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1 **Recycling of End-of-life reverse osmosis membranes**
2 **for Membrane Biofilms Reactors (MBfRs). Effect of**
3 **chlorination on the membrane surface and gas**
4 **permeability.**

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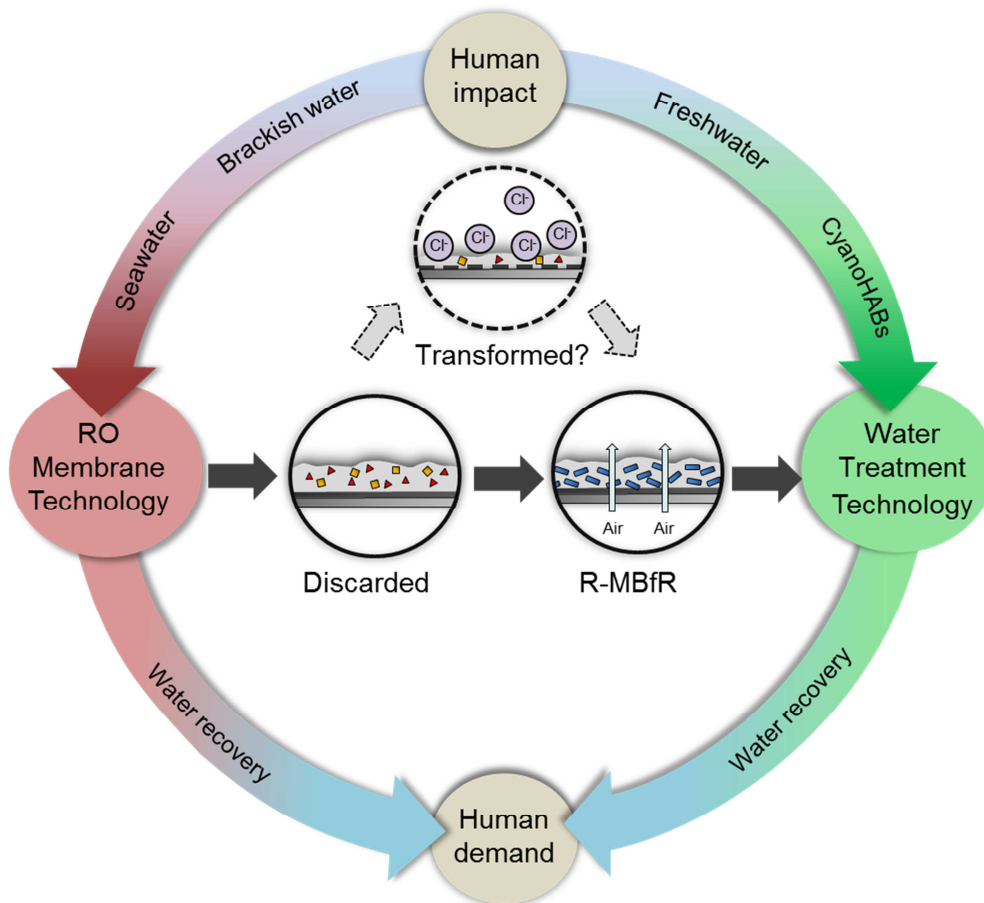
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17 **Keywords:** Recycled membranes, transformation process, chlorination, fouling,
18 biofilm, microcystin, degradation, gas permeability.

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21 **Graphical Abstract**

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30 **Abstract**

31 Reducing human impacts on drinking water is one of the main challenges for
32 the water treatment industry. This work provides new results to support the
33 recycling of EoL desalination reverse osmosis (RO) membranes for Membranes
34 Biofilm Reactors (MBfRs). We investigate if the controlled-removal of fouling
35 and polyamide layer may favor the use of these membranes in MBfRs. It also
36 would allow establishing a normalized methodology of membrane recycling,
37 regardless of inherited fouling during its lifespan. For this purpose, we transform
38 by chlorination discarded brackish (BWd) and seawater (SWd) membranes into
39 nanofiltration (BWt-NF and SWt-NF) and ultrafiltration (BWt-UF and SWt-UF)
40 membranes. Our results show that chlorine attacks allow the fouling cleaning
41 while improves the hydrophilicity and maintains roughness only in BWt-NF.
42 Therefore, the bacterial deposition in this membrane is greater than the other
43 tested membranes. Besides, the microcystin (MC) degradation capacity of BWt-
44 NF verifies the compatibility of the chemical modification for the biological
45 activity of MC-degrading bacteria. Finally, our results also provide that
46 polyamide thin-film composite (PA-TFC) membranes, originally manufactured
47 for salt rejection during desalination processes, offer competitive gases diffusion
48 at low pressures. Therefore, we conclude that the membrane recycling may
49 provide alternative low cost and gas permeable membranes for MBfRs,
50 according to circular economy principles.

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54 1. Introduction

55 Water is essential for human life and plays an important role in many sectors of
56 the economy. The strong anthropogenic impact on freshwater worldwide has
57 led to global concern about water supply (Paerl and Paul, 2012; Vörösmarty et
58 al., 2000). The ever-growing human population, and increasing water pollution
59 and freshwater demands, have allowed major advances to be made in water
60 treatment processes (Ercin and Hoekstra, 2014; M.Mekonnen and Hoekstra,
61 2016). Membrane technology is among the most important fields in the
62 separation processes employed for water treatment. The use of membrane
63 separation technology has several advantages: continuous operation, adjusted
64 properties of membranes to the requirements set in each application, flexibility
65 in systems design and easy scaling-up. Therefore, membrane technology has
66 considerably developed in recent decades to establish a solid, mature and
67 standardized market.

68 Pressure-driven membranes are well-established desalination processes by
69 which freshwater can be obtained from seawater and brackish water (Van Der
70 Bruggen et al., 2003). In particular, reverse osmosis (RO) membranes
71 constitute the most used desalination technology worldwide. Over 95% of
72 existing RO desalination plants use polyamide thin film composite (PA-TFC)
73 membranes, a high-performance material with excellent mechanical and
74 chemical durability (Geise et al., 2010). Consequently, the large desalination
75 market has resulted in increased waste generation associated with this
76 technology, which has led to the disposal of more than 840,000 end-of-life (EoL)
77 membranes (>14,000 Tn/year) every year worldwide (Landaburu-Aguirre et al.,
78 2016; Peng Lee et al., 2011). The landfilling or incineration of millions of

79 membranes is currently the fastest solution for a material considered to be
80 waste. Therefore, following European Directive 2008/98/EC on waste, which
81 sets out a hierarchy of priorities for waste treatment (prevention, reuse,
82 recycling, other types of recovery and, finally, disposal), current membrane
83 management is not coherent with the basic principles of European
84 environmental legislation; hence, new less environmentally harmful handling
85 alternatives are desired to move toward a circular economy system and to
86 achieve a cross-continental recycling society (Darton and Fazel, 2001;
87 European commission, 2008; Lawler et al., 2015; Siddique et al., 2008).

