesberger-Loisl F, et al (2014) The Danube so colourful: A potpourri of  
1318 plastic litter outnumbers fish larvae in Europe's second largest river. *Environ Pollut* 188:177–181.  
1319 doi: 10.1016/j.envpol.2014.02.006

1320 Lechner A, Ramler D (2015) The discharge of certain amounts of industrial microplastic from a  
1321 production plant into the River Danube is permitted by the Austrian legislation. *Environ Pollut*  
1322 200:159–160. doi: 10.1016/j.envpol.2015.02.019

1323 Lee H, Kim Y (2018) Treatment characteristics of microplastics at biological sewage treatment  
1324 facilities in Korea. *Mar Pollut Bull* 137:1–8. doi: 10.1016/j.marpolbul.2018.09.050

1325 Lee WS, Cho H-J, Kim E, et al (2019) Bioaccumulation of polystyrene nanoplastics and their effect on  
1326 the toxicity of Au ions in zebrafish embryos. *Nanoscale* 11:3173–3185. doi:  
1327 10.1039/C8NR09321K

1328 Lenz R, Enders K, Gissel T (2016) Microplastic exposure studies should be environmentally realistic.  
1329 *Proc Natl Acad Sci U S A* 113:2–3. doi: 10.1073/pnas.1606615113

1330 Leslie HA (2014) Review of Microplastics in Cosmetics. *IVM Inst Environ Stud* 476:33

1331 Leslie HA, Brandsma SH, van Velzen MJM, Vethaak AD (2017) Microplastics en route: Field  
1332 measurements in the Dutch river delta and Amsterdam canals, wastewater treatment plants,  
1333 North Sea sediments and biota. *Environ Int* 101:133–142. doi: 10.1016/j.envint.2017.01.018

1334 Lin L, Zuo LZ, Peng JP, et al (2018) Occurrence and distribution of microplastics in an urban river: A  
1335 case study in the Pearl River along Guangzhou City, China. *Sci Total Environ* 644:375–381. doi:  
1336 10.1016/j.scitotenv.2018.06.327

1337 Liu M, Lu S, Song Y, et al (2018) Microplastic and mesoplastic pollution in farmland soils in suburbs of  
1338 Shanghai, China. *Environ Pollut* 242:855–862. doi: 10.1016/j.envpol.2018.07.051

1339 Liu F, Olesen KB, Borregaard AR, Vollertsen J (2019a) Microplastics in urban and highway  
1340 stormwater retention ponds. *Sci Total Environ* 671:992–1000. doi:  
1341 10.1016/j.scitotenv.2019.03.416

1342 Liu X, Yuan W, Di M, et al (2019b) Transfer and fate of microplastics during the conventional activated  
1343 sludge process in one wastewater treatment plant of China. *Chem Eng J* 362:176–182. doi:  
1344 10.1016/j.cej.2019.01.033

1345 Lohmann R (2017) Microplastics are not important for the cycling and bioaccumulation of organic  
1346 pollutants in the oceans—but should microplastics be considered POPs themselves? *Integr*  
1347 *Environ Assess Manag* 13:460–465. doi: 10.1002/ieam.1914

1348 Luo W, Su L, Craig NJ, et al (2018) Accepted Manuscript Comparison of microplastic pollution in  
1349 different water bodies from urban creeks to coastal waters. *Environ Pollut* 246:174–182. doi:  
1350 10.1016/j.envpol.2018.11.081

1351 Maaß S, Daphi D, Lehmann A, Rillig MC (2017) Transport of microplastics by two collembolan  
1352 species. *Environ Pollut* 225:456–459. doi: 10.1016/j.envpol.2017.03.009

1353 Magni S, Binelli A, Pittura L, et al (2019) The fate of microplastics in an Italian Wastewater Treatment  
1354 Plant. *Sci Total Environ* 652:602–610. doi: 10.1016/j.scitotenv.2018.10.269

1355 Magnusson K, Norén F (2014) Screening of microplastic particles in and down-stream a wastewater  
1356 treatment plant. *IVL Swedish Environ Res Inst C* 55:22. doi: naturvardsverket-2226

1357 Mahon AM, O'Connell B, Healy MG, et al (2017) Microplastics in sewage sludge: Effects of treatment.  
1358 *Environ Sci Technol* 51:810–818. doi: 10.1021/acs.est.6b04048

1359 Mai L, Bao LJ, Shi L, et al (2018) A review of methods for measuring microplastics in aquatic  
1360 environments. *Environ Sci Pollut Res* 25:11319–11332. doi: 10.1007/s11356-018-1692-0

1361 Mani T, Hauk A, Walter U, Burkhardt-Holm P (2016) Microplastics profile along the Rhine River. *Sci*  
1362 *Rep* 5:17988. doi: 10.1038/srep17988

1363 Mason SA, Garneau D, Sutton R, et al (2016) Microplastic pollution is widely detected in US municipal  
1364 wastewater treatment plant effluent. *Environ Pollut* 218:1045–1054. doi:

1365 10.1016/j.envpol.2016.08.056

1366 McCormick A, Hoellein TJ, Mason SA, et al (2014) Microplastic is an abundant and distinct microbial

1367 habitat in an urban river. *Environ Sci Technol* 48:11863–11871. doi: 10.1021/es503610r

1368 McCormick AR, Hoellein TJ, London MG, et al (2016) Microplastic in surface waters of urban rivers:

1369 Concentration, sources, and associated bacterial assemblages. *Ecosphere* 7:. doi:

1370 10.1002/ecs2.1556

1371 Mercier A, Gravouil K, Aucher W, et al (2017) Fate of Eight Different Polymers under Uncontrolled

1372 Composting Conditions: Relationships between Deterioration, Biofilm Formation, and the

1373 Material Surface Properties. *Environ Sci Technol* 51:1988–1997. doi: 10.1021/acs.est.6b03530

1374 Michielssen MR, Michielssen ER, Ni J, Duhaime MB (2016) Fate of microplastics and other small

1375 anthropogenic litter (SAL) in wastewater treatment plants depends on unit processes employed.

1376 *Environ Sci Water Res Technol* 2:1064–1073. doi: 10.1039/C6EW00207B

1377 Miller RZ, Watts AJR, Winslow BO, et al (2017) Mountains to the sea: River study of plastic and non-

1378 plastic microfiber pollution in the northeast USA. *Mar Pollut Bull* 124:245–251. doi:

1379 10.1016/j.marpolbul.2017.07.028

1380 Mintenig SM, Int-Veen I, Löder MGJ, et al (2017) Identification of microplastic in effluents of waste

1381 water treatment plants using focal plane array-based micro-Fourier-transform infrared imaging.

1382 *Water Res* 108:365–372. doi: 10.1016/j.watres.2016.11.015

1383 Mintenig SM, Löder MGJ, Primpke S, Gerdts G (2019) Low numbers of microplastics detected in

1384 drinking water from ground water sources. *Sci Total Environ* 648:631–635. doi:

1385 10.1016/j.scitotenv.2018.08.178

1386 Mintenig SM, Bäuerlein PS, Koelmans AA, et al. (2018) Closing the gap between small and smaller:

1387 Towards a framework to analyse nano- and microplastic in aqueous environmental samples.

1388 *Environ Sci Nano* 5:1640-1649. doi: 10.1039/C8EN00186C

1389 Mohamed Nor NH, Koelmans AA (2019) Transfer of PCBs from Microplastics under Simulated Gut

1390 Fluid Conditions Is Biphaseic and Reversible. *Environ Sci Technol* 53:1874–1883. doi:

1391 10.1021/acs.est.8b05143

1392 Moore CJ, Lattin GL, Zellers AF (2011) Quantity and type of plastic debris flowing from two urban

1393 rivers to coastal waters and beaches of Southern California. *Rev Gestão Costeira Integr* 11:65–

1394 73. doi: 10.5894/rgci194

1395 Morritt D, Stefanoudis P V., Pearce D, et al (2014) Plastic in the Thames: A river runs through it. *Mar*

1396 *Pollut Bull* 78:196–200. doi: 10.1016/j.marpolbul.2013.10.035

1397 Mulder M (1998) Basic principle of membrane technology. Netherlands, Kluwer academic publisher.

1398 Murphy F, Ewins C, Carbonnier F, Quinn B (2016) Wastewater Treatment Works (WwTW) as a

1399 Source of Microplastics in the Aquatic Environment. *Environ Sci Technol* 50:5800–5808. doi:

1400 10.1021/acs.est.5b05416

1401 Napper IE, Bakir A, Rowland SJ, Thompson RC (2015) Characterisation, quantity and sorptive

1402 properties of microplastics extracted from cosmetics. *Mar Pollut Bull* 99:178–185. doi:

1403 10.1016/j.marpolbul.2015.07.029

1404 Napper IE, Thompson RC (2016) Release of synthetic microplastic plastic fibres from domestic

1405 washing machines: Effects of fabric type and washing conditions. *Mar Pollut Bull* 112:39–45. doi:

1406 10.1016/j.marpolbul.2016.09.025

1407 Nel HA, Dalu T, Wasserman RJ (2018) Sinks and sources: Assessing microplastic abundance in river

1408 sediment and deposit feeders in an Austral temperate urban river system. *Sci Total Environ*

1409 612:950–956. doi: 10.1016/j.scitotenv.2017.08.298

1410 Ng EL, Huerta Lwanga E, Eldridge SM, et al (2018) An overview of microplastic and nanoplastic

1411 pollution in agroecosystems. *Sci Total Environ* 627:1377–1388. doi:

1412 10.1016/j.scitotenv.2018.01.341

1413 Nizzetto L, Futter M, Langaas S (2016) Are Agricultural Soils Dumps for Microplastics of Urban

1414 Origin? *Environ Sci Technol* 50:10777–10779. doi: 10.1021/acs.est.6b04140

1415 Peng G, Xu P, Zhu B, et al (2018) Microplastics in freshwater river sediments in Shanghai, China: A

1416 case study of risk assessment in mega-cities. *Environ Pollut* 234:448–456. doi:

1417 10.1016/j.envpol.2017.11.034

1418 Picó Y, Barceló D (2019) Analysis and Prevention of Microplastics Pollution in Water: Current

1419 Perspectives and Future Directions. *ACS Omega* 4:6709–6719. doi:

1420 10.1021/acsomega.9b00222

1421 Piehl S, Leibner A, Löder MGJ, et al (2018) Identification and quantification of macro- and

1422 microplastics on an agricultural farmland. *Sci Rep* 8:17950. doi: 10.1038/s41598-018-36172-y

1423 Pirc U, Vidmar M, Mozer A, Kržan A (2016) Emissions of microplastic fibers from microfiber fleece

1424 during domestic washing. *Environ Sci Pollut Res* 23:22206–22211. doi: 10.1007/s11356-016-

1425 7703-0

- 1426 PlasticsEurope (2018) Plastics – the Facts.  
 1427 [https://www.plasticseurope.org/application/files/6315/4510/9658/Plastics\\_the\\_facts\\_2018\\_AF\\_w](https://www.plasticseurope.org/application/files/6315/4510/9658/Plastics_the_facts_2018_AF_w)  
 1428 [eb.pdf](#)
- 1429 Prata JC, da Costa JP, Lopes I, et al (2019) Effects of microplastics on microalgae populations: A  
 1430 critical review. *Sci Total Environ* 665:400–405. doi: 10.1016/j.scitotenv.2019.02.132
- 1431 Ramaswamy V, Sharma HR (2011) RESEARCH ARTICLE PLASTIC BAGS – THREAT TO  
 1432 ENVIRONMENT AND CATTLE HEALTH : A RETROSPECTIVE STUDY FROM GONDAR CITY  
 1433 OF ETHIOPIA. 2:7–12
- 1434 Ramos L, Berenstein G, Hughes EA, et al (2015) Polyethylene film incorporation into the horticultural  
 1435 soil of small periurban production units in Argentina. *Sci Total Environ* 523:74–81. doi:  
 1436 10.1016/j.scitotenv.2015.03.142
- 1437 Rech S, Macaya-Caquilpán V, Pantoja JF, et al (2014) Rivers as a source of marine litter - A study  
 1438 from the SE Pacific. *Mar Pollut Bull* 82:66–75. doi: 10.1016/j.marpolbul.2014.03.019
- 1439 Redondo Hasselerharm PE, Falahudin D, Peeters E, Koelmans AA (2018) Microplastic effect  
 1440 thresholds for freshwater benthic macroinvertebrates. *Environ Sci Technol* acs.est.7b05367. doi:  
 1441 10.1021/acs.est.7b05367
- 1442 Rillig MC, Ziersch L, Hempel S (2017) Microplastic transport in soil by earthworms. *Sci Rep* 7:1362.  
 1443 doi: 10.1038/s41598-017-01594-7
- 1444 Rochman CM, Kross SM, Armstrong JB, et al (2015) Scientific Evidence Supports a Ban on  
 1445 Microbeads. *Environ Sci Technol* 49:10759–10761. doi: 10.1021/acs.est.5b03909
- 1446 Rochman CM, Kurobe T, Flores I, Teh SJ (2014) Early warning signs of endocrine disruption in adult  
 1447 fish from the ingestion of polyethylene with and without sorbed chemical pollutants from the  
 1448 marine environment. *Sci Total Environ* 493:656–661. doi: 10.1016/j.scitotenv.2014.06.051
- 1449 Rodrigues MO, Abrantes N, Gonçalves FJM, et al (2018) Spatial and temporal distribution of  
 1450 microplastics in water and sediments of a freshwater system (Antuã River, Portugal). *Sci Total*  
 1451 *Environ* 633:1549–1559. doi: 10.1016/j.scitotenv.2018.03.233
- 1452 Sato T, Qadir M, Yamamoto S, et al (2013) Global, regional, and country level need for data on  
 1453 wastewater generation, treatment, and use. *Agric Water Manag* 130:1–13. doi:  
 1454 10.1016/j.agwat.2013.08.007
- 1455 SAPEA, Science Advice for Policy by European Academies. (2018). A Scientific Perspective on  
 1456 Microplastics in Nature and Society. Berlin:SAPEA. <https://doi.org/10.26356/microplastics>
- 1457 Scarascia-Mugnozza G, Sica C, Russo G (2012) Plastic Materials in European Agriculture: Actual Use  
 1458 and Perspectives. *J Agric Eng* 42:15. doi: 10.4081/jae.2011.3.15
- 1459 Scheurer M, Bigalke M (2018) Microplastics in Swiss Floodplain Soils. *Environ Sci Technol* 52:3591–  
 1460 3598. doi: 10.1021/acs.est.7b06003
- 1461 Schmidt C, Krauth T, Wagner S (2017a) Export of Plastic Debris by Rivers into the Sea. *Environ Sci*  
 1462 *Technol* 51:acs.est.7b06377. doi: 10.1021/acs.est.7b06377
- 1463 Schmidt C, Krauth T, Wagner S (2017b) Export of Plastic Debris by Rivers into the Sea. *Environ Sci*  
 1464 *Technol* 51:12246–12253. doi: 10.1021/acs.est.7b02368
- 1465 Schür C, Rist S, Baun A, et al (2019) WHEN Fluorescence is not a particle: The tissue translocation of  
 1466 microplastics in *Daphnia magna* seems an artifact. *Environ Toxicol Chem* 0: doi:  
 1467 10.1002/etc.4436
- 1468 Schuyler Q, Hardesty BD, Wilcox C, Townsend K (2012) To eat or not to eat? debris selectivity by  
 1469 marine turtles. *PLoS One* 7:. doi: 10.1371/journal.pone.0040884
- 1470 Shruti VC, Jonathan MP, Rodriguez-Espinosa PF, Rodríguez-González F (2019) Microplastics in  
 1471 freshwater sediments of Atoyac River basin, Puebla City, Mexico. *Sci Total Environ* 654:154–  
 1472 163. doi: 10.1016/j.scitotenv.2018.11.054
- 1473 Sighicelli M, Pietrelli L, Lecce F, et al (2018) Microplastic pollution in the surface waters of Italian  
 1474 Subalpine Lakes. *Environ Pollut* 236:645–651. doi: 10.1016/j.envpol.2018.02.008
- 1475 Simon M, van Alst N, Vollertsen J (2018) Quantification of microplastic mass and removal rates at  
 1476 wastewater treatment plants applying Focal Plane Array (FPA)-based Fourier Transform Infrared  
 1477 (FT-IR) imaging. *Water Res* 142:1–9. doi: 10.1016/j.watres.2018.05.019
- 1478 Stenmarck Å, Belleza EL, Fråne A, et al (2017) Hazardous substances in plastics  
 1479 Su L, Xue Y, Li L, et al (2016) Microplastics in Taihu Lake, China. *Environ Pollut* 216:711–719. doi:  
 1480 10.1016/j.envpol.2016.06.036
- 1481 Sundt P, Schulze P-E, Syversen F (2014) Sources of microplastic- pollution to the marine environment  
 1482 Project report. *Nor Environ Agency*. doi: M-321|2015
- 1483 Talvitie J, Heinonen M, Pääkkönen JP, et al (2015) Do wastewater treatment plants act as a potential  
 1484 point source of microplastics? Preliminary study in the coastal Gulf of Finland, Baltic Sea. *Water*  
 1485 *Sci Technol* 72:1495–1504. doi: 10.2166/wst.2015.360
- 1486 Talvitie J, Mikola A, Koistinen A, Setälä O (2017a) Solutions to microplastic pollution – Removal of