88 One of the main factors that affects RO membrane discarding is biofouling
89 (Nguyen et al., 2012), a serious issue that compromises the membrane filtration
90 process by shortening membrane service life (5-10 years) and raising its
91 replacement rate (10-15% per year) (Darton and Fazel, 2001; Landaburu-
92 Aguirre et al., 2016). Consequently, different alternatives have been proposed
93 to extend the lifespan of discarded membranes. One such alternative is to
94 recycle discarded RO membranes by removing the active polyamide (PA) layer,
95 which is an interesting solution (Lawler et al., 2013). This transformation
96 process is based on chemical modification using oxidant agents such as
97 K_2MnO_4 and NaOCl which, under controlled conditions, convert EoL RO
98 membranes into reusable nanofiltration (NF) or ultrafiltration (UF) membranes
99 (Ambrosi and Tessaro, 2013; García-Pacheco et al., 2015; Raval et al., 2012).
100 However, not all discarded RO membranes can be recycled and reused as
101 pressure-driven membranes because the transformation success is dependent
102 on the nature of PA (Do et al., 2012a, 2012b).

103 Membrane biofilm reactors (MBfRs) are an emerging membrane technology
104 and have been recently used to overcome the growing need for treating poor
105 quality water. To date, MBfRs have tested porous, dense and composite
106 membranes (Martin et al., 2012). However, the role of the membrane in MBfRs
107 differs from that in RO membranes and membrane bioreactors (MBRs) because
108 they neither act as filters (Li and Zhang, 2018; Nerenberg, 2016, 2005), nor are
109 they classical reactors as they provide new functions to surfaces, beyond that of
110 mere supports material (Halan et al., 2012). MBfRs use gas-permeable
111 membranes to remove pollutants from water by supplying electron donors or
112 acceptors across the membrane, with the aim of enhancing the biological
113 activity of the bacterial biofilm immobilized on the membrane (Li et al., 2008;
114 Martin and Nerenberg, 2012; Syron and Casey, 2008; Zhou et al., 2019).
115 Interestingly, companies such as APTwater, Inc. (Long Beach, CA, USA) have
116 developed similar membrane configurations to RO units (Martin and Nerenberg,
117 2012). So even though membrane functions differ among these technologies,
118 similar compositions and layouts can be used.

119 In previous studies, we proposed a novel recycling option for discarded RO
120 membranes by converting them into low-cost thin-film composite support
121 material for MBfRs (Morón-López et al., 2019). We proved that discarded RO
122 membranes could be directly used as surfaces for immobilizing bacteria and
123 carry out a desired reaction. Even we observed that the intrinsic fouling of
124 discarded membranes may act as a conditioning agent and increase the
125 bacterial deposition rate. In this manner, we linked the waste from pressure-
126 driven membranes to MBfR technology for the first time. The biological activity
127 of the biofilm generated on the discarded desalination membrane was proven

128 by removing microcystins (MC), a group of potent hepatotoxins produced by
129 harmful cyanobacterial algal blooms (cyanoHABs) in eutrophic water (Buratti et
130 al., 2017; Dawson, 1998; Lawton and Robertson, 1999). For this reason,
131 although MBfRs have been successfully used to remove a wide variety of
132 wastewater pollutants (Casey et al., 1999; Ontiveros-Valencia et al., 2018), we
133 also demonstrated that our recycled membrane biofilm reactor (R-MBfR) is a
134 suitable method to remove pollutants from surface water.

135 The present work attempted to go one step further by evaluating the effect of
136 the transformation process through chlorination on discarded RO membranes.
137 This process could be also interesting for recycling membranes into MBfRs due
138 to both its simultaneous gas permeability enhancing, after partial or total
139 removal of the polyamide (PA) layer, and the cleaning effect that chlorine may
140 provide by removing the previous fouling, which is a conditioning agent but
141 highly dependent from the water source used. However, the use of chlorine on
142 the surfaces may also modify the membrane characteristics and consequently,
143 the bacterial deposition. Therefore, this study is focused on investigate if: a)
144 chlorination changes key surface characteristics for conditioning and bacterial
145 association, such as previous fouling presence, charge, contact angle and
146 surface roughness; b) the MC-degrading bacteria is able to attach on
147 transformed surfaces and carry out the biological activity; c) the PA removal of
148 the discarded membranes influences on gas permeability, adding value to the
149 transformation process. All these newly acquired characteristics were compared
150 with those already studied in untransformed discarded RO membranes in the
151 previous work (Morón-López et al., 2019). So we provide new knowledge for

152 optimizing the R-MBfR concept by opening up new horizons for future
153 membrane technology applications as part of circular economy principles.

154 **2. Material and Method**

155 *2.1 Membranes and chemical reagents*

156 Experiments were performed on the membranes obtained from EoL PA-TFC
157 RO membranes. For this purpose, membrane coupons (216 cm²) were taken
158 from 8 inch-diameter spiral wound modules. The EoL membranes had originally
159 been used for water desalination for more than 3 years and presented fouling of
160 different natures. On the one hand, the TM 720-400 (*Toray*) module had treated
161 brackish water (BW). On the other hand, the HSWC3 (*Hydranautics*) module
162 had treated seawater (SW). The end-of-life RO membranes were transformed in
163 recycled NF and UF membranes using two different dose level of NaOCl. All the
164 membranes were conserved in Milli-Q water before being analyzed. NaOCl
165 (10%) was purchased from Scharlab and used to transform the EoL RO
166 membranes.

167 *2.2. Transformation process of the EoL RO membranes*

168 The EoL RO membranes were washed out with Milli-Q water, followed by
169 exposing them to a dose level of 6,200 ppm·h and 300,000 ppm·h of NaOCl at
170 pH 10, for 24 h at room temperature and under static conditions to obtain NF
171 and UF membranes, respectively. These dose levels were selected based on
172 the previous confirmation that they fell within a proper dose range for converting
173 EoL RO membranes into NF and UF recycled membranes (García-Pacheco et
174 al., 2015; Molina et al., 2018, 2015). Thus the obtained membranes were: BW
175 transformed into NF (BWt-NF) and into UF (BWt-UF); SW transformed into NF

176 (SWt-NF) and UF (SWt-UF). After these transformation processes, coupons
177 were taken out of the containers and thoroughly washed with Milli-Q water until
178 a pure water pH was accomplished. For comparison purposes, the same non
179 transformed type membranes (BWd and SWd) were also included (Morón-
180 López et al., 2019). All the membranes are kept in Milli-Q water until used.

181 *2.3. Membrane surface characterization*

182 Similar experimental set-up and analyzing methodology to our previous work
183 were performed in order to compare with discarded untransformed membranes
184 results (Morón-López et al., 2019). For membrane characterization, all the
185 membranes were previously dried at 100 °C for 24 h to avoid the interaction of
186 occluded water.

187 *2.3.1. Zeta potential measurements*

188 A Surface Zeta Potential Cell (ZEN 1020, Malvern) was employed to measure
189 the surface Zeta potential via electrophoretic light scattering (Zetasizer Nano
190 ZS), under the same conditions applied in previous studies (Morón-López et al.,
191 2019; Santiago-Morales et al., 2016).

192 *2.3.2. Contact angle measurement*

193 Surface wettability tests were performed by a static contact angle meter via the
194 sessile drop technique in a KSV CAM200 instrument (KSV Instruments, USA),
195 under identical conditions as in previous studies (Molina et al., 2015; Morón-
196 López et al., 2019).

197 *2.3.3. Atomic-force microscopy (AFM)*

198 Following the same experimental conditions of previous works (Molina et al.,
199 2018; Morón-López et al., 2019), atomic-force microscopy (AFM) experiments

200 for the roughness analysis were carried out in the tapping mode by a Multimode
201 AFM (Veeco Instruments, Santa Barbara, CA, USA), equipped with a
202 Nanoscope Iva control system (software version 6.14r1).