1487 microplastics from wastewater effluent with advanced wastewater treatment technologies. *Water*

1488 *Res* 123:401–407. doi: 10.1016/j.watres.2017.07.005

1489 Talvitie J, Mikola A, Setälä O, et al (2017b) How well is microlitter purified from wastewater? – A

1490 detailed study on the stepwise removal of microlitter in a tertiary level wastewater treatment

1491 plant. *Water Res* 109:164–172. doi: 10.1016/j.watres.2016.11.046

1492 Tan X, Xubiao Yu, Cai L, et al (2019) Microplastics and associated PAHs in surface water from the

1493 Feilaixia Reservoir in the Beijiing River, China. *Chemosphere* 221:834–840. doi:

1494 10.1016/j.chemosphere.2019.01.022

1495 Teuten EL, Saquing JM, Knappe DRU, et al (2009) Transport and release of chemicals from plastics

1496 to the environment and to wildlife. *Philos Trans R Soc B Biol Sci* 364:2027–2045. doi:

1497 10.1098/rstb.2008.0284

1498 Tibbetts J, Krause S, Lynch I, Smith GHS (2018) Abundance, distribution, and drivers of microplastic

1499 contamination in urban river environments. *Water (Switzerland)* 10:. doi: 10.3390/w10111597

1500 Unice KM, Kreider ML, Panko JM (2013) Comparison of tire and road wear particle concentrations in

1501 sediment for watersheds in France, Japan, and the United States by quantitative pyrolysis

1502 GC/MS analysis. *Environ Sci Technol* 47:8138–8147. doi: 10.1021/es400871j

1503 US Congress 2015. Microbead-Free Waters Act of 2015. Public Law 114 – 231 114th Congress An

1504 Act. Public Law 114-144. <https://www.congress.gov/bill/114th-congress/house-bill/1321/text>.

1505 Accessed 26 May 2019.

1506 van Emmerik T, Kieu-Le T-C, Loozen M, et al (2018) A Methodology to Characterize Riverine

1507 Macroplastic Emission Into the Ocean. *Front Mar Sci* 5:1–11. doi: 10.3389/fmars.2018.00372

1508 Van Franeker JA, Blaize C, Danielsen J, et al (2011) Monitoring plastic ingestion by the northern

1509 fulmar *Fulmarus glacialis* in the North Sea. *Environ Pollut* 159:2609–2615. doi:

1510 10.1016/j.envpol.2011.06.008

1511 Vaughan R, Turner SD, Rose NL (2017) Microplastics in the sediments of a UK urban lake. *Environ*

1512 *Pollut* 229:10–18. doi: 10.1016/j.envpol.2017.05.057

1513 Verma R, Vinoda KS, Papireddy M, Gowda ANS (2016) Toxic Pollutants from Plastic Waste- A

1514 Review. *Procedia Environ Sci* 35:701–708. doi: 10.1016/j.proenv.2016.07.069

1515 Vermaire JC, Pomeroy C, Herczegh SM, et al (2017) Microplastic abundance and distribution in the

1516 open water and sediment of the Ottawa River, Canada, and its tributaries. *Facets* 2:301–314.

1517 doi: 10.1139/facets-2016-0070

1518 Vianello A, Boldrin A, Guerriero P, et al (2013) Microplastic particles in sediments of Lagoon of

1519 Venice, Italy: First observations on occurrence, spatial patterns and identification. *Estuar Coast*

1520 *Shelf Sci* 130:54–61. doi: 10.1016/j.ecss.2013.03.022

1521 Vollertsen J, Hansen AA (ed) (2017) Microplastic in Danish wastewater: Sources, occurrences and

1522 fate. Danish Environmental Protection Agency

1523 Wang J, Peng J, Tan Z, et al (2017a) Microplastics in the surface sediments from the Beijiing River

1524 littoral zone: Composition, abundance, surface textures and interaction with heavy metals.

1525 *Chemosphere* 171:248–258. doi: 10.1016/j.chemosphere.2016.12.074

1526 Wang W, Ndungu AW, Li Z, Wang J (2017b) Microplastics pollution in inland freshwaters of China: A

1527 case study in urban surface waters of Wuhan, China. *Sci Total Environ* 575:1369–1374. doi:

1528 10.1016/j.scitotenv.2016.09.213

1529 Wang W, Yuan W, Chen Y, Wang J (2018) Microplastics in surface waters of Dongting Lake and Hong

1530 Lake, China. *Sci Total Environ* 633:539–545. doi: 10.1016/j.scitotenv.2018.03.211

1531 Weithmann N, Möller JN, Löder MGJ, et al (2018) Organic fertilizer as a vehicle for the entry of

1532 microplastic into the environment. *Sci Adv* 4:1–8. doi: 10.1126/sciadv.aap8060

1533 Wen X, Du C, Xu P, et al (2018) Microplastic pollution in surface sediments of urban water areas in

1534 Changsha, China: Abundance, composition, surface textures. *Mar Pollut Bull* 136:414–423. doi:

1535 10.1016/j.marpolbul.2018.09.043

1536 Wetherbee GA, Baldwin AK, Ranville JF (2019) It is raining plastic. Reston, VA

1537 Williams AT, Simmons SL (1999) Sources of riverine litter: The river Taff, South Wales, UK. *Water Air*

1538 *Soil Pollut* 112:197–216. doi: 10.1023/A:1005000724803

1539 Williams AT, Simmons SL (1996) The degradation of plastic litter in rivers: Implications for beaches. *J*

1540 *Coast Conserv* 2:63–72. doi: 10.1007/BF02743038

1541 Wolff S, Kerpen J, Prediger J, et al (2018) Determination of the Microplastics Emission in the Effluent

1542 of a Municipal Waste Water Treatment Plant using Raman Microspectroscopy. *Water Res X*

1543 2:100014. doi: 10.1016/J.WROA.2018.100014

1544 Wright SL, Thompson RC, Galloway TS (2013) The physical impacts of microplastics on marine

1545 organisms: A review. *Environ Pollut* 178:483–492. doi: 10.1016/j.envpol.2013.02.031

1546 Xanthos D, Walker TR (2017) International policies to reduce plastic marine pollution from single-use

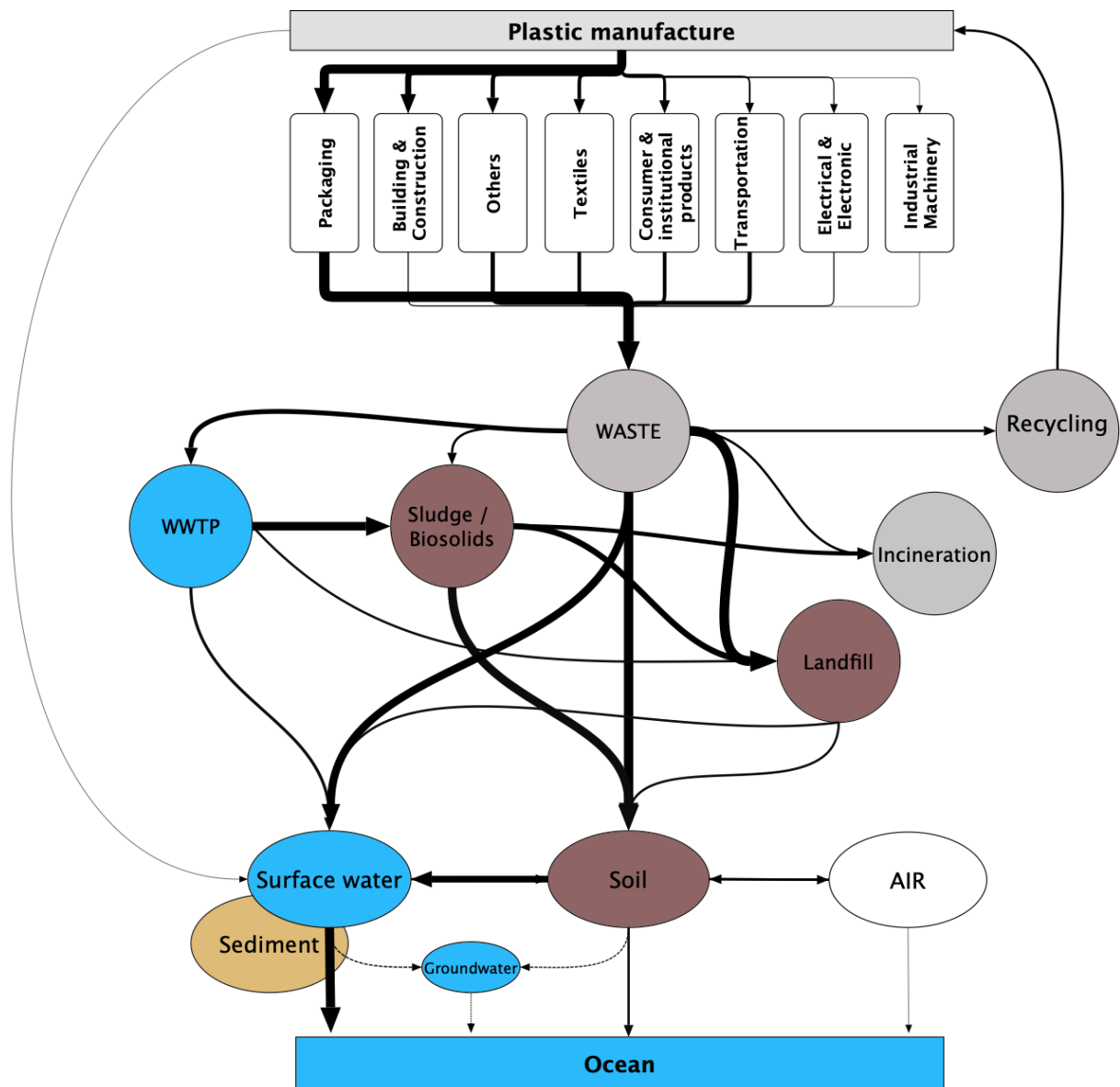
1547 plastics (plastic bags and microbeads): A review. *Mar Pollut Bull* 118:17–26. doi:

1548 10.1016/j.marpolbul.2017.02.048  
1549 Xiong X, Wu C, Elser JJ, et al (2019) Occurrence and fate of microplastic debris in middle and lower  
1550 reaches of the Yangtze River – From inland to the sea. *Sci Total Environ* 659:66–73. doi:  
1551 10.1016/j.scitotenv.2018.12.313  
1552 Yan M, Nie H, Xu K, et al (2019) Microplastic abundance, distribution and composition in the Pearl  
1553 River along Guangzhou city and Pearl River estuary, China. *Chemosphere* 217:879–886. doi:  
1554 10.1016/j.chemosphere.2018.11.093  
1555 Yang S-S, Brandon AM, Andrew Flanagan JC, et al (2018) Biodegradation of polystyrene wastes in  
1556 yellow mealworms (larvae of *Tenebrio molitor* Linnaeus): Factors affecting biodegradation rates  
1557 and the ability of polystyrene-fed larvae to complete their life cycle. *Chemosphere* 191:979–989.  
1558 doi: <https://doi.org/10.1016/j.chemosphere.2017.10.117>  
1559 Ye S, Andrady AL (1991) Fouling of Floating Plastic Debris Under Biscayne Bay Exposure Conditions.  
1560 *Mar Pollut Bull* 22:  
1561 Yoshida S, Hiraga K, Takanaha T, et al (2016) A bacterium that degrades and assimilates  
1562 poly(ethyleneterephthalate). *Science* (80- ) 351:1196–1199. doi: 10.1126/science.aad6359  
1563 Yuan W, Liu X, Wang W, et al (2019) Microplastic abundance, distribution and composition in water,  
1564 sediments, and wild fish from Poyang Lake, China. *Ecotoxicol Environ Saf* 170:180–187. doi:  
1565 10.1016/j.ecoenv.2018.11.126  
1566 Zbyszewski M, Corcoran PL (2011) Distribution and degradation of fresh water plastic particles along  
1567 the beaches of Lake Huron, Canada. *Water Air Soil Pollut* 220:365–372. doi: 10.1007/s11270-  
1568 011-0760-6  
1569 Zbyszewski M, Corcoran PL, Hockin A (2014) Comparison of the distribution and degradation of  
1570 plastic debris along shorelines of the Great Lakes, North America. *J Great Lakes Res* 40:288–  
1571 299. doi: 10.1016/j.jglr.2014.02.012  
1572 Zettler ER, Mincer TJ, Amaral-Zettler LA (2013) Life in the “Plastisphere”: Microbial Communities on  
1573 Plastic Marine Debris. *Environ Sci Technol* 47:7137–7146. doi: 10.1021/es401288x  
1574 Zhang GS, Liu YF (2018) The distribution of microplastics in soil aggregate fractions in southwestern  
1575 China. *Sci Total Environ* 642:12–20. doi: 10.1016/j.scitotenv.2018.06.004  
1576 Zhang K, Gong W, Lv J, et al (2015) Accumulation of floating microplastics behind the Three Gorges  
1577 Dam. *Environ Pollut* 204:117–123. doi: 10.1016/j.envpol.2015.04.023  
1578 Zhang K, Su J, Xiong X, et al (2016) Microplastic pollution of lakeshore sediments from remote lakes  
1579 in Tibet plateau, China. *Environ Pollut* 219:450–455. doi: 10.1016/j.envpol.2016.05.048  
1580 Zhang S, Yang X, Gertsen H, et al (2018) A simple method for the extraction and identification of light  
1581 density microplastics from soil. *Sci Total Environ* 616–617:1056–1065. doi:  
1582 10.1016/j.scitotenv.2017.10.213  
1583 Zhao S, Zhu L, Li D (2016) Microscopic anthropogenic litter in terrestrial birds from Shanghai, China:  
1584 Not only plastics but also natural fibers. *Sci Total Environ* 550:1110–1115. doi:  
1585 10.1016/j.scitotenv.2016.01.112  
1586 Ziajahromi S, Neale PA, Leusch FDLL (2016) Wastewater treatment plant effluent as a source of  
1587 microplastics: review of the fate, chemical interactions and potential risks to aquatic organisms.  
1588 *Water Sci Technol* 74:2253–2269. doi: 10.2166/wst.2016.414  
1589 Ziajahromi S, Neale PA, Rintoul L, Leusch FDL (2017) Wastewater treatment plants as a pathway for  
1590 microplastics: Development of a new approach to sample wastewater-based microplastics.  
1591 *Water Res* 112:93–99. doi: 10.1016/j.watres.2017.01.042  
1592 Zubris KA V., Richards BK (2005a) Synthetic fibers as an indicator of land application of sludge.  
1593 *Environ Pollut* 138:201–211. doi: 10.1016/j.envpol.2005.04.013  
1594 Zubris KA V, Richards BK (2005b) Synthetic fibers as an indicator of land application of sludge.  
1595 *Environ Pollut* 138:201–211. doi: 10.1016/j.envpol.2005.04.013  
1596 Zylstra ER (2013) Accumulation of wind-dispersed trash in desert environments. *J Arid Environ* 89:13–  
1597 15. doi: 10.1016/j.jaridenv.2012.10.004  
1598  
1599  
1600  
1601 Table 1. Percentage of EU population connected to WWTPs in 2015 (EEA, 2019).

No treatment or no connection	Primary	Secondary	Tertiary
-------------------------------------	---------	-----------	----------

with sewerage				
Northern	15.1	5.6	2.3	77
Central	3.4	0	16.5	80.1
Southern	23	2.2	21.3	53.4
Eastern	26	0.2	13.6	60.6
South-Eastern	40	16.7	22.8	20.6
<b>Weighted average respect to population</b>	<b>13</b>	<b>2</b>	<b>18</b>	<b>67</b>

1602

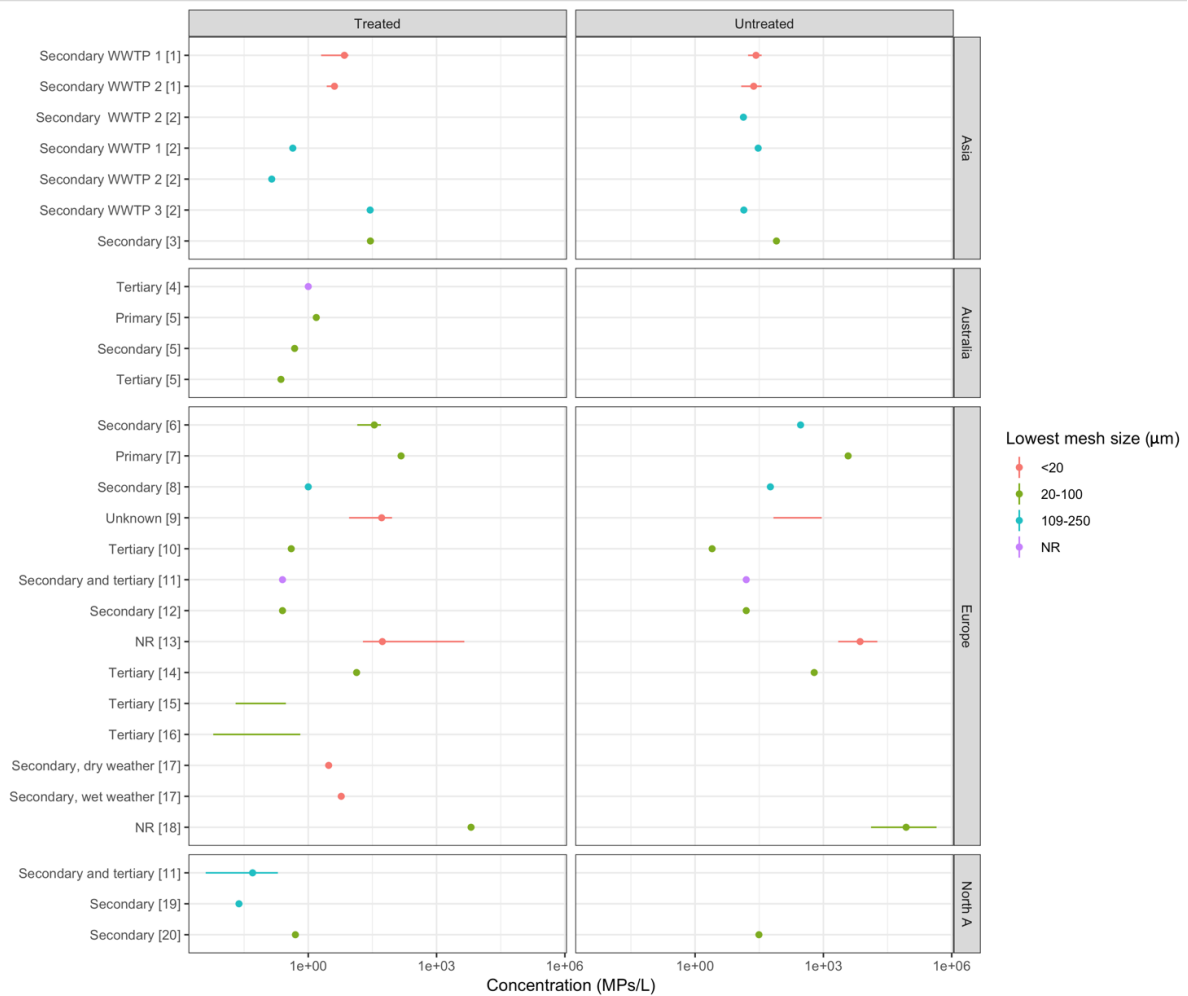


1603  
1604  
1605  
1606

Fig. 1. Production and pathways of plastics into the different environmental compartments. Thickness of the different arrows are related to the relevance of the different mass flows. The relevance of the different plastic

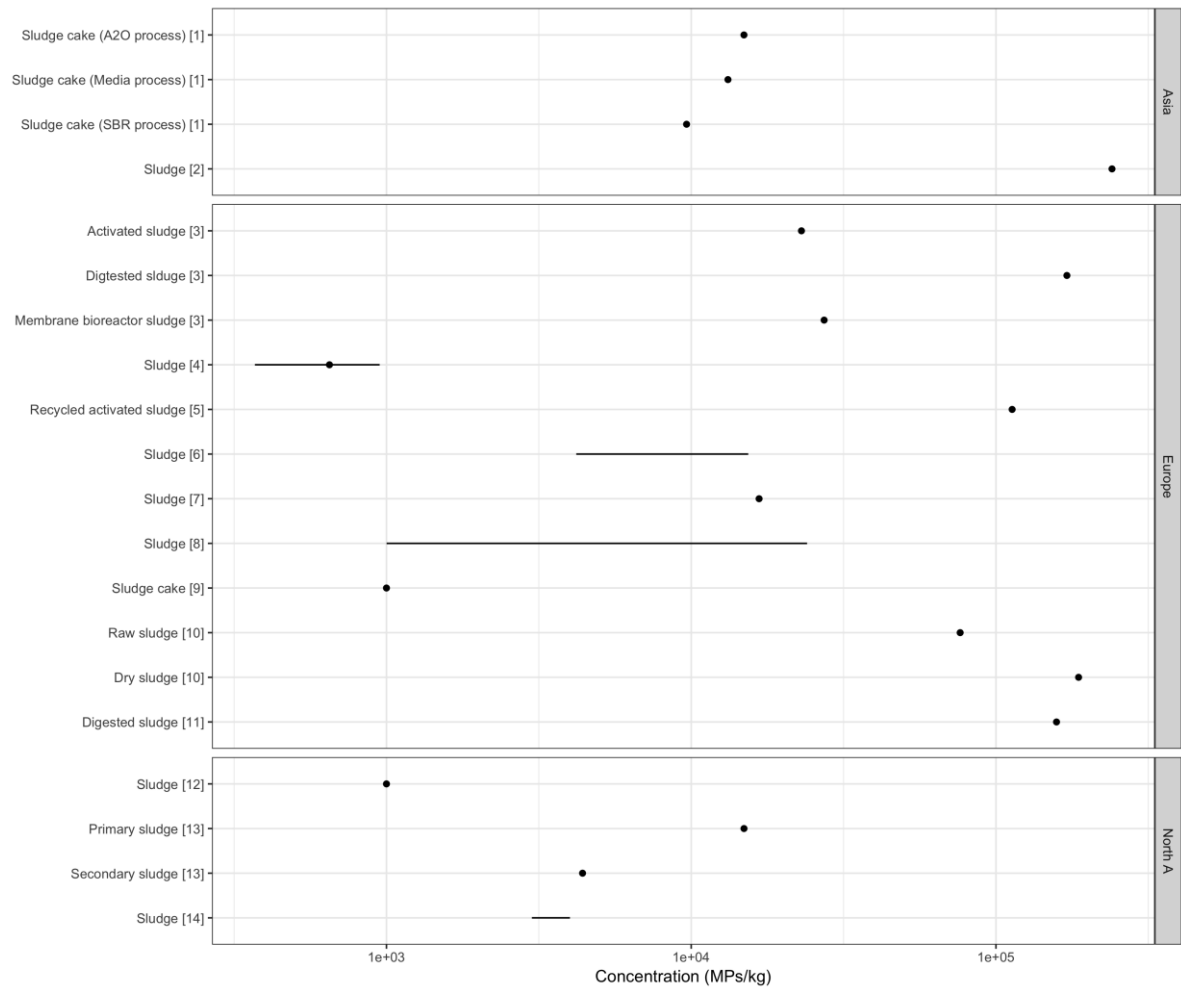
1607  
1608  
1609

source mass flows is based on Geyer et al. (2017), while the relevance of the environmental flows is based on the reviewed literature or assumptions. Dashed lines indicate yet completely unexplored pathways with unknown relevance.



1610  
1611

1612 *Fig. 2. MP concentrations in untreated and treated wastewaters (MPs/L) from WWTPs with different treatment*  
 1613 *types. NR= not reported. [1] Gündoğdu et al. (2018) [2] Lee and Kim 2018 (2018) [3] Liu et al. (2019b) [4] Browne*  
 1614 *et al. (2011) [5] Ziajahromi et al. (2017) [6] Dris et al. (2015) [7] Helcom (2014) [8] Lares et al. (2018) [9] Leslie et*  
 1615 *al. (2017) [10] Magni et al. (2019) [11] Mason et al. (2016) [12] Murphy et al. (2016) [13] Simon et al. (2018) [14]*  
 1616 *Talvitie et al. (2015) [15] Talvitie et al. (2017a) [16] Talvitie et al. (2017b) [17] Wolff et al. (2018) [18] Vollertsen*  
 1617 *and Hansen (2017) [19] Dyachenko et al, 2017 (2017) [20] Gies et al. (2018).*



1618

1619

1620

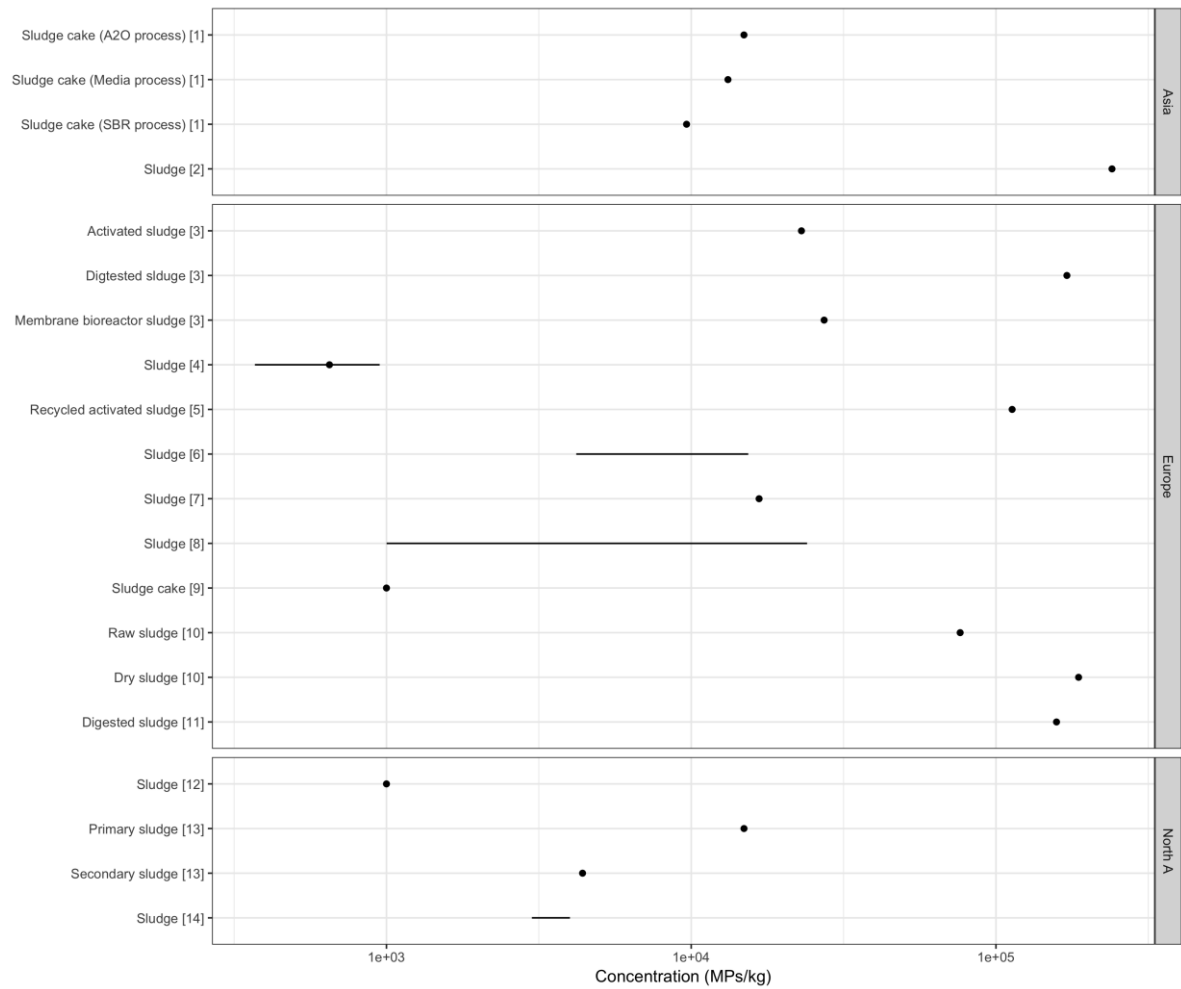
1621

1622

1623

Fig. 3. MP concentrations in sludge samples (MPs/kg dw). North A = North America; A2O = anaerobic-anoxic-aerobic; SBR= sequence batch reactor [1] Lee and Kim (2018) [2] Liu et al. (2019b) [3] Lares et al. 2018 (Lares et al. 2018) [4] Leslie et al. (2017) [5] Magni et al. (2019) [6] Mahon et al. (2017) [7] Magnusson and Norén (2014) [8] Mintenig et al. (2017) [9] Murphy et al. (2016) [10] Talvitie et al. (2017b) [11] Vollertsen and Hansen (2017) [12] Carr et al. (2016) [13] Gies et al. (2018) [14] Zubris and Richards (2005)