203 *2.4. Bacterial deposition test*

204 MC-degrading bacteria *Sphingopyxis* sp. strain IM-1 has been proven to
205 efficiently degrade the MC molecule until small peptides and amino acids
206 (Jones et al., 1994; Lezcano et al., 2016). For this reason, this strain was
207 selected to analyze its attachment to membranes BWt-NF, BWt-UF, SWt-NF
208 and SWt-UF. The bacterial growth on tested membranes was performed by
209 following the steps previously described (Morón-López et al., 2019). Then,
210 samples were analyzed by confocal laser scanning microscopy (CLSM) and
211 scanning electron microscopy (SEM). The negative controls without strain IM-1
212 were also performed for the comparisons.

213 *2.5. Confocal laser scanning microscopy (CLSM)*

214 The bacterial coverage images were obtained with a confocal laser scanning
215 microscope (CLSM Leica SP5, Leica Microsystems), using the same
216 experimental conditions reported in a previous study (Morón-López et al.,
217 2019). The bacterial coverage percent (%) on the transformed membrane
218 surfaces was analyzed by the ImageJ software (Abràmoff et al., 2004).

219 *2.6. Scanning electron microscopy (SEM)*

220 Several imaging devices have been used. The first (XL30 ESEM Model
221 (Phillips)) was used to observe the cross-section of membranes. For this
222 purpose, membranes were broken properly after being frozen in liquid nitrogen.

223 Samples were dried and then gold-sputtered with a Sputter Coater Polaron
224 SC7640 model to achieve a 13-15 nm thickness prior to the SEM analysis.
225 Second, two other SEMs were employed to examine the surfaces of the
226 membranes. The transformed membrane surfaces were observed by an S-8000
227 Model (Hitachi) device. A SEM Zeiss DSM 950 (Germany) was used to observe
228 the bacterial attachment of BWt-NF after the bacterial deposition test, following
229 the steps previously reported (Morón-López et al., 2019).

230 *2.7. MC degradation test*

231 Toxins were obtained by extracting the fresh cyanobacterial scum collected
232 from natural water (Lezcano et al., 2016). The MC mixture was composed of
233 different variants: 84.5% of -LR; 9.86 % of -RR and 5.64 of -YR.

234 Having completed the bacterial deposition test, we chose the membrane with
235 the highest bacterial coverage for the MC degradation test. After growing the
236 MC-degrading bacteria, the MC degradation test of selected membrane was
237 performed under the same experimental conditions previously reported (Morón-
238 López et al., 2019). The controls with the same membrane and conditions, but
239 no bacteria, were also analyzed.

240 According to the reference (Morón-López et al., 2017), an HPLC-MS-TOF
241 (Agilent 6230 accurate mass TOF Agilent Technologies, Santa Clara, CA, USA)
242 was used to quantify the MC concentration. After that, the MC degradation
243 kinetics of the bacteria attached to the transformed membrane was calculated
244 following previous works (Li et al., 2014; Xu et al., 2011). Because of a lag
245 phase was observed, calculates for determining k constant were done taking
246 the first point at which biodegradation began (Ho et al., 2012).

247 2.8. Gas permeability test

248 The air permeability of the studied membranes was investigated with an MBfR
249 cell by measuring the air flow across the membrane by a bubble flowmeter at
250 room temperature and pressure at 1 bar. For the permeability calculations, a 0.2
251 μm membrane thickness was taken into account for the discarded and
252 transformed NF types, while the thickness of the transformed UF was 50, and
253 was 30 μm for the BW model and the SW model, respectively. All the
254 membranes measurements were taken in triplicate using different parts of the
255 membrane coupons. Given the dissimilar ways to show the gas permeability of
256 the membranes found in the literature, air permeability was calculated in
257 $\text{Ncm}^3 \cdot \text{cm} \cdot \text{cm}^{-2} \cdot \text{s}^{-1} \cdot \text{cmHg}^{-1}$ (Barrers) and air flux in $\text{Ncm}^3 \cdot \text{cm}^{-2} \cdot \text{s}^{-1} \cdot \text{cmHg}^{-1}$ (J).
258 Additionally, to provide more information of tested membranes for other
259 hypothetical applications, the pure hydrogen permeability was also tested
260 following the same steps and criteria commented above.

261 2.9. Data analysis

262 Statistical analyses were conducted using the Statistical Package for Social
263 Sciences (SPSS, Inc.) software, v.17. Normality and homogeneity of variances
264 were performed using the Shapiro-Wilk and Levene tests, respectively. To
265 determine significant differences between bacterial coverage and zeta potential
266 a one-way analysis of variance (ANOVA) and a Turkey HSD analysis were run.
267 One-way ANOVA and Tamhane *post hoc* analyses were performed for
268 remaining variables with non-homogeneous variances.

269 3. Results and discussion

270 3.1. *Characterization of the transformed EoL RO membranes*

271 The membranes used in this study were EoL RO polyamide thin film composite
272 (PA-TFC) membranes because of many existing discarded modules (Geise et
273 al., 2010). PA-TFC membranes are composed for a three-layer structure,
274 typically based on a non-woven fibrous support, a porous polysulfone (PSF)
275 sublayer and a dense polyamide (PA) ultrathin surface. To transform the EoL
276 discarded membranes, which came from treating brackish water (BWd type)
277 and seawater (SWd type), we chemically attacked the membrane surface with
278 NaOCl. This process removes not only previously attached fouling, but also the
279 PA layer in a controlled manner to obtain transformed NF and UF membranes
280 (BWt-NF, BWt-UF, SWt-NF and SWt-UF types) from the discarded types (Kwon
281 and Leckie, 2006; Rodríguez et al., 2002).

282 The chemical attack with 6,200 ppm·h of NaOCl partially removed the PA layer,
283 and BWt-NF and SWt-NF membranes were obtained (Fig. 1). Conversely with
284 the BWt-UF and SWt-UF membranes obtained after treating them with
285 300,000 ppm·h of NaOCl, PA removal was total, and a porous surface which
286 belonged to the PSF layer was observed. In both cases, a cleaning effect was
287 achieved and no fouling was noted. These results agree with authors who
288 generated recycled NF and UF membranes using the same NaOCl doses in
289 previous works (García-Pacheco et al., 2015; Molina et al., 2018).

290 The transformation process by chlorination also modifies key surface
291 characteristics for bacterial attachment, such as charges, hydrophobicity and
292 roughness (Table 1). Zeta potential measurements showed a negatively
293 charged surface after chlorine attack on all surfaces, as in other studies (Do et

294 al., 2012a; Kwon and Leckie, 2006; Xu et al., 2013). Contact angle changes
295 were also observed after chlorine transformation. The transformed membranes
296 that belonged to the SW model were more hydrophobic than those for the BW
297 model, and the most hydrophilic surface was observed in BWt-NF. These
298 results agree with studies in which the incorporation of chlorine onto the surface
299 led to greater hydrophobicity (Kwon and Leckie, 2006; Simon et al., 2009), while
300 other studies have shown that chlorine attack triggers hydrophilic surfaces (Do
301 et al., 2012a; Molina et al., 2018). Besides, fouling removal seems to more
302 strongly affect the contact angle of the SW model, possibly due to its more
303 organic nature (Morón-López et al., 2019).