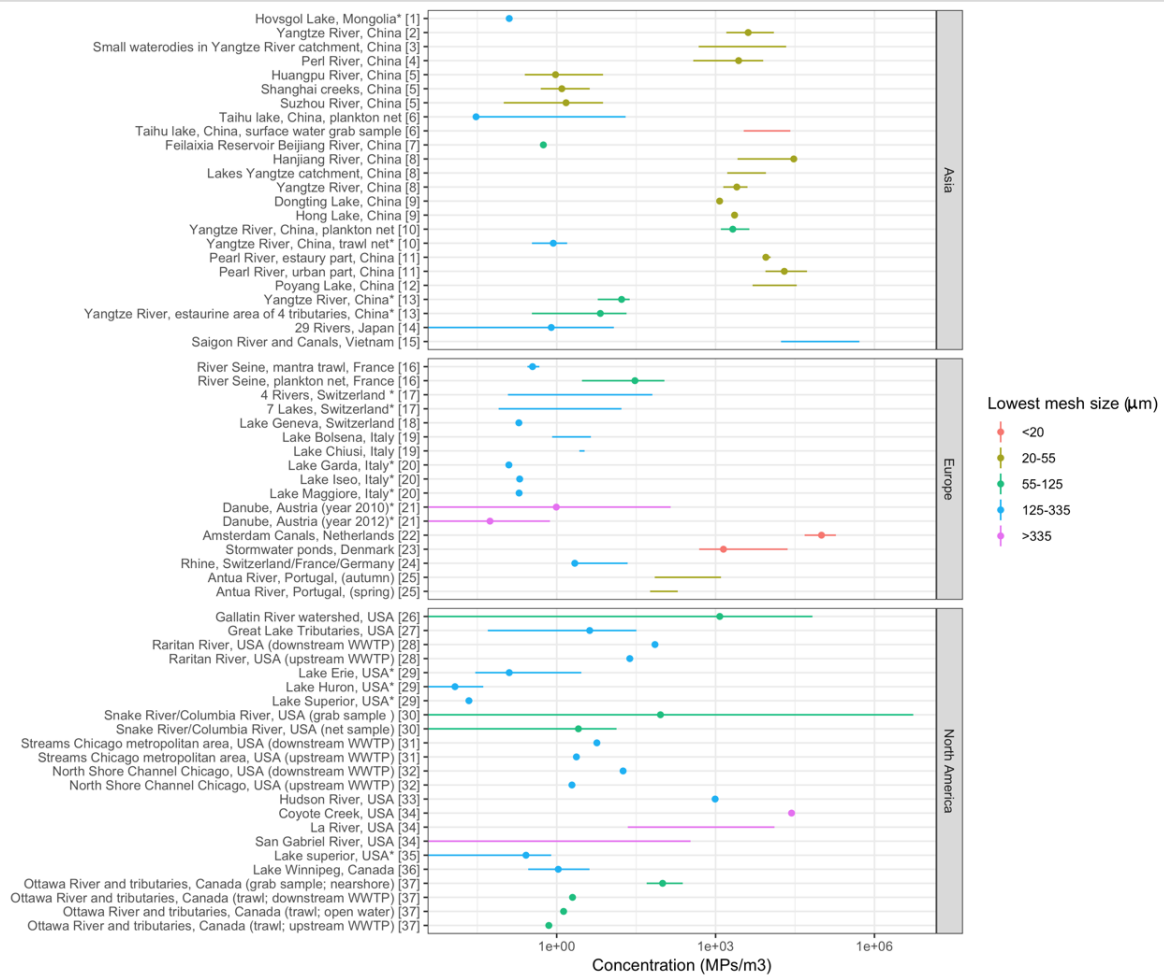




1624

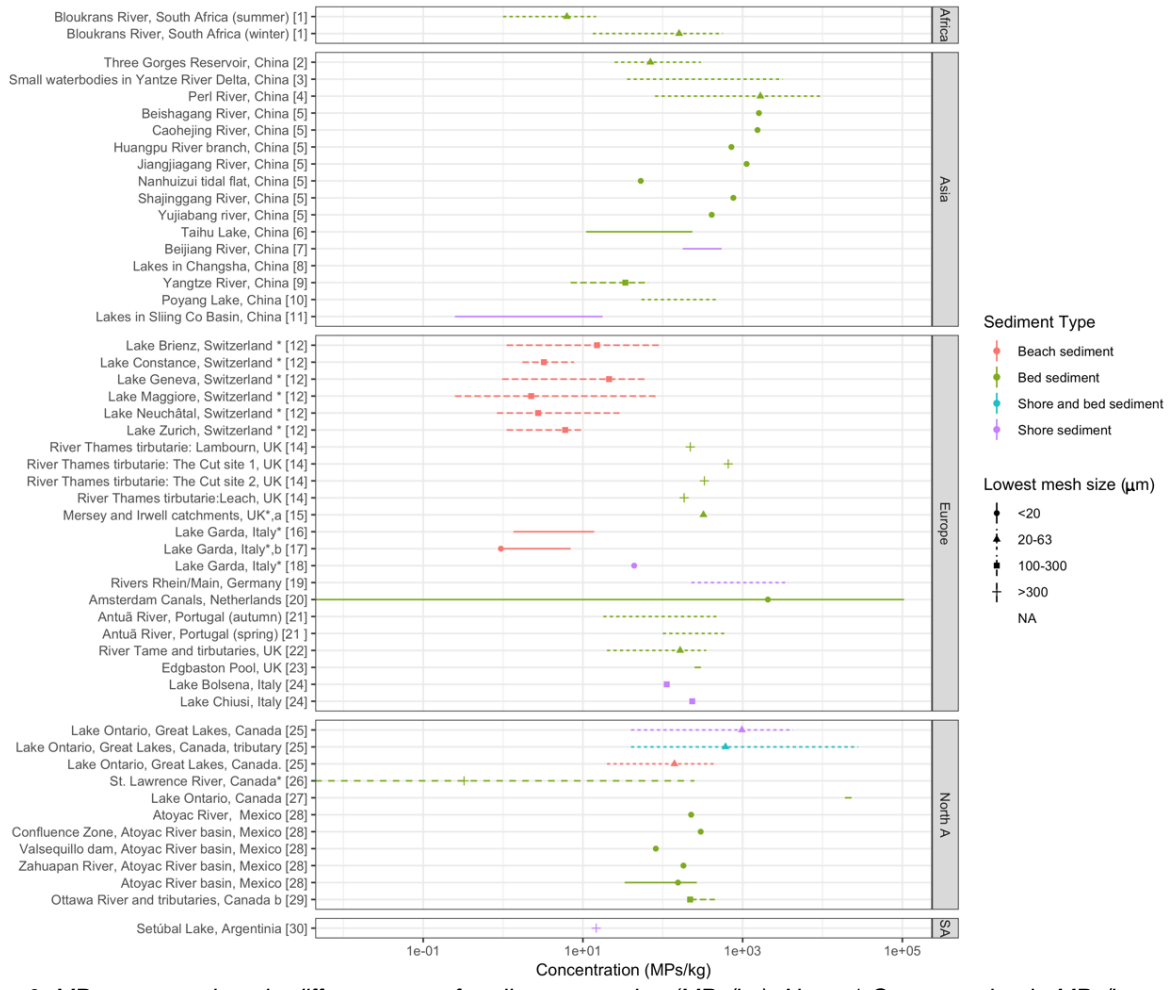
1625  
 1626  
 1627  
 1628  
 1629

Fig. 4. MP concentrations in different soil samples (MPs/kg dw). AUS = Australia; C.A. = Central America; N.A = North America. [1] Fuller and Gautam (2016) [2] Zhang et al. (2018) [3] Zhang and Liu (2018) [4] Liu et al. (2018) [5] Huerta Lwanga et al. (2017b) [6] Corradini et al. (2019b) [7] Vollertsen and Hansen (2017) [8] Piehl et al. (2018) [9] Scheurer and Bigalke (2018) [10] Zubris and Richards (2005).



1630  
1631  
1632  
1633  
1634  
1635  
1636  
1637  
1638  
1639  
1640  
1641

Fig. 5. MP concentrations in surface water samples ( $\text{MPs}/\text{m}^3$ ). \* Concentration in  $\text{MPs}/\text{m}^3$  was estimated by dividing the reported concentration in particles per area by the height of the net used for sampling. [1] Free et al. (2014) [2] Di and Wang (2018) [3] Hu et al. (2018) [4] Lin et al. (2018) [5] Luo et al. (2018) [6] Su et al. (2016) [7] Tan et al. (2019) [8] Wang et al. (2017a) [9] Wang et al. (2018) [10] Xiong et al. (2019) [11] Yan et al. (2019) [12] Yuan et al. (2019) [13] Zhang et al. (2015) [14] Kataoka et al. (2019) [15] Lahens et al. (2018) [16] Dris et al. (2015) [17] Faure et al. (2015) [18] Faure et al. (2012) [19] Fischer et al. (2016) [20] Sighicelli et al. (2018) [21] Lechner et al. (2014) [22] Leslie et al. (2017) [23] (Liu et al. 2019a) [24] Mani et al. (2016) [25] Rodrigues et al. (2018) [26] Barrows et al. (2018) [27] (Baldwin et al. 2016) [28] Estahbanati and Fahrenfeld (2016) [29] Eriksen et al. (2013) [30] Kapp and Yeatman (2018) [31] McCormick et al. (2016) [32] McCormick et al. (2014) [33] Miller et al. (2017) [34] Moore et al. (2011) [35] Hendrickson et al. (2018) [36] Anderson et al. (2017) [37] Vermaire et al. (2017).



1642  
 1643  
 1644  
 1645  
 1646  
 1647  
 1648  
 1649  
 1650  
 1651

Fig. 6. MP concentrations in different type of sediment samples (MPs/kg). Notes: \* Concentration in MPs/kg was estimated by using the sample depth and assuming a density of 1.6 g/cm<sup>3</sup> for the sediment <sup>a</sup> maximum value is shown; <sup>b</sup> no lower value reported. North A. = North America; SA = South America. [1] Nel et al. (2018) [2] Di and Wang (2018) [3] Hu et al. (2018) [4] Lin et al. (2018) [5] Peng et al. (2018) [6] Su et al. (2016) [7] Wang et al. (2017a) [8] Wen et al. (2018) [9] Xiong et al. (2019) [10] Yuan et al. (2019) [11] Zhang et al. (2016) [12] Faure et al. (2015) [14] Horton et al. (2017) [15] Hurley et al. (2018a) [16] Imhof et al. (2013) [17] Imhof et al. (2016) [18] Imhof et al. (2018) [19] Klein et al. (2015) [20] Leslie et al. (2017) [21] Rodrigues et al. (2018) [22] Tibbetts et al. (2018) [23] Vaughan et al. (2017) [24] Fischer et al. (2016) [25] Ballent et al. (2016) [26] Castañeda et al. (2014) [27] Corcoran et al. (2015) [28] Shruti et al. (2019) [29] Vermaire et al. (2017) [30] (Blettler et al. 2017

1652  
1653  
1654  
1655  
1656  
1657

**Supplemental material**

Table S1. Concentration and removal of MPs in municipal WWTPs for different treatment types; **bold** numbers represent the median concentration instead of the mean concentration; NR = not reported.

Treatment type (no. of WWTP)	Location	Mean concentration in MP/L <sup>a</sup> ± SD or (minimum - maximum)		Removal rate (%)	Lowest mesh sizes or lowest size limit (µm)	Identification method	Dominant shapes in effluent	Dominant polymer compositio n in effluent	Referenc e
		Influent	Effluent						
Tertiary (1)	Australia	NR	1	NR	NR	Visual and FTIR	Fibres	Polyester, Acrylic	[1]
Primary (1)	Australia	NR	1.54	NR	25	Visual and FTIR	Fibres	PET	[2]
Secondary (1)		NR	0.48	NR					
Tertiary (1)		NR	0.28	NR					
Secondary (3)	Korea	29.8	0.435	98-99	109	Visual and FTIR	Fibres, black particles <sup>c</sup>	NR	[3]
		13.5	0.14						
		13.8	0.28						
Secondary (2)	China	79.9 ± 9.3	28.4 ± 7	64.4	47	Visual and Raman	Fragments	Nylon	[4]
Secondary (2)	Turkey	26.6 (17.3 - 36)	7 (2 - 8.7)	73	5	Visual and µ- Raman	Fibres	PET	[5]
		23.4 (12 - 36)	4.1 (2.7 - 4.7)	79					
Secondary (1)	France	293 (260 - 320)	35 (14 - 50)	NR	100	Visual	Fibres	NR	[6]
Primary (1)	Russia	3787 <sup>b</sup>	148 <sup>b</sup>	96	20	Visual	Black particles		[7]
Secondary (1)	Finland	57.6 ± 12.4	<b>1.0 ± 0.4</b>	98.3	250	Visual and FTIR/Raman	Fibre	PES	[8]
NR (7)	The Netherlands	68 - 910 <sup>d</sup>	51 - 81 <sup>d</sup>	72	10	Visual and FTIR	Fibres		[9]
Secondary (1)	Sweden	15.1 ± 0.89	0.00825 ± 0.00085	99.9	300	Visual and FTIR	Fibres		[10]
Tertiary (1)	Italy	2.5 ± 0.3	0.4 ± 0.1	84	63	Visual and FTIR	Lines and Films	PES	[11]

Secondary and tertiary (12)	Germany	NR	> 500 µm: 0-0.05; <500 µm: 0.01-9	NR	20	Visual and FTIR	Fibres	PES	[12]
Secondary (1)	Scotland	15.7 ± 5.23	0.25 ± 0.04	98.4	65	Visual and FTIR	Flakes	PES, Polyamine	[13]
Secondary (10)	Denmark	7,771 ± 4,283 <sup>k</sup> (2,223 - 18,285), <b>7216</b> ; 341 ± 323.9 µg/L <sup>k</sup> (61 - 1189 µg/L), <b>250 µg/L</b>	114.3 ± 133.5 <sup>k</sup> (min-max: 19 - 447) <b>54</b> ; 4.3 ± 4.25 µg/L <sup>k</sup> (0.5-11.0 µg/L), <b>3.7 µg/L</b>	99.3; 98.3 <sup>e</sup>	10 <sup>f</sup>	FTIR and infrared map	Particles	PE and Polyester	[14]
Tertiary (1)	Finland	180 fibres; 430 particles	4.9 ± 1.4 fibres; 8.6 ± 2.5 particles	NR	20	Visual	Particles	NR	[15]
Tertiary (4) <sup>g</sup>	Finland	NR	0.02-0.3	NR	20	Visual and FTIR	NR	PES	[16]
Tertiary (1)	Finland	NR	0.006-0.651 <sup>b</sup>	> 99	20	Visual and FTIR	Fragments	Cotton, Polyester <sup>h</sup>	[17]
Secondary (1)	Germany	NR	5.9 (wet weather); 3 (dry weather)		10	Raman	Fragments	PET	[18]
NR (10)	Denmark	127,000; <b>86,000</b> ; 8,000 µg/L; <b>5900 µg/L</b>	5,800, <b>6,400</b> ; 34 µg/L, <b>16 µg/L</b>	99.7%	20	Visual and FTIR	NR	Nylon	[19]
Tertiary (1)	USA	1	0.0009	99.9	150	Visual and FTIR	NR	NR	[20]
Secondary (1)	USA	NR	0.024 (24h sample); 0.17 (2h peak flow event)	NR	125	Visual and FTIR	Fragments	NR	[21]
Secondary and tertiary (17)	USA	NA	0.05 ± 0.024 (0.004-0.195)	NR	125	Visual	Fibres	NR	[22]
Secondary (1)	USA	NA	5.9 <sup>b</sup>	95.6	20	Visual	Fibres	NR	[23]
Tertiary (1)		NA	2.6 <sup>b</sup>	97.2					
Tertiary (1) <sup>j</sup>		NA	0.5 <sup>b</sup>	99.4					
Secondary (1)	Canada	31.1 ± 6.7	0.5 ± 0.2	97.1-99.1	63	Visual and FTIR	Fibres	Polyester	[24]

1658 Notes: <sup>a</sup> If not indicated otherwise <sup>b</sup> Anthropogenic litter in the micro range in general and not only microplastic considered, <sup>c</sup> Suspected tyre particles; <sup>d</sup> Range of  
1659 mean concentrations between different WWTP; <sup>e</sup> Retention based on particle mass; <sup>f</sup> Upper size limit was 500 µm; <sup>g</sup> Four different advanced treatment methods  
1660 were tested; <sup>h</sup> Only fibres considered; <sup>i</sup> Pilot scale anaerobic membrane bioreactor; <sup>k</sup> numbers were calculated based on data in the publication. [1] Browne et al.  
1661 (2011) [2] Ziajahromi et al. (2017) [3] Lee and Kim (2018) [4] Liu et al. (2019b) [5] Gündoğdu et al. (2018) [6] Dris et al. (2015) [7] HELCOM (2014) [8] Lares et al.  
1662 (2018) [9] Leslie et al. (2017) [10] Magnusson et al. (2016) [11] Magni et al. (2019) [12] Mintenig et al. (2017) [13] Murphy et al. (2016) [14] Simon et al. (2018)

1663 [15] Talvitie et al. (2015) [16] Talvitie et al. (2017a) [17] Talvitie et al. (2017b) [18] Wolff et al. (2018) [19] Vollertsen and Hansen (2017) [20] Carr et al. (2016) [21]  
 1664 Dyachenko et al. (2017) [22] Mason et al. (2016) [23] Michielssen et al. (2016) [24] Gies et al. (2018)  
 1665