304 In spite of changes in the aforementioned properties, the most obvious surface
305 alteration between membrane types was roughness (Table 1, Fig. 2). Average
306 roughness (R_a , the average deviation of the peaks and valleys from the mean
307 height) and root mean square roughness (R_q , standard deviation of the peaks
308 and valleys) showed that the roughness of both membrane models significantly
309 decreased at higher chlorine doses (BWt-UF and SWt-UF). This significant drop
310 in roughness could be due to the PA layer being removed (Al-Jeshi and Neville,
311 2006; Jiang et al., 2018; Molina et al., 2018). The fact that roughness remained
312 similar for BWt-NF and BWd, but significantly lowered more in SWt-NF than
313 SWd, was remarkable, and agrees with the works which have indicated that
314 modification by chlorination is PA nature-dependent (Do et al., 2012b; Molina et
315 al., 2015). It also suggests that fouling nature triggers more roughness on SWd
316 than BWd and, hence, its removal affects SWd roughness more.

317 *3.2. Conditioning and bacterial attachment*

318 A biofilm is referred to as a surface-associated microbial community enclosed
319 by extracellular polymeric substances (EPS). Biofilm development is a complex
320 process whose stages comprise a first substratum conditioning, initial bacterial
321 attachment, following by biofilm formation and bacterial dispersion when biofilm
322 matures (Characklis and Cooksey, 1983). In order to identify if the
323 transformation process affected to conditioning and bacterial deposition, and
324 therefore for biofilm formation, we focused on early, but no less important,
325 stages where the surface plays a relevant role (Fig. 3). For this purpose, we
326 immersed the transformed membranes into a culture of MC-degrading bacteria
327 *Sphingopyxis* sp. strain IM-1. Worth mentioning that although MBfRs are
328 composed of multi-species biofilms, the MC degradation capacity has been
329 confined to limited number of species so far (Li et al., 2017). For this reason, as
330 well as to avoid possible interferences in biological activity interpretation due to
331 interactions between microorganisms, we have used single specie as MC-
332 degrading biofilm. Additionally, because of the cell deposition is dependent of
333 interaction between cells and surfaces (Donlan, 2002), we also included
334 Calcium (Ca^{2+}) as a *quorum-sensing* molecule and a surface conditioner (He et
335 al., 2016; Li and Elimelech, 2004; Mangwani et al., 2014).

336 Unlike the untransformed discarded membranes, the cleaning of the surface
337 during the transformation process could lead to direct interaction between Ca^{2+}
338 and the dissolved nutrients with the surface without fouling, as outlined in Figure
339 3. We observed that the BWt-NF membrane was the only membrane favorable
340 for conditioning, and showed the best bacterial attachment in the CLMS images
341 (Fig. 4). This membrane presented a similar roughness, but a smaller contact
342 angle, than SWt-NF. Hence, hydrophilicity could be considered the beneficial

343 factor for bacterial attachment because water-miscible conditioners have better
344 access of to the surface (Characklis and Wilderer, 1989; Hou et al., 2013).
345 Unlike SWt-NF, BWt-UF and SWt-UF did not show any bacterial deposition,
346 possibly due to their drastic decrease in roughness, and also to the
347 hydrophobicity of their surfaces. Even the least negative charge of BWt-UF
348 could affect its low bacterial coverage as a result of its minor attraction to Ca^{2+}
349 and other nutritive substances (Fang et al., 2018). All these results agree with
350 other studies in which hydrophilic and roughness surfaces enhanced bacterial
351 deposition (Chen et al., 2013; Guo et al., 2013; Subramani et al., 2009). They
352 are also consistent with those authors who conclude that not all cultures
353 successfully colonize certain membrane types (Rothmund et al., 1996).

354 Compared to the untransformed discarded membranes, the amount of bacterial
355 attached to the BWt-NF surface was significantly higher than BWd and similar
356 to SWd (Fig. 4). Therefore, these results suggest that, at certain doses of
357 NaOCl, the chlorination process could simultaneously remove fouling and
358 improve the key surface characteristics for conditioning and bacterial deposition
359 in some membranes (Fig. 3). Chlorine could interact with certain types of PAs to
360 result in a more hydrophilic active-charged surface for Ca^{2+} and nutrient
361 absorption (Herzberg et al., 2009; Li and Elimelech, 2004). In this manner,
362 transforming discarded membranes could be advantageous for avoiding the
363 highly variable fouling nature of untransformed discarded types, which could be
364 relevant to establish a hypothetical methodology of recycling. On the other
365 hand, in reference to the worst results obtained in remaining membranes, it
366 should be noted that this prior conditioning to the initial bacterial attachment
367 could also modify these properties with time (Jin et al., 2009; Zhao et al., 2015).

368 Hence, mid- and long-term studies are necessary to confirm the rejection of
369 some membrane types for biofilm formation.

370 3.3. MC degradation capability

371 Once the strain IM-1 was attached to the transformed surfaces in BWt-NF, an
372 MC degradation test was run to observe if the chlorine modifications affected its
373 biological activity. Figure 5.1 shows that, after an acclimation time in the
374 presence of the toxin, strain IM-1 was able to remove the total MC
375 concentration ($1 \text{ mg}\cdot\text{L}^{-1}$) in 6 h. Besides, no MC removal was observed in the
376 negative control without bacteria (Fig. 5.1 and 5.2.b), which indicates that MC
377 degradation was biologically-mediated. The MC degradation kinetics follows the
378 pseudo-first order model, where k and $T_{1/2}$ agree with those obtained previously
379 with untransformed discarded membranes (Morón-López et al., 2019).
380 Therefore, it could be concluded that the chemical modification during the
381 transformation process maintains viable conditions for biological reactions in
382 certain EoL membranes. Nonetheless, we also highlighted that the potential use
383 of membranes transformed into MBfRs would still be less appealing than using
384 untransformed discarded membranes due to the environmental impacts derived
385 from the transformation process (Nguyen et al., 2012). Consequently, further
386 experiments and techno-economic and environmental analysis are needed to
387 clarify the advantages and disadvantages of transforming discarded
388 membranes in larger scales.

389 3.4. Gas permeability

390 One of the most crucial aspects in MBfRs is membrane typology due to
391 differences in each membrane's gas transfer properties (Li et al., 2008;

392 Nerenberg, 2016; Syron and Casey, 2008; Tang et al., 2012). The gas transfer
393 resistance of these membranes depends on the material's character and
394 thickness. Porous membranes provide high gas transfer rates given the faster
395 gaseous diffusion in gas-filled pores (Semmens, 2005). However, drawbacks
396 such as a low bubble point, clogging and wetting are some of the limitations of
397 porous materials (Casey et al., 1999). Conversely, dense membranes are free
398 of clogging and wetting, but their gas transfer is lower than porous membranes
399 because gas is firstly dissolved through the membrane's material. To overcome
400 this diffusion resistance, dense membranes work at high intermembrane
401 pressure. As an alternative, a thinner dense layer achieves less mass
402 resistance. Hence, composite membranes offer a promising, but more
403 expensive, option than others (Martin and Nerenberg, 2012).

404 Using recycled desalination composite membranes for MBfRs could be an
405 interesting low-cost option to achieve the desired throughputs. To study if the
406 untransformed and transformed membranes allowed gas permeability, we
407 investigated air permeability before and after dense PA layer removal. As
408 shown in Table 2, all tested membranes were able to offer air flux at low
409 pressure. Consequently, up to our knowledge, we proved for first time that
410 membranes originally manufactured for desalination purposes are capable of
411 diffusing gases from the inner non-woven fibrous layer. In addition, unequally
412 fluxes were obtaining depending on the chlorine doses used during the
413 transformation process. The PSF porous BWt-UF and SWt-UF have an air
414 permeability that is between two and three orders of magnitude higher than the
415 untransformed discarded membranes and those transformed into NF.
416 Conversely, similar air permeability was observed between the untransformed

417 discarded membranes and those transformed into NF. Therefore, as expected,
418 our results suggested that total PA removal from discarded membranes would
419 lead to higher air permeability. However, no increase of gas permeability is
420 achieved at doses levels of 6,200 ppm·h of NaOCl. Besides, our results also
421 indicated higher air permeability in the BW model than in the SW model. This
422 could be due to the difference in selective layer thickness. Figure 1 shows the
423 cross-section micrographs of the PSF layer in the transformed membranes. The
424 thickness of the recycled membranes used originally as the RO BW membranes
425 almost doubled those which treated seawater. This variation in thickness could
426 be due to differences in membrane compaction, as BW membranes have been
427 subjected to lower pressure than SW membranes during their previous service
428 times in desalination plants (Molina et al., 2018).

429 When comparing the air permeability of the tested membranes with the
430 membranes made especially for gas transfer from studies based on MBfRs, we
431 obtained promising results. We observed that SWd and SWt-NF were of the
432 same order of magnitude as the commercial polyethylene composite membrane
433 used by (Ahmed et al., 2004), while the BWd and BWt-NF membranes were
434 160-fold more permeable. The composite polyolefin membrane of (Motlagh et
435 al., 2008), and that commercialized for air separation made of Teflon of
436 Cerqueira et al., (2013), showed an oxygen flux of $3.00 \cdot 10^{-5}$ and $4.91 \cdot 10^{-4}$
437 $\text{Ncm}^3 \cdot \text{cm}^{-2} \cdot \text{s}^{-1} \cdot \text{cmHg}^{-1}$, respectively. Otherwise, the comparison made of the
438 polyurethane composite membranes by Tang et al., (2012) displayed lower
439 oxygen fluxes than previous references. Therefore, our recycled composite
440 membranes were far more permeable than those in other studies, which
441 suggests that these membranes could be a green low-cost alternative for

442 MBfRs. Worth mentioning that pure hydrogen permeability of these membranes
443 was also tested (Table S1) and the hydrogen fluxes obtained were higher than
444 those composite membranes permeabilities reported by Tang et al., (2012).
445 Accordingly, although the hydrogen application is not required for removing MC
446 by an aerobic biofilm, our preliminary data suggest that recycled membranes
447 could also be appropriate for other uses of hydrogen-based MBfRs.

448 By focusing on transformed to porous membranes (BWt-UF and SWt-UF), the
449 total PA removal also provided higher air fluxes than those of the
450 aforementioned composite membranes. However, when compared with the
451 oxygen permeabilities of the other porous materials used in MBfRs, such as
452 polyethylene ($8.00 \cdot 10^{-1}$ Barrers), polypropylene ($2.00 \cdot 10^{-1}$ Barrers), polyvinyl
453 chloride ($1.40 \cdot 10^{-2}$ Barrers), polyvinyl alcohol ($1.00 \cdot 10^{-2}$ Barrers) and
454 polyvinylidene fluoride ($3.00 \cdot 10^{-3}$ Barrers), our porous membranes were less
455 permeable by several orders of magnitude (Cote, 1989; Hou et al., 2013).
456 These results could be due to the nature of the PSF of the transformed
457 membranes. Therefore, by taking into account the possible environmental
458 impact and minor bacterial deposition rate, the transformation process done to
459 UF membranes for MBfRs did not seem to provide any added value.
460 Nonetheless, BWt-NF is especially interesting because the transformation into
461 NF cleaned the surfaces and improved the conditioning and bacterial
462 attachment, maintaining a competitive gas permeability regarding to the
463 aforementioned studies. The thin film dense PA layer of the composite
464 discarded RO membranes could supply competitive bubbleless aeration with
465 low mass resistance, providing a low-cost alternative for MBfRs. Therefore, as
466 is schematically illustrated in Fig. 6, this gas permeability could benefits the

467 biological activity of the biofilms thanks to the high oxygenic-surface provided,
468 which would enhance the pollutant removal, such as MC, by R-MBfRs.

469 **4. Conclusion**

470 This work provides new findings for recycling the huge amount of EoL RO
471 desalination membrane for MBfRs. We prove that chlorination process of
472 certain discarded membranes could be beneficial for removing previous fouling
473 and enhancing the attachment of the desired bacteria. This is of interest to
474 make the membrane recycling totally independent of previous fouling nature,
475 which is important to establish a hypothetical process of membrane recycling for
476 MBfR. Besides, the chlorination process of the BWt-NF did not affect to the MC
477 degradation of the attached strain IM-1; hence, this chemical modification does
478 not have a negative effect for later biological activity. On the other hand, our
479 results also demonstrate that PA-TFC membranes originally manufactured for
480 desalination purposes are capable to allow gases fluxes at low pressures, which
481 is an essential property of MBfRs. Promising outcome is shown by the
482 composite typology in BWt-NF due to display higher air permeabilities than
483 other composite membranes reported in the literature, together with meeting the
484 rest of properties for biofilm developments. Therefore, the current work gives
485 further evidence of the potential of recycling discarded desalination membranes
486 for R-MBfRs, offering a solution for poor quality water treatments and following
487 circular economy principles.

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496 **Compliance with ethical standards**

497 **Conflict of interest**

498 The authors declare that they have no conflict of interest.

499 **References**

500 Abràmoff, M.D., Magalhães, P.J., Ram, S.J., 2004. Image processing with
501 imageJ. *Biophotonics Int.* 11, 36–41. doi:10.1117/1.3589100

502 Ahmed, T., Semmens, M.J., Voss, M.A., 2004. Oxygen transfer characteristics
503 of hollow-fiber, composite membranes. *Adv. Environ. Res.* 8, 637–646.
504 doi:10.1016/S1093-0191(03)00036-4

505 Al-Jeshi, S., Neville, A., 2006. An investigation into the relationship between flux
506 and roughness on RO membranes using scanning probe microscopy.
507 *Desalination* 189, 221–228. doi:10.1016/j.desal.2005.08.001

508 Ambrosi, A., Tessaro, I.C., 2013. Study on Potassium Permanganate Chemical
509 Treatment of Discarded Reverse Osmosis Membranes Aiming their Reuse.
510 *Sep. Sci. Technol.* 48, 1537–1543. doi:10.1080/01496395.2012.745876