1666 Table S2. Concentration of MPs in sludge from municipal WWTPs. **Bold** numbers represent the median concentration instead of the mean concentration; NR =  
 1667 not reported.  
 1668

Sludge type (no of WWTPs)	Location	Mean Concentration $\pm$ SD in in MP/kg dw <sup>a</sup> or minimum-maximum	Retained in sludge (%)	Lowest mesh sizes or assessed size ( $\mu$ m)	Identification method	Dominant shapes in effluent	Dominant polymer composition in effluent	Reference
Secondary sludge after thickening and dehydration	Korea,	14,895	49.3	106	Visual and FTIR	Fragments (mainly black)	NR	[1]
Secondary sludge after thickening and dehydration	Korea	9,655	44.7	106	Visual and FTIR	Fragments (slightly more than fibres (only fibres and fragments reported))	NR	[1]
Mix of primary and secondary sludge after thickening and dehydration	Korea	13,200	49.0	106	Visual and FTIR	Fragments (slightly more than fibres (only fibres and fragments reported))	NR	[1]

Mix of primary and secondary sludge	China	240,300	NR	20	Visual and Raman	Fragments	Nylon	[2]
Sludge	Spain	NR	NR	20 / 200	visual and FTIR, differential scanning calorimeter	NR	NR	[3]
Sludge (3)	The Netherlands	650 (370-950) <sup>b</sup>	72	10	visual and FTIR	Fibres	NR	[4]
Recycled activated sludge (1)	Italy	113,000 ± 57,000 <sup>b</sup>	NR		visual and FTIR	Fibres	PES	[5]
Sludge (7)	Ireland	4,196 - 15,386	NR	45	visual and FTIR/Raman	Fibres	PE, PES acrylic, PET, PP, polyamide	[6]
Activated sludge (1)	Finnland	23,000 ± 4,200	NR	20	Visual and FTIR/Raman	Fibres	Polyester	[7]
Digested sludge (1)	Finnland	170,900 ± 28,700	NR	20				
Membrane bioreactor sludge (1)	Finnland	27,300 ± 4,700	NR	20				
Sludge (1)	Sweden	16,700 ± 1,960; 720 ± 112 <sup>b</sup>	NR	300	Visual	Fibres	NR	[8]
Primary Sludge (6)	Germany	1x 10 <sup>3</sup> - 2.4 x 10 <sup>4</sup>	NR	20	Visual and FTIR	Fibres	PE	[9]
Sludge (1) (24 h duplicate)	Scotland	800 <sup>b,c</sup>	NR	65	Visual and FTIR	NR	PES, acrylic, PP, alkyd, PS	[10]

Raw sludge	Finland, Europe	76,300		20	Visual and FTIR		NR	[11]
Dry sludge		186700	99.9					
Digested sludge (5)	Denmark, Europe	4.5 mg/g; 169 000 MPs/g  <b>158,000</b> MPs/g; <b>6.5</b> mg/g			Visual and FTIR	NR	PE	[12]
Sludge as biosolid (1)	USA, North America	1,000	99.9 (in grid and biosolids)	20		NR	NR	[13]
Primary sludge (1)	Canada, North America	14,900 <sup>b</sup>	NR	1	visual	Fibres	NR	[14]
Secondary sludge (1)		4,400 <sup>b</sup>	NR	1				
Sludge as biosolid	North America	3,000 - 4,000	NR	NR	visual	Fibres	NR	[15]

1669  
1670  
1671  
1672  
1673  
1674

Notes: <sup>a</sup> If not indicated otherwise; <sup>b</sup> concentration in wet weight, <sup>c</sup> concentration estimated from figure  
[1] (Lee et al. 2019) [2] (Liu et al. 2019b) [3] (Bayo et al. 2016) [4] (Leslie et al. 2017) [5] (Magni et al. 2019) [6] (Mahon et al. 2017) [7] (Lares et al. 2018) [8]  
(Magnusson and Norén 2014) [9] (Mintenig et al. 2017) [10] (Murphy et al. 2016) [11] (Talvitie et al. 2017a) [12] (Vollertsen and Hansen 2017) [13] (Carr et al.  
2016) [14] (Gies et al. 2018) [15] (Zubris and Richards 2005)

1675  
1676  
1677

Table S3. Concentration of MPs and MAPs in different soil types. **Bold** numbers represent the median concentration; \*\* Most common shape or polymer type  
observed. NR = not reported. Dw = dry weight; ww = wet weight.

Soil type (number of fields)	Location	Plastic type (size in mm)	Mean concentration MPs/kg dw <sup>a</sup> (minimum- maximum)	Mean concentration of MeP or MaP/kg	Identification method	Reported shapes	Reported polymer composition	Reference
Industrial	Australia	MPs (< 1)	300 - 67,500 mg/kg	NR	FTIR	NR	<b>PVC</b> , PE, PS,	[1]
Agricultural (vegetable fields)	China	MPs (0.02- 5) / MePs (4- 20)	0-3 cm sample depth: 78.00 ± 12.91; 3-6 sample	0-3 cm sample depth: 6.75 ± 1.51; 3-6 cm	Visually, FTIR	<b>Fibres,</b> <b>Fragments,</b> films,	<b>PP, PE, PES</b>	[2]



			depth cm: 62.50 ± 12.97	sample depth: 3.25 ± 1.04		pellets		
Agricultural (Greenhouse) (4)	China	MPs /MePs (0.05-10)	18,760 (7,100 - 42,960) <sup>d</sup>	NR	Visually	<b>Fibres,</b> fragments, films	NR	[3]
Buffer (former crop land)			14,360 (8,180 - 18,100) <sup>d</sup>	NR				
Agricultural	China		0-10 cm sample depth: 40 ± 126; 0.008 ± 0.025mg/kg;	NR	Visually	PE, PP		[4]
Fruit field			10-30 cm sample depth: 100 ± 141; 0.368 ± 0.740 mg/kg					
			0-10 cm sample depth: 320 ± 329; 0.540 ± 0.603 mg/kg;					
			10-30 cm sample depth: 120 ± 169; 0.460 ± 0.735 mg/kg	NR				
Agricultural (Greenhouse)			0-10 cm sample depth: 100 ± 254; 0.130 ± 0.307 mg/kg;	NR				
			10-30 cm sample depth: 80 ± 193; 0.024 ± 0.051 mg/kg					
Agricultural	Germany	MPs (1-5) and MaPs (>5)	0.34 ± 0.36	206 MaPs/ha or 0.066 kg MaPs/ha;	Visual and FTIR	<b>Fragments, Films,</b>	MaPs: <b>PE;</b> PS ,PP, PVC, PET,	[5]

						Fibres	PMMA MPs: PE, PP, PS	
Floodplain	Switzerland	MPs (0.125-5) and MePs (5- 25)	5 mg/kg dw (0- 55mg/kg dw)	NR	FTIR	PE, PA, natural latex, PS, PVC, SBR, PP		[6]
Agricultural	Denmark	MPs (0.02-0.5)	145,000 <sup>f</sup> ; 12 mg/kg <sup>f</sup>	NR	Visual and FTIR	PE, Nylon and PP		[7]
Agricultural (sewage sludge applied)			71,000 <sup>f</sup> ; 5.8mg/kg <sup>f</sup>					
Agricultural	USA	Synthetic fibres	580 (± 403) – 1210 (± 250) <sup>g</sup>	NR	Polarized microscopy	Fibres		[8]
Agricultural (6) 0-25 cm	Chile	MPs <sup>i</sup>	1.37 - 4.38 <sup>h</sup> (0.73 -12.9) mg/kg dw	NR	Visual	Fibres, Films, Fragments, Pellets		[9]
Rural home gardens (agro-forestry land-use system)	Mexico	MPs (<5 <sup>i</sup> )/ MaPs (> 5)	870 ± 1,900	744,000 ± 204 000 PE bottles /ha 74 000 ± 65 000 MaPs/m <sup>2</sup>	Visual	NR	NR	[10]
Horticulture	Argentina	PE MaP (> 20)	NR	3 ± 1.9 g/m <sup>2</sup>	Visually	Films <sup>k</sup>	PE <sup>k</sup>	[11]
Dessert	USA	MaP	NR	0.056 – 0.344 bags /ha; 0.392-0.627 balloon clusters/ha	Visually	Bags, balloons <sup>l</sup>	PE, latex	[12]

1678  
1679  
1680

Notes: <sup>a</sup> If not indicated otherwise; <sup>b</sup> Wet weight; <sup>c</sup> Calculated mean concentration from all dept fraction; <sup>d</sup> Not distinguished into concentration of MPs and MePs but 95% of the observed plastic particles are in the MPs size (0,05 -1mm) ; <sup>e</sup> Extraction method was only suitable for low density plastics; <sup>f</sup> Not reported if dry or

1681 wet weight; <sup>g</sup> Only synthetic fibres concentration assessed; <sup>h</sup>Range of medians is shown, which was increasing with increasing number of sludge applications; <sup>i</sup>  
 1682 Lower detection limit not reported; <sup>k</sup> Only PE film concentration assessed; <sup>l</sup> Only bags and balloons assessed. [1] Fuller and Gautam (2016) [2] Liu et al. (2018)  
 1683 [3] Zhang and Liu (2018) [4] Zhang et al. (2018) [5] Piehl et al. (2018) [6] Scheurer and Bigalke (2018) [7] Vollertsen and Hansen (2017) [8] Zubris and Richards  
 1684 (2005) [9] Corradini et al. (2019) [10] Huerta Lwanga et al. (2017) [11] Ramos et al. (2015) [12] Zylstra (2013)  
 1685

1686

1687 Table S4. MaPs reported in surface waters.

Waterbody (number of waterbodies)	Location	Sample type (sampling type)	Reported Concentration range	Lowest assessed size (mm)	Observed plastic types	Reference
Saigon River	Vietnam	Water (floated plastic intercepted by nets)	Median estimated amount entering the river: 4.43 (0.96 - 19.9) g/inhabitant/day	20	Bags, bottles, drinking recipients, plastic cutlery PE (79%), PP (15%), PET (4%)	[1]
Saigon River	Vietnam	Water (floated plastic intercepted by nets)	0.2 - 0.3 tons per day emitted to the ocean	50	PS food container fragments), PS foam polyolefin bags and food wrappings, caps, lids, polyolefin hard plastic, fragments, cups, bottles, straws, others	[2]
Yangtze River	China	Water (trawl net)	Mean: $8.74 \times 10^3$ items/km <sup>2</sup> ( $1.94 \times 10^3$ - $2.78 \times 10^4$ items/km <sup>2</sup> )	5 <sup>b</sup>		[3]
Rhone River	France	Water (visual observation from elevated points)	NR	70	Bags, sheets, bottles, covers/packaging, others	[4]
Tiber River	Italy	Water (visual observation from elevated points)	1,270 litter items/km <sup>2</sup> ; 190 litter items > 20 cm/km <sup>2.c</sup>	25	Plastic pieces, bottles, covers, polystyrene pieces, cover/packaging, foam	[5]
Lakes (7)	Switzerland	Water (Manta trawl)	Mean: $1,800 \pm 3,100$ items/km <sup>2</sup> , $44,000 \pm 80,000$ mg/km <sup>2</sup> ; median: 860 items/km <sup>2</sup> , 12,000 mg/km <sup>2</sup>	5	PE (mainly packaging films), PP (mainly fragments), PS (mainly foams)	[6]

Rivers (5)	Switzerland	Water	Mean: 0.012±0.034 items/m <sup>3</sup> , 0.43 ± 12 mg/m <sup>3</sup> ; median: 0	5		[6]
River Seine	France	Water (plastic trapped by floating debris retention booms)	27,000 tons floating plastic intercepted annually		PP, PE, PET	[7]
Upper Thames estuary	UK	Water (plastic intercepted by nets close to riverbed)	Total number of items: 8480 <sup>c</sup>	NR	Bags, cups, plates, forks, food wrappers, Tabaco packaging, sanitary components, others	[8]
Los Angeles River	USA	Water (plastic intercepted by nets)	819 items/m <sup>3</sup>	4.75	NR	[9]
San Garbiel River	USA	Water (plastic intercepted by nets)	125 items/m <sup>3</sup>	4.75	NR	[9]

1688 Notes: <sup>b</sup> only mesoplastics and not MaPs assessed but the upper size limit is unknown; <sup>c</sup> Floating litter in general assessed and not only plastic. [1] Lahens et al.  
1689 (2018) [2] van Emmerik et al. (2018) [3] Xiong et al. (2019) [4] Castro-Jiménez et al. (2019) [5] Crosti et al. (2018) [6] Faure et al. (2015) [7] Gasperi et al. (2014)  
1690 [8] Morritt et al. (2014) [9] Moore et al. (2011)

1691  
1692  
1693  
1694  
1695  
1696  
1697

Table S5. Concentration of MPs in different waterbodies with sample type, mesh size limit, identification methods, reported shapes and polymer compositions. **Bold** numbers represent the median concentration; \* Concentration in MPs/ m<sup>3</sup> was estimated by dividing the reported concentration in particles per area by the height of the net used for sampling; \*\* Most common shape or polymer type observed. NR = not reported. Dw = dry weight; ww = wet weight.

Waterbody (number of waterbodies)	Location	Mean concentration in MP/m <sup>3</sup> <sup>a</sup> ± SD or (minimum - maximum)	Sample type	Lowest mesh sizes or lowest size limit (µm)	Identification method	Reported shapes	Reported polymer composition	Reference
Hovsgol Lake (1)	Mongolia	0.127 (0.01 - 0.28)*	Manta trawl	333	Visual	Fragments films, lines/fibres	NR	[1]