511 Buratti, F.M., Manganeli, M., Vichi, S., Stefanelli, M., Scardala, S., Testai, E.,

- 512 Funari, E., 2017. Cyanotoxins: producing organisms, occurrence, toxicity,
513 mechanism of action and human health toxicological risk evaluation. Arch.
514 Toxicol. 91, 1049–1130. doi:10.1007/s00204-016-1913-6
- 515 Casey, E., Glennon, B., Hamer, G., 1999. Review of membrane aerated biofilm
516 reactors. Resour. Conserv. Recycl. 27, 203–215. doi:10.1016/S0921-
517 3449(99)00007-5
- 518 Cerqueira, A.C., Nobrega, R., Jr, G.L.S.A., Dezotti, M., 2013. Oxygen air
519 enrichment through composite membranes: Application to an Aerated
520 Biofilm Reactor 30, 771–779.
- 521 Characklis, W.G., Cooksey, K.E., 1983. Biofilms and Microbial Fouling,
522 Advances in Applied Microbiology. doi:10.1016/S0065-2164(08)70355-1
- 523 Characklis, W.G., Wilderer, P.A., 1989. Structure and Function of Biofilms 436.
- 524 Chen, Y.P., Zhang, P., Guo, J.S., Fang, F., Gao, X., Li, C., 2013. Functional
525 groups characteristics of EPS in biofilm growing on different carriers.
526 Chemosphere 92, 633–638. doi:10.1016/j.chemosphere.2013.01.059
- 527 Cote, P., 1989. Bubble-free aeration transfer analysis. J. Memb. Sci. 47, 91–
528 106.
- 529 Darton, E., Fazel, M., 2001. A statistical review of 150 membrane autopsies.
530 62nd Annu. Int. Water Conf.
- 531 Dawson, R.M., 1998. The toxicology of microcystins. Toxicon 36, 953–962.
532 doi:10.1016/S0041-0101(97)00102-5
- 533 Do, V.T., Tang, C.Y., Reinhard, M., Leckie, J.O., 2012a. Effects of hypochlorous
534 acid exposure on the rejection of salt, polyethylene glycols, boron and
535 arsenic(V) by nanofiltration and reverse osmosis membranes. Water Res.
536 46, 5217–5223. doi:10.1016/j.watres.2012.06.044

- 537 Do, V.T., Tang, C.Y., Reinhard, M., Leckie, J.O., 2012b. Degradation of
538 polyamide nanofiltration and reverse osmosis membranes by hypochlorite.
539 Environ. Sci. Technol. 46, 852–859. doi:10.1021/es203090y
- 540 Donlan, R.M., 2002. Biofilms: microbial life on surfaces. Emerg Infect Diseases.
541 <http://www.cdc.gov/ncidod/EID/vol8no9/02-0063.htm> 8, 881–890.
542 doi:10.3201/eid0809.020063
- 543 Ercin, A.E., Hoekstra, A.Y., 2014. Water footprint scenarios for 2050: A global
544 analysis. Environ. Int. 64, 71–82. doi:10.1016/j.envint.2013.11.019
- 545 European commission, 2008. 19.11.2008.
- 546 Fang, C., Sun, J., Zhang, B., Sun, Y., Zhu, L., Matsuyama, H., 2018.
547 Preparation of positively charged composite nanofiltration membranes by
548 quaternization crosslinking for precise molecular and ionic separations. J.
549 Colloid Interface Sci. 531, 168–180. doi:10.1016/j.jcis.2018.07.034
- 550 García-Pacheco, R., Landaburu-Aguirre, J., Molina, S., Rodríguez-Sáez, L.,
551 Teli, S.B., García-Calvo, E., 2015. Transformation of end-of-life RO
552 membranes into NF and UF membranes: Evaluation of membrane
553 performance. J. Memb. Sci. 495, 305–315.
554 doi:10.1016/j.memsci.2015.08.025
- 555 Geise, G.M., Lee, H.-S., Miller, D.J., Freeman, B.D., Mcgrath, J.E., Paul, D.R.,
556 2010. Effect of MolecularWeight and Temperature on Physical Aging of
557 ThinGlassy Poly(2,6-dimethyl-1,4-phenylene oxide) Films. J. Polym. Sci.
558 Part B Polym. Phys. 45, 1390–1398. doi:10.1002/polb
- 559 Guo, K., Freguia, S., Dennis, P.G., Chen, X., Donose, B.C., Keller, J., Gooding,
560 J.J., Rabaey, K., 2013. Effects of surface charge and hydrophobicity on
561 anodic biofilm formation, community composition, and current generation in

- 562 bioelectrochemical systems. *Environ. Sci. Technol.* 47, 7563–7570.
563 doi:10.1021/es400901u
- 564 Halan, B., Buehler, K., Schmid, A., 2012. Biofilms as living catalysts in
565 continuous chemical syntheses. *Trends Biotechnol.* 30, 453–465.
566 doi:10.1016/j.tibtech.2012.05.003
- 567 He, X., Wang, J., Abdoli, L., Li, H., 2016. Mg²⁺/Ca²⁺ promotes the adhesion of
568 marine bacteria and algae and enhances following biofilm formation in
569 artificial seawater. *Colloids Surfaces B Biointerfaces* 146, 289–295.
570 doi:10.1016/j.colsurfb.2016.06.029
- 571 Herzberg, M., Kang, S., Elimelech, M., 2009. Role of Extracellular Polymeric
572 Substances (EPS) in Biofouling of Reverse Osmosis Membranes. *Environ.*
573 *Sci. Technol.* 43, 4393–4398. doi:10.1021/es900087j
- 574 Ho, L., Tang, T., Hoefel, D., Vigneswaran, B., 2012. Determination of rate
575 constants and half-lives for the simultaneous biodegradation of several
576 cyanobacterial metabolites in Australian source waters. *Water Res.* 46,
577 5735–5746. doi:10.1016/j.watres.2012.08.003
- 578 Hou, F., Li, B., Xing, M., Wang, Q., Hu, L., Wang, S., 2013. Surface modification
579 of PVDF hollow fiber membrane and its application in membrane aerated
580 biofilm reactor (MABR). *Bioresour. Technol.* 140, 1–9.
581 doi:10.1016/j.biortech.2013.04.056
- 582 Jiang, B., Zhang, N., Zhang, L., Sun, Y., Huang, Z., Wang, B., Dou, H., Guan,
583 H., 2018. Enhanced separation performance of PES ultrafiltration
584 membranes by imidazole-based deep eutectic solvents as novel functional
585 additives. *J. Memb. Sci.* 564, 247–258. doi:10.1016/j.memsci.2018.07.034
- 586 Jin, X., Huang, X., Hoek, E.M. V, 2009. Role of specific ion interactions in