Yangtze River (1)	China	4,700 ± 2,800; <b>4130</b> (1600 - 1.26 x10 <sup>4</sup> )	Water pumped through stainless steel sieve	48	Visual and Raman	Fibres**, Fragments, (Pellets)	PS**, PP, PE, PC, PVC, Vinyl chloride, others	[2]
Small Waterbodies in Yangtze River catchment (25)	China	480 - 21,520	Grab water	20	Visual and FTIR	Fibres	PES	[3]
Perl River (1)	China	2,724 (379 - 7,924)	Water sieved	20	Visual and FTIR	Fibres	PE, PP	[4]
Shanghai creeks (1)	China	1.25 (0.5 -4 .2) <sup>b</sup>	Pump or metal pail	20	Visual and FTIR	Fibres**, Fragments, Films/Pellets	PES**, Rayon, PP	[5]
Suzhou River (1)		1.5 (0.1 - 7.5) <sup>b</sup>						
Huangpu River (1)		0.95 (0.25 - 7.5) <sup>b</sup>						
Taihu lake (1)	China	0.03 (20 <sup>d</sup> )	Plankton net	333	Visual and FTIR	Fibres**, Fragments, Films/Pellets	Cellophane**, PET, PES, Terephthalic acid, PP	[6]
		3,400 - 25,800	Surface water grab sample	5				
Feilaixia Reservoir Beijing River (1)	China	0.56	Plankton net	112	Visual and FTIR	Foams**, Fragments, Films, Fibres	PP**, PE, EPS, PS, PET, PVC	[7]
Yangtze River (1)	China	2,516 (1,400 - 4,000)	Water pumped through stainless steel sieve	50	Visual and FTIR	Fibres**, Granules, Films, (Pellets)	PET**, PP, PE, Nylon, PS	[8]
Hanjiang River (1)		29,933 (2,600 - 3,200)						
Surface waters of Wuhan: Lakes (20), Yangtze River (1) and Hanjiang River (1)		1,660 ± 6,391 – 8,925 ±1,591						
Dongting Lake (1)	China	1,191.7	Water pumped through stainless steel sieve	50	Visual and Raman	Fibres**, Granules, Films	PE**, PP**, PS, PVC	[9]
Hong Lake (1)		2,282.5						
Yangtze River (1)	China	0.86 (0.34 - 1.58)*	Trawl net	333	Visual and Raman	Sheets**, Fragments**, Foams, Lines	PP**, PE, PS, others	[10]
Yangtze River (1)		2,113 (1,260 - 4,340)	Water filtered through plankton net	64				
Pearl River, urban	China	19,860 (8,750 - 53,250)	Water filtered	50	Visual and Raman	Films**, Polyamide**, [11]		

part (1)			through stainless steel sieve			Granules, Fibres	Cellophane, PP, PE	
Pearl River, estuary part (1)		8,902 (7,850 - 1.1x10 <sup>4</sup> )				Granules**, Films**, Fibres		
Poyang Lake (1)	China	5,000 – 34,000	Water filtered through stainless steel sieve	50	Visual and Raman	Fibres**, Films, Fragments, Pellets	PP, PE, Nylon, PVC	[12]
Yangtze River (1)	China	16.8 (5.96 - 23.83)*	Trawl net	112	Visual and FTIR	Sheets**, Fragments, Lines, Foam	PE**, PP**, PS	[13]
Yangtze River Tributaries (4)		6.663 (0.34 - 20.81)*						
Rivers (29)	Japan	<b>7.9</b> ; 1.6 ± 2.3 (0 - 12)	Plankton net	335	FTIR	Fragments (only mentioned)	PE**, PP, PS, others	[14]
Saigon River and Canals (1)	Vietnam	10 - 223	Net sample			Fragments	PE**, PP, PE-PP, PS, others	[15]
		17,200 - 519,000	Grab water sample	300	Visual and FTIR	Fibres**	PES**, PET PE, PP, others	
River Seine (1)	France	30 (3 - 108)	Plankton net	80	Visual	Fibres	NR	[16]
		0.35 (0.28 - 0.47)	Mantra trawl	330		Fibres, Fragments, Spheres		
Lakes (7)	Switzerland	0.51 ± 0.67; <b>0.27*</b>	Manta trawl	300	Visual and FTIR	Fragments**, Foams, Films, Fibres, Others	PE**, PP, PS	[17]
Rivers (4)		7 ± 0.2; <b>0.36</b>						
Lake Geneva (1)	Switzerland	0.193*	Manta trawl	300	Visual	NR	NR	[18]
Lake Bolsena (1)	Italy	0.82 - 4.42	Manta trawl	300	Visual (UV microscope) and SEM	Fragments**, Fibres**	NR	[19]
Lake Chiusi (1)		2.68 - 3.36				Fibres**, Fragments		

Lake Iseo (1)	Italy	0.2*	Manta trawl	300	Visual and FTIR	Fragments**, Filaments, sheets, others	PE**, EPS; PP, others	[20]
Lake Maggiore (1)		0.195*						
Lake Garda (1)		0.125*						
Danube (1, Year 2010)	Austria	0.983 (0 - 141.66)*	Conical drift nets	500	Visual	Spherules**, Flakes, Pellets	NR	[21]
Danube (1, Year 2012)		0.055 (0 - 0.75)*				Pellets**, Flakes, Spherules		
Amsterdam Canals (1)	Netherlands	100,000 ± 49,000 (48,000 - 187,000)	Grab sample	10	Visual and FTIR	Fibres**, Spheres, Foils	NR	[22]
Storm water ponds (1)	Denmark	1,409 (490 - 22,894)	Filtered through stainless steel mesh	10	Visual and FTIR	NR	PP**, PVC, PES, PE, PS, Others	[23]
Rhine (1)	Switzerland, France, Germany	4.96; <b>2.196</b> ; 2, 684 <sup>d,*</sup>	Manta trawl	300	Visual and FTIR	Spherules**, Fragments, Fibres	PS**, PP, Others	[24]
Antua River, Portugal, March (1)	Portugal	58 - 193	Surface and bottom water filtered	55	Visual and FTIR	Fragments**, Fibres, Foams, Films, Pellets	PE**, PP, Others	[25]
Antua River, Portugal, October (1)		71 - 1,265						
Gallatin River watershed (1)	USA	1,200 (0 - 67,500)	Grab sample	100	Visual and FTIR	Fibres**, Fragments, (Beads)	Semi-synthetic cellulose**, PET, PES, PVA, Neoprene	[26]
Great Lake Tributaries (29)	USA	1.9; <b>4.2</b> (0.05 - 32)	Neuston net	333	Visual	Fibres**, Fragments, Foams, Films	NR	[27]
Raritan River	USA	24.0	Plankton net	125	Visual	NR	NR	[28]

upstream WWTP (1)								
Raritan River downstream WWTP (1)		71.7						
Lake Superior (1)	USA	<b>0.022</b> (0.01 - 0.08)*	Manta trawl	333	Visual and SEM/EDS	Pellets**, Fragments, Foams, Films, Lines	NR	[29]
Lake Huron (1)		<b>0.012</b> (0 - 0.04)*						
Lake Erie (1)		<b>0.127</b> (0.03 - 2.91)*						
Snake River/Columbia River (1)	USA	91 ± 1,140 (0 - 5405,000)	Grab sample	100	Raman	Fibres**, Fragments, Beads, Films	PP, PE, PET, PES	[30]
Snake River/Columbia River (1)		2.57 ± 2.95 (0 - 13.5)	Net sample	100				
Lakes in Lubbock Texas (1)	USA	53 - 105 µm: 0.79 - 1.56 mg/L; 106 - 179 µm: 0.31 - 1.25 mg/L	Grab sample	53-179	Visual	Beads, Filamentous MPs, Irregular MPs	NR	[31]
Wetlands in Texas (1)		53 - 105 µm: 0.64 - 5.51 mg/L; 106 - 179 µm: 0 - 1.79 mg/L						
Streams Chicago metropolitan area, upstream WWTP (1)	USA	2.355 ± 0.375	Neuston nets	333	Visual and Py-GCMS	Pellets**, Fibres**, Fragments**, others	PP**, PE**, PS**	[32]
Streams Chicago metropolitan area, downstream WWTP (1)		5.733 ± 0.850						



North Shore Channel Chicago, upstream WWTP (1)	USA	1.94 ± 0.81	Neuston nets	333	SEM	Fibres**, Fragments	NR	[33]
North Shore Channel Chicago, downstream WWTP (1)		17.93 ± 11.05				Fibres**, Fragments, Pellets, Styrofoam		
Hudson River (1)	USA	980°	Grab sample	330	FTIR	Fibres	NR	[34]
La River (1)	USA	22 - 12,932	Manta trawl, Hand nets, Rectangular nets, streambed sampler	1000	Visual	Foamed plastic**, Pellets, Fragments, Films, Lines, whole items	NR	[35]
San Gabriel River (1)		0 - 337				Foamed plastic**, Fragments, Films, whole items, Lines, Pellets		
Coyote Creek (1)		27,211				Fragments**, Foams, Lines, Pellets, whole items		
Estuarine Rivers Chesapeake Bay (1)	USA	0 - 0.036g/m <sup>3</sup> *	Surface trawl	330	Visual and Raman	Fragments**, Sheets**, Fibres, EPS, others	PE	[36]

Lake Superior (1)	USA	0.263 ± 0.193 (0-0.786)* 0.086 (0 - 0.0253 mg/m <sup>3</sup> )	Manta trawl	333- 4 mm?	Visual and yr-GC/MS and ATR-FTIR	Fibres**, Fragments, Films, (Beads, Foams, Others)	PE, PVC, PP, PET, others	[37]
Lake Winnipeg (1)	Canada	1.07 (0.29 - 4.16)	Manta trawl	333	Visual and SEM-XDS	Fibres**, Fragments, Films, Foams	NR	[38]
Ottawa river and tributaries, nearshore (1)	Canada	100 (50 - 240)	Grab sample filtered	100	Visual	Fibres	NR	[39]
Ottawa river and tributaries, open water (1)		1.35	Manta trawl			Fibres**, Fragments, Beads		
Ottawa river upstream WWTP (1)		0.71				NR		
Ottawa river downstream WWTP (1)		1.99				NR		

1698  
1699  
1700  
1701  
1702  
1703  
1704  
1705  
1706  
1707  
1708  
1709  
1710

Notes: <sup>a</sup> If not indicated otherwise; <sup>b</sup> Numbers for different sample sites were estimated from graph and mean numbers were calculated; <sup>c</sup> Only fibres included; <sup>d</sup> Maximum observed concentration. [1] Free et al. (2014) [2] Di and Wang (2018) [3] Hu et al. (2018) [4] Lin et al. (2018) [5] Luo et al. (2018) [6] Su et al. (2016) [7] Tan et al. (2019) [8] Wang et al. (2017b) [9] Wang et al. (2018) [10] Xiong et al. (2019) [11] Yan et al. (2019) [12] Yuan et al. (2019) [13] Zhang et al. (2015) [14] Kataoka et al. (2019) [15] Lahens et al. (2018) [16] Dris et al. (2015) [17] Faure et al. (2015) [18] Faure et al. (2012) [19] Fischer et al. (2016) [20] Sighicelli et al. (2018) [21] Lechner et al. (2014) [22] Leslie et al. (2017) [23] Liu et al. (2019a) [24] Mani et al. (2016) [25] Rodrigues et al. (2018) [26] Barrows et al. (2018) [27] Baldwin et al. (2016) [28] Estahbanati and Fahrenfeld (2016) [29] Eriksen et al. (2013) [30] Kapp and Yeatman (2018) [31] Lasee et al. (2017) [32] McCormick et al. (2016) [33] McCormick et al. (2014) [34] Miller et al. (2017) [35] Moore et al. (2011) [36] Yonkos et al. (2014) [37] Hendrickson et al. (2018) [38] Anderson et al. (2017) [39] Vermaire et al. (2017)

Table S6. Concentration of MaPs in sediments. **Bold** numbers represent the median concentration

Waterbody (no of waterbodies)	Location	Sample type	Reported mean (min-max) concentration <sup>a</sup>	Lowest assessed size (mm)	Observed plastic types	Reference
Rivers (6)	Switzerland	Beach sediment	90 ± 250 items/m <sup>2</sup> ; 14,000 ± 33,000 mg	5	PE (mainly packaging films), PP (mainly fragments), PS (mainly	[1]

			/m; <b>11 items /m<sup>2</sup>, 480 mg/m<sup>2</sup></b>		foams)	
Lake Bolsena	Italy	Beach sediment	North shore: 2.57 items/m <sup>2</sup> , 2.6 g/m <sup>2</sup> ; South shore: 0.28 items/m <sup>2</sup> , 1.1 g/m <sup>2</sup>	5	Industrial packaging (PE, PP), food packaging (PE), net/rope/string/cord (polyamide, PVC, polyacrylonitrile), others	[2]
Lake Chiusi	Italy	Beach sediment	East shore: 5 items/m <sup>2</sup> , 4.5 g/m <sup>2</sup> ; west shore: 0.22 items/m <sup>2</sup> , 0.2 g/m <sup>2</sup>	5	Cigarette butts (cellulose acetate), net/rope/string/cord (polyamide, PVC, polyacrylonitrile), others	[2]
Albegna River	Italy	Sediment	Winter: 7-12 items/kg; Summer: 16 -43 items/kg	5.1 – 25	Filaments, fragments, others	[3]
Osa River			Winter: 7-12 items/kg; Summer: 16 -43 items/kg			
Ombro River			Winter: 0 items/kg; Summer 0-14 items/kg			
Lake Garda	Italy	Beach sediment	North shore: 483 ± 236 items/m <sup>2</sup> ; South shore: 0 - 8.3 items/m <sup>2</sup>	5	NR	[4]
Lake Garda	Italy	Beach sediment	5 ± 9 items/m <sup>2</sup>	5	NR	[5]
Edgbaston Pool,	UK	Sediment	2- 20 items per sampling site	NR	Food wrappers, bottle caps, plastic bags, Styrofoam, bottles, films, fragments, ropes, straws, syringe, cosmetic tubes, fibrous clothing	[6]
River Taff	UK	River bank	584 plastic items/0.1 km river bank <sup>c</sup>	NR	Plastic, packaging, others	[7]
Lake Ontario	Canada	Beach sediment	366 items/kg <sup>b</sup>	5	Fragments	[8]
Lake shoreline	Argentina	Shoreline sediment	meso: 25 items/m <sup>2</sup> , 19 g/m <sup>2</sup> ; MaP: 1.15	meso: 5 -25 MaP > 25	Food wrappers (PP &PS), bags (PE), bottles (PET), Styrofoam food	[9]

			items/ m <sup>2</sup> ; 4.9 g m <sup>2</sup> ;		containers (expanded PS)	
Rivers (4)	Chile	River bank	up to <b>3.4</b> items/m <sup>2</sup> . <sup>c</sup>	1.5	NR	[10]

1711 Notes: <sup>a</sup> If not indicated otherwise; <sup>b</sup> Mean has been calculated across different sampling dates and includes only fragments; <sup>c</sup> Persistent buoyant litter in general  
1712 assessed and not only plastic. [1] Faure et al. (2015) [2] Fischer et al. (2016) [3] Guerranti et al. (2017) [4] Imhof et al. (2013) [5] Imhof et al. (2018) [6] Vaughan  
1713 et al. (2017) [7] Williams and Simmons (1999) [8] Corcoran et al. (2015) [9] Blettler et al. (2017) [10] Rech et al. (2014)

1714

1715

1716

1717

1718

1719

1720

1721

Table S7. Concentration of MPs sediments of different waterbodies with sample type, mesh size limit, Identification methods, reported shapes and polymer compositions.

**Bold** numbers represent the median concentration; \* Concentration in MPs/kg was estimated by using the sample depth and assuming a density of 1.6 g/cm<sup>3</sup> for the sediment; \*\* Most common shape or polymer type observed. NR = not reported.

Waterbody (number of waterbodies)	location	Sample type	Average concentration in MP/kg dry weight $\pm$ SD or (minimum-maximum) <sup>a</sup>	Type of analysis	Lowest measured size ( $\mu$ m)	Reported shapes	Reported polymer compositions	Reference
Bloukrans River (summer)	South Africa	Bed sediment	6.3 $\pm$ 4.3 (1 - 14.61)	Visual	63	Fibres	NR	[1]
Bloukrans River (winter)			160.1 $\pm$ 139.5 (13.3 - 563.8)					
Three Gorges Reservoir	China	Bed sediment	82 $\pm$ 60 (25 - 300) <sup>b</sup>	Visual and Raman	48	Fibres**, Fragments, Pellets, Film, Styrofoam	PS**, PP, PE	[2]
Small waterbodies in Yangtze River Delta	China	Bed sediment	35.8 - 3185 <sup>c</sup>	Visual and FTIR	20	Fibres**, Fragment, Granule	PP, PE, PES	[3]
Pearl River	China	Bed sediment	1,669 (80 - 9,597) <sup>c</sup>	Visual and FTIR	20	Fibres, Fragments, Films	PE**, PP,	[4]
Rivers Shanghai (6)	China	Bed sediment	802 $\pm$ 594	Visual and FTIR	1	Spheres**, Fibres, Fragments	PP**, PES, Rayon, Others.	[5]
Vembanad Lake	India	Bed sediment	252.8 $\pm$ 25.76 (96 - 496)	Visual and Raman		Films**, Foams**, Fragments, Fibres	High density PE, Low density PE, PS; PP	[6]

			MPs/m <sup>2.g</sup>					
Taihu Lake	China	Bed sediment	11 - 234.6	Visual and Raman	1.2	Fibres, Fragments, Films, Pellets	Cellophane**, PET, PES, terephthalic acid, PP	[7]
Beijiang River	China	Shore sediment	178 ± 69 - 544 ± 107 <sup>d</sup>	Visual and FTIR	1	NR	PE**, PP, Copolymer	[8]
Lakes in Changsha (12)	China	Bed sediment	270.17 ± 48.23 - 866.54 ± 37.96 <sup>d</sup>	Visual and Raman	<500	Fragments**, Fibres, Films, Foams	PS**, PE, PET, PP, Polyamide, PVC	[9]
Yangtze River	China	Bed sediment	34 (7 - 66) <sup>c</sup>	Visual and Raman	333	NR	NR	[10]
Poyang Lake	China	Bed sediment	54 - 506	Visual and Raman	50	Fibres**, Fragments, Films, Pellets	PP**, PE PVC, Nylon	[11]
Lakes Siling Co Basin (4)	China	Shore sediment	<0.125 – 17.63 ± 35.3	Visual and Raman	1	Sheets, Lines, Fragments Foams	PP**, PE**, PVC, PET, PS	[12]
Lakes (6)	Switzerland	Beach sediment	16.25 ± 25 (10 - 86.25) <sup>*</sup>	Visual and FTIR	300	Foams, Fragments, Fibres, Films, Pellets. Lines, Beads	PE, PP, PS, PVC	[13]
Lakes Bolsena				UV microscope and Scanning Electronic Microscope	300	Fragments**, Fibres		[14]
Lake Chiusi	Italy	shore sediment	44.10 ± 14.48 <sup>*</sup>			Fibres**, Fragments		
Thames tributaries								[15]
Leach			185 ± 42					
Lambourn			221 ± 95					
Cut Site 1	UK		665 ± 77		1000 <sup>e</sup>	Fibres**, Fragments, Films		
Cut Site 2		Bed sediment	332 ± 161 <sup>*</sup>	Visual and Raman / x-XRF		Fragments**, Fibres, Films	PES**, PET**, PP polyacrylsulphane, PE, PS PVC and others	
Upper Mersey and Irwell catchments	UK	Bed sediment	323.13 <sup>*.f</sup>	Visual and FTIR	50	Beads**, Fragments, Fibres	NR	[16]

Lake Garda	Italy	Beach sediment	43.85 ± 110.69*	Visual and Raman	1	NR	NR	[17]
Lake Garda	Italy	Beach sediment	0.94 ± 1.68	Visual and Raman	1	Plastic and paint particles	Polyamide, PE, PS, PP and others	[18]
Lake Garda	Italy	Beach sediment	1.35 ± 0.69 - 13.85 ± 12.29*	Visual and Raman	9	Fragments**, Fibres	PS, PE	[19]
Rivers Rhine and Main	Germany	Shore sediment	228 - 3 763	Visual and FTIR	63	Fragments**, Fibres**, Spherules	PS**, PE, PP, Polyamide, Others	[20]
Amsterdam canals	Netherlands	Bed sediments	2,071 ± 4,246 (0 - 10,500)	Visual and FTIR	10	Spheres, Fibres, Foils	NR	[21]
Antuã River (Spring)	Portugal	Bed sediment	100 - 624	Visual and FTIR	55	Fragments**, Fibres, Foams, Pellets	PE**, PP**, PS, PET	[22]
Antuã River (Autumn)			18 - 514					
River Tames and tributaries	UK	Bed sediment	165 (20 - 350)	Visual and FTIR	63	Fragments**, Fibres, Spheres, Foams, Films, irregular Spheres, commercial Fragments	PE**, PVC, Polyamide,	[23]
Edgbaston Pool	UK	Bed sediment	250 - 300	Visual	500	Fibres, films, foams, Fragments	NR	[24]
Lake Ontario	Canada	Shore and beach sediment	760 (20 - 27,830)	Visual and Raman/x-XRF (X-ray fluorescence spectroscopy)	63	Fragments**, Fibres, Beads	PE**, PS, Polyurethane, PP, PVC, PSS, Others	[25]
St Lawrence River	Canada	Bed Sediment	85.99 ± 86.53*; <b>0.325*</b> (0 -248.75*)	Visual	500	NR	NR	[26]
Lake Ontario	Great Lakes, Canada	Beach sediment	19,175.2 – 27,774.79 <sup>h</sup>	Visual and Raman, FTIR	<1	Pellets**, Fragments and polystyrene	PE**, PP, NC	[27]
Atoyac River	Mexico	Bed sediment	155.17 (33.33 - 266.67) <sup>d</sup>	Visual	1.2	Films**, Fragments, Fibres, pellets		[28]
Ottawa River and tributaries	Canada	Bed sediment	220	Visual	100	Fibres**, beads fragments		[29]
Lake Huron	Great	Beach	408 MPs/m <sup>2</sup> .	Visual and	NR <sup>i</sup>	Pellets**,	PE**, PP, PET	[30]

	Lakes, Canada	sediment	f,g	FTIR/ SEM		fragments, Styrofoam		
Lakes Erie	Canada	Beach sediment	0.36 - 3.7 MPs/m <sup>2,g</sup>	Visual and FTIR	NR <sup>i</sup>	Fragments**, Pellets, Styrofoam	PE, PP	[31]
Lake St. Clair			0.18 - 8.38 MPs/m <sup>2,g</sup>			Fragments**, Styrofoam, Pellets,		
Lake Huron			0.98 - 34 MPs/m <sup>2,g</sup>			Pellets**, Fragments, Styrofoam		
Setúbal Lake	Argentina	Shore sediment	14.67*	Visual and FTIR	350	Hard plastic, Fibres		[32]
River Albegna	Italy	Bed sediment	Winter: 305 - 477; Summer: 202-253	Visual	10	Filaments, Fragments, (Films)	NR	[33]
River Osa			Winter: 312; Summer 259					
River Ombrone			Winter: 75 - 188; Summer: 137 - 168					

1722  
1723  
1724  
1725  
1726  
1727  
1728  
1729  
1730

Notes: <sup>a</sup> If not indicated otherwise; <sup>b</sup>Wet weight; <sup>c</sup>Not mentioned if wet or dry, <sup>d</sup>Range of means between different sampling sites, <sup>e</sup>Upper size limit 4000 µm; <sup>f</sup>Maximum observed concentration; <sup>g</sup>Not sufficient information available to estimate the number in MPs/kg, <sup>h</sup> Calculated from data in the paper.

[1] Nel et al. (2018) [2] Di and Wang (2018) [3] Hu et al. (2018) [4] Lin et al. (2018) [5] Peng et al. (2018) [6] Sruthy and Ramasamy (2017) [7] Su et al. (2016) [8] Wang et al. (2017a) [9] Wen et al. (2018) [10] Xiong et al. (2019) [11] Yuan et al. (2019) [12] Zhang et al. (2016) [13] Faure et al. (2015) [14] Fischer et al. (2016) [15] Horton et al. (2017) [16] Hurley et al. (2018) [17] Imhof et al. (2018) [18] Imhof et al. (2016) [19] Imhof et al. (2013) [20] Klein et al. (2015) [21] Leslie et al. (2017) [22] Rodrigues et al. (2018) [23] Tibbetts et al. (2018) [24] Vaughan et al. (2017) [25] Ballent et al. (2016) [26] Castañeda et al. (2014) [27] Corcoran et al. (2015) [28] Shruti et al. (2019) [29] Vermaire et al. (2017) [30] Zbyszewski and Corcoran (2011) [31] Zbyszewski et al. (2014) [32] Blettler et al. (2017) [33] Guerranti et al. (2017)

## References

- Anderson PJ, Warrack S, Langen V, et al (2017) Microplastic contamination in Lake Winnipeg, Canada. *Environ Pollut* 225:223–231. doi: 10.1016/j.envpol.2017.02.072
- Baldwin AK, Corsi SR, Mason SA (2016) Plastic Debris in 29 Great Lakes Tributaries: Relations to Watershed Attributes and Hydrology. *Environ Sci Technol* 50:10377–10385. doi: 10.1021/acs.est.6b02917
- Ballent A, Corcoran PL, Madden O, et al (2016) Sources and sinks of microplastics in Canadian Lake Ontario nearshore, tributary and beach sediments. *Mar Pollut Bull* 110:383–395. doi: 10.1016/j.marpolbul.2016.06.037
- Barrows APW, Christiansen KS, Bode ET, Hoellein TJ (2018) A watershed-scale, citizen science approach to quantifying microplastic concentration in a mixed land-use river. *Water Res* 147:382–392. doi: 10.1016/j.watres.2018.10.013
- Bayo J, Olmos S, López-Castellanos J, Alcolea A (2016) Microplastics and microfibers in the sludge of a municipal wastewater treatment plant. *Int J Sustain Dev Plan* 11:812–821. doi: 10.2495/SDP-V11-N5-812-821
- Blettler MCM, Ulla MA, Rabuffetti AP, Garelo N (2017) Plastic pollution in freshwater ecosystems: macro-, meso-, and microplastic debris in a floodplain lake. *Environ Monit Assess* 189:. doi: 10.1007/s10661-017-6305-8
- Browne MA, Crump P, Niven SJ, et al (2011) Accumulations of microplastic on shorelines worldwide: sources and sinks. *Environ Sci Technol* 9175–9179. doi: 10.1021/es201811s
- Carr SA, Liu J, Tesoro AG (2016) Transport and fate of microplastic particles in wastewater treatment plants. *Water Res* 91:174–182. doi: 10.1016/j.watres.2016.01.002
- Castañeda RA, Avlijas S, Simard MAA, et al (2014) Microplastic pollution in St. Lawrence River sediments. *Can J Fish Aquat Sci* 71:1767–1771. doi: 10.1139/cjfas-2014-0281
- Castro-Jiménez J, González-Fernández D, Fornier M, et al (2019) Macro-litter in surface waters from the Rhone River: Plastic pollution and flows to the NW Mediterranean Sea. *Mar Pollut Bull* 146:60–66. doi: 10.1016/j.marpolbul.2019.05.067
- Corcoran PL, Norris T, Ceccanese T, et al (2015) Hidden plastics of Lake Ontario, Canada and their potential preservation in the sediment record. *Environ Pollut* 204:17–25. doi: 10.1016/j.envpol.2015.04.009
- Corradini F, Bartholomeus H, Lwanga EH, et al (2019) Predicting soil microplastic concentration using vis-NIR spectroscopy. *Sci Total Environ* 650:922–932. doi: <https://doi.org/10.1016/j.scitotenv.2018.09.101>
- Crosti R, Arcangeli A, Campana I, et al (2018) ‘Down to the river’: amount, composition, and economic sector of litter entering the marine compartment, through the Tiber river in the Western Mediterranean Sea. *Rend Lincei* 29:859–866. doi: 10.1007/s12210-018-0747-y
- Di M, Wang J (2018) Microplastics in surface waters and sediments of the Three Gorges Reservoir, China. *Sci Total Environ* 616–617:1620–1627. doi: 10.1016/j.scitotenv.2017.10.150
- Dris R, Gasperi J, Rocher V, et al (2015) Microplastic contamination in an urban area: A case study in Greater Paris. *Environ Chem* 12:592–599. doi: 10.1071/EN14167
- Dyachenko A, Mitchell J, Arsem N (2017) Extraction and identification of microplastic particles from secondary wastewater treatment plant (WWTP) effluent. *Anal Methods* 9:1412–1418. doi: 10.1039/c6ay02397e
- Eriksen M, Mason S, Wilson S, et al (2013) Microplastic pollution in the surface waters of the Laurentian Great Lakes. *Mar Pollut Bull* 77:177–182. doi: 10.1016/j.marpolbul.2013.10.007
- Estahbanati S, Fahrenfeld NL (2016) Influence of wastewater treatment plant discharges on microplastic concentrations in surface water. *Chemosphere* 162:277–284. doi: 10.1016/j.chemosphere.2016.07.083
- Faure F, Corbaz M, Baecher H, De Alencastro LF (2012) Pollution due to plastics and microplastics in lake Geneva and in the Mediterranean Sea. *Arch des Sci* 65:157–164. doi: 10.1071/EN14218
- Faure F, Demars C, Wieser O, et al (2015) Plastic pollution in Swiss surface waters: Nature and concentrations, interaction with pollutants. *Environ Chem* 12:582–591. doi: 10.1071/EN14218
- Fischer EK, Paglialonga L, Czech E, Tamminga M (2016) Microplastic pollution in lakes and lake shoreline sediments - A case study on Lake Bolsena and Lake Chiusi (central Italy). *Environ Pollut* 213:648–657. doi: 10.1016/j.envpol.2016.03.012
- Free CM, Jensen OP, Mason SA, et al (2014) High-levels of microplastic pollution in a large, remote, mountain lake. *Mar Pollut Bull* 85:156–163. doi: 10.1016/j.marpolbul.2014.06.001
- Fuller S, Gautam A (2016) A Procedure for Measuring Microplastics using Pressurized Fluid



- Extraction. *Environ Sci Technol* 50:5774–5780. doi: 10.1021/acs.est.6b00816
- Gasperi J, Dris R, Bonin T, et al (2014) Assessment of floating plastic debris in surface water along the Seine River. *Environ Pollut* 195:163–166. doi: 10.1016/j.envpol.2014.09.001
- Gies EA, Lenoble JL, Noël M, et al (2018) Retention of microplastics in a major secondary wastewater treatment plant in Vancouver, Canada. *Mar Pollut Bull* 133:553–561. doi: 10.1016/j.marpolbul.2018.06.006
- Guerranti C, Cannas S, Scopetani C, et al (2017) Plastic litter in aquatic environments of Maremma Regional Park (Tyrrhenian Sea, Italy): Contribution by the Ombrone river and levels in marine sediments. *Mar Pollut Bull* 117:366–370. doi: 10.1016/j.marpolbul.2017.02.021
- Gündoğdu S, Çevik C, Güzel E, Kilercioğlu S (2018) Microplastics in municipal wastewater treatment plants in Turkey: a comparison of the influent and secondary effluent concentrations. *Environ Monit Assess* 190:. doi: 10.1007/s10661-018-7010-y
- HELCOM (2014) Preliminary study on synthetic microfibers and particles at a municipal waste water treatment plant. *Balt Mar Environ Prot Comm HELCOM* 14 p.
- Hendrickson E, Minor EC, Schreiner K (2018) Microplastic Abundance and Composition in Western Lake Superior As Determined via Microscopy, Pyr-GC/MS, and FTIR. *Environ Sci Technol* 52:1787–1796. doi: 10.1021/acs.est.7b05829
- Horton AA, Walton A, Spurgeon DJ, et al (2017) Microplastics in freshwater and terrestrial environments: Evaluating the current understanding to identify the knowledge gaps and future research priorities. *Sci Total Environ* 586:127–141. doi: 10.1016/j.scitotenv.2017.01.190
- Hu L, Chernick M, Hinton DE, Shi H (2018) Microplastics in Small Waterbodies and Tadpoles from Yangtze River Delta, China. *Environ Sci Technol* 52:8885–8893. doi: 10.1021/acs.est.8b02279
- Huerta Lwanga E, Vega JM, Quej VK, et al (2017) Field evidence for transfer of plastic debris along a terrestrial food chain. 1–7. doi: 10.1038/s41598-017-14588-2
- Hurley R, Woodward J, Rothwell JJ (2018) Microplastic contamination of river beds significantly reduced by catchment-wide flooding. *Nat Geosci* 11:251–257. doi: 10.1038/s41561-018-0080-1
- Imhof AHK, Ivleva NP, Schmid J, et al (2013) Contamination of beach sediments of a subalpine lake with microplastic particles. *Curr Biol* 23:867–868. doi: 10.1016/j.cub.2013.09.001
- Imhof HK, Laforsch C, Wiesheu AC, et al (2016) Pigments and plastic in limnetic ecosystems: A qualitative and quantitative study on microparticles of different size classes. *Water Res* 98:64–74. doi: 10.1016/j.watres.2016.03.015
- Imhof HK, Wiesheu AC, Anger PM, et al (2018) Variation in plastic abundance at different lake beach zones - A case study. *Sci Total Environ* 613–614:530–537. doi: 10.1016/j.scitotenv.2017.08.300
- Kapp KJ, Yeatman E (2018) Microplastic hotspots in the Snake and Lower Columbia rivers: A journey from the Greater Yellowstone Ecosystem to the Pacific Ocean. *Environ Pollut* 241:1082–1090. doi: 10.1016/j.envpol.2018.06.033
- Kataoka T, Nihei Y, Kudou K, Hinata H (2019) Assessment of the sources and inflow processes of microplastics in the river environments of Japan. *Environ Pollut* 244:958–965. doi: 10.1016/j.envpol.2018.10.111
- Klein S, Worch E, Knepper TP (2015) Occurrence and spatial distribution of microplastics in river shore sediments of the rhine-main area in Germany. *Environ Sci Technol* 49:6070–6076. doi: 10.1021/acs.est.5b00492
- Lahens L, Strady E, Kieu-Le TC, et al (2018) Macroplastic and microplastic contamination assessment of a tropical river (Saigon River, Vietnam) transversed by a developing megacity. *Environ Pollut* 236:661–671. doi: 10.1016/j.envpol.2018.02.005
- Lares M, Ncibi MC, Sillanpää M, Sillanpää M (2018) Occurrence, identification and removal of microplastic particles and fibers in conventional activated sludge process and advanced MBR technology. *Water Res* 133:236–246. doi: 10.1016/j.watres.2018.01.049
- Lasee S, Mauricio J, Thompson WA, et al (2017) Microplastics in a freshwater environment receiving treated wastewater effluent. *Integr Environ Assess Manag* 13:528–532. doi: 10.1002/ieam.1915
- Lechner A, Keckeis H, Lumesberger-Loisl F, et al (2014) The Danube so colourful: A potpourri of plastic litter outnumbers fish larvae in Europe's second largest river. *Environ Pollut* 188:177–181. doi: 10.1016/j.envpol.2014.02.006
- Lee H, Kim Y (2018) Treatment characteristics of microplastics at biological sewage treatment facilities in Korea. *Mar Pollut Bull* 137:1–8. doi: 10.1016/j.marpolbul.2018.09.050
- Lee WS, Cho H-J, Kim E, et al (2019) Bioaccumulation of polystyrene nanoplastics and their effect on the toxicity of Au ions in zebrafish embryos. *Nanoscale* 11:3173–3185. doi: 10.1039/C8NR09321K
- Leslie HA, Brandsma SH, van Velzen MJM, Vethaak AD (2017) Microplastics en route: Field measurements in the Dutch river delta and Amsterdam canals, wastewater treatment plants, North Sea sediments and biota. *Environ Int* 101:133–142. doi: 10.1016/j.envint.2017.01.018