- 587 seawater RO membrane fouling by alginic acid. *Environ. Sci. Technol.* 43,
588 3580–3587. doi:10.1021/es8036498
- 589 Jones, G.J., Bourne, D.G., Blakeley, R.L., Doelle, H., 1994. Degradation of the
590 Cyanobacterial Hepatotoxin Microcystin by Aquatic Bacteria. *Nat. Toxins* 2,
591 228–235.
- 592 Kwon, Y.N., Leckie, J.O., 2006. Hypochlorite degradation of crosslinked
593 polyamide membranes. II. Changes in hydrogen bonding behavior and
594 performance. *J. Memb. Sci.* 282, 456–464.
595 doi:10.1016/j.memsci.2006.06.004
- 596 Landaburu-Aguirre, J., García-Pacheco, R., Molina, S., Rodríguez-Sáez, L.,
597 Rabadan, J., García-Calvo, E., 2016. Fouling prevention, preparing for re-
598 use and membrane recycling. Towards circular economy in RO
599 desalination. *Desalination* 393, 16–30. doi:10.1016/j.desal.2016.04.002
- 600 Lawler, W., Alvarez-Gaitan, J., Leslie, G., Le-Clech, P., 2015. Comparative life
601 cycle assessment of end-of-life options for reverse osmosis membranes.
602 *Desalination* 357, 45–54. doi:10.1016/j.desal.2014.10.013
- 603 Lawler, W., Antony, A., Cran, M., Duke, M., Leslie, G., Le-Clech, P., 2013.
604 Production and characterisation of UF membranes by chemical conversion
605 of used RO membranes. *J. Memb. Sci.* 447, 203–211.
606 doi:10.1016/j.memsci.2013.07.015
- 607 Lawton, L. a., Robertson, P.K.J., 1999. Physico-chemical treatment methods for
608 the removal of microcystins (cyanobacterial hepatotoxins) from potable
609 waters. *Chem. Soc. Rev.* 28, 217–224. doi:10.1039/a805416i
- 610 Lezcano, M.Á., Morón-López, J., Agha, R., López-Heras, I., Nozal, L., Quesada,
611 A., El-Shehawy, R., 2016. Presence or absence of mlr genes and nutrient

- 612 concentrations co-determine the microcystin biodegradation efficiency of a
613 natural bacterial community. *Toxins (Basel)*. 8. doi:10.3390/toxins8110318
- 614 Li, J., Li, R., Li, J., 2017. Current research scenario for microcystins
615 biodegradation – A review on fundamental knowledge, application
616 prospects and challenges. *Sci. Total Environ.* 595, 615–632.
617 doi:10.1016/j.scitotenv.2017.03.285
- 618 Li, J., Peng, L., Li, J., Qiao, Y., 2014. Divergent responses of functional gene
619 expression to various nutrient conditions during microcystin-LR
620 biodegradation by *Novosphingobium* sp. THN1 strain. *Bioresour. Technol.*
621 156, 335–341. doi:10.1016/j.biortech.2013.12.118
- 622 Li, Q., Elimelech, M., 2004. Organic fouling and chemical cleaning of
623 nanofiltration membranes: Measurements and mechanisms. *Environ. Sci.*
624 *Technol.* 38, 4683–4693. doi:10.1021/es0354162
- 625 Li, T., Liu, J., Bai, R., 2008. Membrane aerated biofilm reactors: a brief current
626 review. *Recent Pat. Biotechnol.* 2, 88–93.
627 doi:10.2174/187220808784619739
- 628 Li, Y., Zhang, K., 2018. Pilot scale treatment of polluted surface waters using
629 membrane-aerated biofilm reactor (MABR). *Biotechnol. Biotechnol. Equip.*
630 32, 376–386. doi:10.1080/13102818.2017.1399826
- 631 M.Mekonnen, M., Hoekstra, Y.A., 2016. Four Billion People Experience Water
632 Scarcity. *Sci. Adv.* 2, 1–7. doi:10.1126/sciadv.1500323
- 633 Mangwani, N., Shukla, S.K., Rao, T.S., Das, S., 2014. Calcium-mediated
634 modulation of *Pseudomonas mendocina* NR802 biofilm influences the
635 phenanthrene degradation. *Colloids Surfaces B Biointerfaces* 114, 301–
636 309. doi:10.1016/j.colsurfb.2013.10.003

- 637 Martin, K.J., Boltz, J.P., Nerenberg, R., 2012. The Membrane Biofilm Reactor
638 (MBfR) for Wastewater Treatment: Applications, Design Considerations,
639 and Technology Outlook. *Proc. Water Environ. Fed.* 2012, 4032–4044.
640 doi:10.2175/193864712811708347
- 641 Martin, K.J., Nerenberg, R., 2012. The membrane biofilm reactor (MBfR) for
642 water and wastewater treatment: Principles, applications, and recent
643 developments. *Bioresour. Technol.* 122, 83–94.
644 doi:10.1016/j.biortech.2012.02.110
- 645 Molina, S., García Pacheco, R., Rodríguez Sáez, L., García-Calvo, E.,
646 Campos Pozuelo, E., Zarco Martínez, D., González de la Campa, J., De
647 Abajo González, J., 2015. Transformation of end-of-life RO membrane into
648 recycled NF and UF membranes, surface characterization, in: *The*
649 *International Desalination Association World Congress on Desalination and*
650 *Water Reuse.* p. 20.
- 651 Molina, S., Landaburu-Aguirre, J., Rodríguez-Sáez, L., García-Pacheco, R., de
652 la Campa, J.G., García-Calvo, E., 2018. Effect of sodium hypochlorite
653 exposure on polysulfone recycled UF membranes and their surface
654 characterization. *Polym. Degrad. Stab.* 150, 46–56.
655 doi:10.1016/j.polymdegradstab.2018.02.012
- 656 Morón-López, J., Nieto-Reyes, L., El-Shehawy, R., 2017. Assessment of the
657 influence of key abiotic factors on the alternative microcystin degradation
658 pathway(s) (mlr –): A detailed comparison with the mlr route (mlr +). *Sci.*
659 *Total Environ.* 599–600, 1945–1953. doi:10.1016/j.scitotenv.2017.04.042
- 660 Morón-López, J., Nieto-Reyes, L., Senán-Salinas, J., Molina, S., El-Shehawy,
661 R., 2019. Recycled desalination membranes as a support material for

- 662 biofilm development: A new approach for microcystin removal during water
663 treatment. *Sci. Total Environ.* 647, 785–793.
664 doi:10.1016/j.scitotenv.2018.07.435
- 665 Motlagh, A.R.A., LaPara, T.M., Semmens, M.J., 2008. Ammonium removal in
666 advective-flow membrane-aerated biofilm reactors (AF-MABRs). *J. Memb.*
667 *Sci.* 319, 76–81. doi:10.1016/j.memsci.2008.03.020
- 668 Nerenberg, R., 2016. The membrane-biofilm reactor (MBfR) as a counter-
669 diffusional biofilm process. *Curr. Opin. Biotechnol.* 38, 131–136.
670 doi:10.1016/j.copbio.2016.01.015
- 671 Nerenberg, R., 2005. Membrane Biofilm Reactors for Water and Wastewater
672 Treatment. *A Semin. Adv. Water Wastewater Treat.* 1–20.
- 673 Nguyen, T., Roddick, F.A., Fan, L., 2012. Biofouling of water treatment
674 membranes: A review of the underlying causes, monitoring techniques and
675 control measures. *Membranes (Basel)*. 2, 804–840.
676 doi:10.3390/membranes2040804
- 677 Ontiveros-Valencia, A., Zhou, C., Zhao, H., Tang, Y., Rittmann, B.E., 2018.
678 Managing microbial communities in membrane biofilm reactors. *Appl.*
679 *Microbiol. Biotechnol.* 9003–9014.
- 680 Paerl, H.W., Paul, V.J., 2012. Climate change: Links to global expansion of
681 harmful cyanobacteria. *Water Res.* 46, 1349–1363.
682 doi:10.1016/j.watres.2011.08.002
- 683 Peng Lee, K., C. Arnot, T., Mattia, D., 2011. A Review of Reverse Osmosis
684 Membrane Materials for Desalination – Development to date and Future
685 Potential. *Authors* 51, 32–37.
- 686 Raval, H.D., Chauhan, V.R., Raval, A.H., Mishra, S., 2012. Rejuvenation of