- Lin L, Zuo LZ, Peng JP, et al (2018) Occurrence and distribution of microplastics in an urban river: A case study in the Pearl River along Guangzhou City, China. *Sci Total Environ* 644:375–381. doi: 10.1016/j.scitotenv.2018.06.327
- Liu F, Olesen KB, Borregaard AR, Vollertsen J (2019a) Microplastics in urban and highway stormwater retention ponds. *Sci Total Environ* 671:992–1000. doi: 10.1016/j.scitotenv.2019.03.416
- Liu M, Lu S, Song Y, et al (2018) Microplastic and mesoplastic pollution in farmland soils in suburbs of Shanghai, China. *Environ Pollut* 242:855–862. doi: 10.1016/j.envpol.2018.07.051
- Liu X, Yuan W, Di M, et al (2019b) Transfer and fate of microplastics during the conventional activated sludge process in one wastewater treatment plant of China. *Chem Eng J* 362:176–182. doi: 10.1016/j.cej.2019.01.033
- Luo W, Su L, Craig NJ, et al (2018) Comparison of microplastic pollution in different water bodies from urban creeks to coastal waters. *Environ Pollut* 246:174–182. doi: 10.1016/j.envpol.2018.11.081
- Magni S, Binelli A, Pittura L, et al (2019) The fate of microplastics in an Italian Wastewater Treatment Plant. *Sci Total Environ* 652:602–610. doi: 10.1016/j.scitotenv.2018.10.269
- Magnusson K, Eliasson K, Fråne A, et al (2016) Swedish sources and pathways for microplastic to the marine environment: a review of existing data. 88
- Magnusson K, Norén F (2014) Screening of microplastic particles in and down-stream a wastewater treatment plant. *IVL Swedish Environ Res Inst C* 55:22. doi: naturvardsverket-2226
- Mahon AM, O’Connell B, Healy MG, et al (2017) Microplastics in sewage sludge: Effects of treatment. *Environ Sci Technol* 51:810–818. doi: 10.1021/acs.est.6b04048
- Mani T, Hauk A, Walter U, Burkhardt-Holm P (2016) Microplastics profile along the Rhine River. *Sci Rep* 5:17988. doi: 10.1038/srep17988
- Mason SA, Garneau D, Sutton R, et al (2016) Microplastic pollution is widely detected in US municipal wastewater treatment plant effluent. *Environ Pollut* 218:1045–1054. doi: 10.1016/j.envpol.2016.08.056
- McCormick A, Hoellein TJ, Mason SA, et al (2014) Microplastic is an abundant and distinct microbial habitat in an urban river. *Environ Sci Technol* 48:11863–11871. doi: 10.1021/es503610r
- McCormick AR, Hoellein TJ, London MG, et al (2016) Microplastic in surface waters of urban rivers: Concentration, sources, and associated bacterial assemblages. *Ecosphere* 7:. doi: 10.1002/ecs2.1556
- Michielssen MR, Michielssen ER, Ni J, Duhaime MB (2016) Fate of microplastics and other small anthropogenic litter (SAL) in wastewater treatment plants depends on unit processes employed. *Environ Sci Water Res Technol* 2:1064–1073. doi: 10.1039/C6EW00207B
- Miller RZ, Watts AJR, Winslow BO, et al (2017) Mountains to the sea: River study of plastic and non-plastic microfiber pollution in the northeast USA. *Mar Pollut Bull* 124:245–251. doi: 10.1016/j.marpolbul.2017.07.028
- Mintenig SM, Int-Veen I, Löder MGJ, et al (2017) Identification of microplastic in effluents of waste water treatment plants using focal plane array-based micro-Fourier-transform infrared imaging. *Water Res* 108:365–372. doi: 10.1016/j.watres.2016.11.015
- Moore CJ, Lattin GL, Zellers AF (2011) Quantity and type of plastic debris flowing from two urban rivers to coastal waters and beaches of Southern California. *Rev Gestão Costeira Integr* 11:65–73. doi: 10.5894/rgci194
- Morritt D, Stefanoudis P V., Pearce D, et al (2014) Plastic in the Thames: A river runs through it. *Mar Pollut Bull* 78:196–200. doi: 10.1016/j.marpolbul.2013.10.035
- Murphy F, Ewins C, Carbonnier F, Quinn B (2016) Wastewater Treatment Works (WwTW) as a Source of Microplastics in the Aquatic Environment. *Environ Sci Technol* 50:5800–5808. doi: 10.1021/acs.est.5b05416
- Nel HA, Dalu T, Wasserman RJ (2018) Sinks and sources: Assessing microplastic abundance in river sediment and deposit feeders in an Austral temperate urban river system. *Sci Total Environ* 612:950–956. doi: 10.1016/j.scitotenv.2017.08.298
- Peng G, Xu P, Zhu B, et al (2018) Microplastics in freshwater river sediments in Shanghai, China: A case study of risk assessment in mega-cities. *Environ Pollut* 234:448–456. doi: 10.1016/j.envpol.2017.11.034
- Piehl S, Leibner A, Löder MGJ, et al (2018) Identification and quantification of macro- and microplastics on an agricultural farmland. *Sci Rep* 8:17950. doi: 10.1038/s41598-018-36172-y
- Ramos L, Berenstein G, Hughes EA, et al (2015) Polyethylene film incorporation into the horticultural soil of small periurban production units in Argentina. *Sci Total Environ* 523:74–81. doi: 10.1016/j.scitotenv.2015.03.142
- Rech S, Macaya-Caquilpán V, Pantoja JF, et al (2014) Rivers as a source of marine litter - A study from the SE Pacific. *Mar Pollut Bull* 82:66–75. doi: 10.1016/j.marpolbul.2014.03.019

- Rodrigues MO, Abrantes N, Gonçalves FJM, et al (2018) Spatial and temporal distribution of microplastics in water and sediments of a freshwater system (Antuã River, Portugal). *Sci Total Environ* 633:1549–1559. doi: 10.1016/j.scitotenv.2018.03.233
- Scheurer M, Bigalke M (2018) Microplastics in Swiss Floodplain Soils. *Environ Sci Technol* 52:3591–3598. doi: 10.1021/acs.est.7b06003
- Shruti VC, Jonathan MP, Rodriguez-Espinosa PF, Rodríguez-González F (2019) Microplastics in freshwater sediments of Atoyac River basin, Puebla City, Mexico. *Sci Total Environ* 654:154–163. doi: 10.1016/j.scitotenv.2018.11.054
- Sighicelli M, Pietrelli L, Lecce F, et al (2018) Microplastic pollution in the surface waters of Italian Subalpine Lakes. *Environ Pollut* 236:645–651. doi: 10.1016/j.envpol.2018.02.008
- Simon M, van Alst N, Vollertsen J (2018) Quantification of microplastic mass and removal rates at wastewater treatment plants applying Focal Plane Array (FPA)-based Fourier Transform Infrared (FT-IR) imaging. *Water Res* 142:1–9. doi: 10.1016/j.watres.2018.05.019
- Sruthy S, Ramasamy E V. (2017) Microplastic pollution in Vembanad Lake, Kerala, India: The first report of microplastics in lake and estuarine sediments in India. *Environ Pollut* 222:315–322. doi: 10.1016/j.envpol.2016.12.038
- Su L, Xue Y, Li L, et al (2016) Microplastics in Taihu Lake, China. *Environ Pollut* 216:711–719. doi: 10.1016/j.envpol.2016.06.036
- Talvitie J, Heinonen M, Pääkkönen JP, et al (2015) Do wastewater treatment plants act as a potential point source of microplastics? Preliminary study in the coastal Gulf of Finland, Baltic Sea. *Water Sci Technol* 72:1495–1504. doi: 10.2166/wst.2015.360
- Talvitie J, Mikola A, Koistinen A, Setälä O (2017a) Solutions to microplastic pollution – Removal of microplastics from wastewater effluent with advanced wastewater treatment technologies. *Water Res* 123:401–407. doi: 10.1016/j.watres.2017.07.005
- Talvitie J, Mikola A, Setälä O, et al (2017b) How well is microlitter purified from wastewater? – A detailed study on the stepwise removal of microlitter in a tertiary level wastewater treatment plant. *Water Res* 109:164–172. doi: 10.1016/j.watres.2016.11.046
- Tan X, Xubiao Yu, Cai L, et al (2019) Microplastics and associated PAHs in surface water from the Feilaixia Reservoir in the Beiji River, China. *Chemosphere* 221:834–840. doi: 10.1016/j.chemosphere.2019.01.022
- Tibbetts J, Krause S, Lynch I, Smith GHS (2018) Abundance, distribution, and drivers of microplastic contamination in urban river environments. *Water (Switzerland)* 10:. doi: 10.3390/w10111597
- van Emmerik T, Kieu-Le T-C, Loozen M, et al (2018) A Methodology to Characterize Riverine Macroplastic Emission Into the Ocean. *Front Mar Sci* 5:1–11. doi: 10.3389/fmars.2018.00372
- Vaughan R, Turner SD, Rose NL (2017) Microplastics in the sediments of a UK urban lake. *Environ Pollut* 229:10–18. doi: 10.1016/j.envpol.2017.05.057
- Vermaire JC, Pomeroy C, Herczegh SM, et al (2017) Microplastic abundance and distribution in the open water and sediment of the Ottawa River, Canada, and its tributaries. *Facets* 2:301–314. doi: 10.1139/facets-2016-0070
- Vollertsen J (ed), Hansen AA (ed) (2017) *Microplastic in Danish wastewater: Sources, occurrences and fate*
- Wang J, Peng J, Tan Z, et al (2017a) Microplastics in the surface sediments from the Beiji River littoral zone: Composition, abundance, surface textures and interaction with heavy metals. *Chemosphere* 171:248–258. doi: 10.1016/j.chemosphere.2016.12.074
- Wang W, Ndungu AW, Li Z, Wang J (2017b) Microplastics pollution in inland freshwaters of China: A case study in urban surface waters of Wuhan, China. *Sci Total Environ* 575:1369–1374. doi: 10.1016/j.scitotenv.2016.09.213
- Wang W, Yuan W, Chen Y, Wang J (2018) Microplastics in surface waters of Dongting Lake and Hong Lake, China. *Sci Total Environ* 633:539–545. doi: 10.1016/j.scitotenv.2018.03.211
- Wen X, Du C, Xu P, et al (2018) Microplastic pollution in surface sediments of urban water areas in Changsha, China: Abundance, composition, surface textures. *Mar Pollut Bull* 136:414–423. doi: 10.1016/j.marpolbul.2018.09.043
- Williams AT, Simmons SL (1999) Sources of riverine litter: The river Taff, South Wales, UK. *Water Air Soil Pollut* 112:197–216. doi: 10.1023/A:1005000724803
- Wolff S, Kerpen J, Prediger J, et al (2018) Determination of the Microplastics Emission in the Effluent of a Municipal Waste Water Treatment Plant using Raman Microspectroscopy. *Water Res X* 2:100014. doi: 10.1016/J.WROA.2018.100014
- Xiong X, Wu C, Elser JJ, et al (2019) Occurrence and fate of microplastic debris in middle and lower reaches of the Yangtze River – From inland to the sea. *Sci Total Environ* 659:66–73. doi: 10.1016/j.scitotenv.2018.12.313
- Yan M, Nie H, Xu K, et al (2019) Microplastic abundance, distribution and composition in the Pearl

- River along Guangzhou city and Pearl River estuary, China. *Chemosphere* 217:879–886. doi: 10.1016/j.chemosphere.2018.11.093
- Yonkos LT, Friedel EA, Perez-Reyes AC, et al (2014) Microplastics in Four Estuarine Rivers in the Chesapeake Bay, U.S.A. *Environ Sci Technol* 48:14195–14202. doi: 10.1021/es5036317
- Yuan W, Liu X, Wang W, et al (2019) Microplastic abundance, distribution and composition in water, sediments, and wild fish from Poyang Lake, China. *Ecotoxicol Environ Saf* 170:180–187. doi: 10.1016/j.ecoenv.2018.11.126
- Zbyszewski M, Corcoran PL (2011) Distribution and degradation of fresh water plastic particles along the beaches of Lake Huron, Canada. *Water Air Soil Pollut* 220:365–372. doi: 10.1007/s11270-011-0760-6
- Zbyszewski M, Corcoran PL, Hockin A (2014) Comparison of the distribution and degradation of plastic debris along shorelines of the Great Lakes, North America. *J Great Lakes Res* 40:288–299. doi: 10.1016/j.jglr.2014.02.012
- Zhang GS, Liu YF (2018) The distribution of microplastics in soil aggregate fractions in southwestern China. *Sci Total Environ* 642:12–20. doi: 10.1016/j.scitotenv.2018.06.004
- Zhang K, Gong W, Lv J, et al (2015) Accumulation of floating microplastics behind the Three Gorges Dam. *Environ Pollut* 204:117–123. doi: 10.1016/j.envpol.2015.04.023
- Zhang K, Su J, Xiong X, et al (2016) Microplastic pollution of lakeshore sediments from remote lakes in Tibet plateau, China. *Environ Pollut* 219:450–455. doi: 10.1016/j.envpol.2016.05.048
- Zhang S, Yang X, Gertsen H, et al (2018) A simple method for the extraction and identification of light density microplastics from soil. *Sci Total Environ* 616–617:1056–1065. doi: 10.1016/j.scitotenv.2017.10.213
- Ziajahromi S, Neale PA, Rintoul L, Leusch FDL (2017) Wastewater treatment plants as a pathway for microplastics: Development of a new approach to sample wastewater-based microplastics. *Water Res* 112:93–99. doi: 10.1016/j.watres.2017.01.042
- Zubris KA V, Richards BK (2005) Synthetic fibers as an indicator of land application of sludge. *Environ Pollut* 138:201–211. doi: 10.1016/j.envpol.2005.04.013
- Zylstra ER (2013) Accumulation of wind-dispersed trash in desert environments. *J Arid Environ* 89:13–15. doi: 10.1016/j.jaridenv.2012.10