- 687 discarded RO membrane for new applications. *Desalin. Water Treat.* 48,
688 349–359. doi:10.1080/19443994.2012.704727
- 689 Rodríguez, J.J., Jiménez, V., Trujillo, O., Veza, J., 2002. Reuse of reverse
690 osmosis membranes in advanced wastewater treatment. *Desalination* 150,
691 219–225. doi:10.1016/S0011-9164(02)00977-3
- 692 Rothmund, C., Amann, R., Klugbauer, S., Manz, W., Bieber, C., Schleifer,
693 K.H., Wilderer, P., 1996. Microflora of 2,4-dichlorophenoxyacetic acid
694 degrading biofilms on gas permeable membranes. *Syst. Appl. Microbiol.*
695 19, 608–615. doi:10.1016/S0723-2020(96)80033-6
- 696 Santiago-Morales, J., Amariei, G., Letón, P., Rosal, R., 2016. Antimicrobial
697 activity of poly(vinyl alcohol)-poly(acrylic acid) electrospun nanofibers.
698 *Colloids Surfaces B Biointerfaces* 146, 144–151.
699 doi:10.1016/j.colsurfb.2016.04.052
- 700 Semmens, M.J., 2005. *Membrane Technology: Pilot Studies of Membrane-*
701 *Aerated Bioreactors.* Water Environment Research Foundation, Alexandria.
702 doi:10.2166/9781843397045
- 703 Siddique, R., Khatib, J., Kaur, I., 2008. Use of recycled plastic in concrete: A
704 review. *Waste Manag.* 28, 1835–1852. doi:10.1016/j.wasman.2007.09.011
- 705 Simon, A., Nghiem, L.D., Le-Clech, P., Khan, S.J., Drewes, J.E., 2009. Effects
706 of membrane degradation on the removal of pharmaceutically active
707 compounds (PhACs) by NF/RO filtration processes. *J. Memb. Sci.* 340, 16–
708 25. doi:10.1016/j.memsci.2009.05.005
- 709 Subramani, A., Huang, X., Hoek, E.M. V., 2009. Direct observation of bacterial
710 deposition onto clean and organic-fouled polyamide membranes. *J. Colloid*
711 *Interface Sci.* 336, 13–20. doi:10.1016/j.jcis.2009.03.063

- 712 Syron, E., Casey, E., 2008. Membrane-aerated biofilms for high rate
713 biotreatment: Performance appraisal, engineering principles, scale-up, and
714 development requirements. *Environ. Sci. Technol.* 42, 1833–1844.
715 doi:10.1021/es0719428
- 716 Tang, Y., Zhou, C., Van Ginkel, S.W., Ontiveros-Valencia, A., Shin, J.,
717 Rittmann, B.E., 2012. Hydrogen permeability of the hollow fibers used in
718 H₂-based membrane biofilm reactors. *J. Memb. Sci.* 407–408, 176–183.
719 doi:10.1016/j.memsci.2012.03.040
- 720 Van Der Bruggen, B., Vandecasteele, C., Gestel, T. Van, Doyenb, W., Leysenb,
721 R., 2003. Review of Pressure-Driven Membrane Processes. *Environ. Prog.*
722 22, 46–56. doi:10.1002/ep.670220116
- 723 Vörösmarty, C.J., Green, P., Salisbury, J., Lammers, R.B., 2000. Global Water
724 Resources: Vulnerability from Climate Change and Population Growth.
725 *Science* (80-.). 289, 284–288. doi:10.1126/science.289.5477.284
- 726 Xu, B., Mao, D., Luo, Y., Xu, L., 2011. Sulfamethoxazole biodegradation and
727 biotransformation in the water-sediment system of a natural river.
728 *Bioresour. Technol.* 102, 7069–7076. doi:10.1016/j.biortech.2011.04.086
- 729 Xu, J., Wang, Z., Wei, X., Yang, S., Wang, J., Wang, S., 2013. The chlorination
730 process of crosslinked aromatic polyamide reverse osmosis membrane:
731 New insights from the study of self-made membrane. *Desalination* 313,
732 145–155. doi:10.1016/j.desal.2012.12.020
- 733 Zhao, F., Xu, K., Ren, H., Ding, L., Geng, J., Zhang, Y., 2015. Combined effects
734 of organic matter and calcium on biofouling of nanofiltration membranes. *J.*
735 *Memb. Sci.* 486, 177–188. doi:10.1016/j.memsci.2015.03.032
- 736 Zhou, C., Ontiveros-valencia, A., Nerenberg, R., Tang, Y., Zhou, C., 2019.

737 Hydrogenotrophic Microbial Reduction of Oxyanions With the Membrane

738 Biofilm Reactor 9, 1–14. doi:10.3389/fmicb.2018.03268

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Type	Zeta potential (mV)	Contact angle (°)	Roughness	
			Ra (nm)	Rq (nm)
BWd ^a	-31.68 ± 4.89 ^{AB}	50.74 ± 2.96 ^C	32.01 ± 4.77 ^B	42.69 ± 5.26 ^A
BWt-NF	-35.19 ± 4.23 ^A	45.66 ± 1.55 ^D	32.18 ± 4.01 ^B	40.71 ± 5.04 ^A
BWt-UF	-22.49 ± 5.22 ^B	51.85 ± 2.14 ^C	4.77 ± 0.42 ^C	6.19 ± 0.47 ^B
SWd ^a	-30.53 ± 3.40 ^{AB}	29.48 ± 1.54 ^E	88.75 ± 28.89 ^A	98.71 ± 30.03 ^A
SWt-NF	-37.11 ± 4.42 ^A	68.19 ± 2.79 ^A	47.61 ± 12.05 ^B	58.64 ± 13.59 ^A
SWt-UF	-33.55 ± 4.48 ^A	60.23 ± 1.25 ^B	4.32 ± 0.63 ^C	5.74 ± 0.93 ^B

^aDiscarded RO membranes already reported (Morón-López et al., 2019).

Table 1. Zeta potential, contact angle and roughness of the transformed EoL membranes. Statistics analyses are shown per column, where each different capital letter indicates the significant differences between membranes at $p < 0.05$ after the one-way ANOVA.

Type	Air permeability (Barrer)	Air flux (J) ($\text{Ncm}^3 \cdot \text{cm}^{-2} \cdot \text{s}^{-1} \cdot \text{cmHg}^{-1}$)
BWd	$1.60 \cdot 10^{-7}$	$8.00 \cdot 10^{-3}$
BWt-NF	$1.01 \cdot 10^{-7}$	$5.06 \cdot 10^{-3}$
BWt-UF	$1.79 \cdot 10^{-4}$	$1.79 \cdot 10^{-2}$
SWd	$4.89 \cdot 10^{-9}$	$2.44 \cdot 10^{-4}$
SWt-NF	$4.81 \cdot 10^{-9}$	$3.01 \cdot 10^{-4}$
SWt-UF	$2.78 \cdot 10^{-5}$	$2.78 \cdot 10^{-3}$

* 1 Barrier = $10^{-10} \text{Ncm}^3 \cdot \text{cm} \cdot \text{cm}^{-2} \cdot \text{s}^{-1} \cdot \text{cmHg}^{-1}$

Table 2. Air permeability and flux (J) at room temperature and pressure at 1 bar.

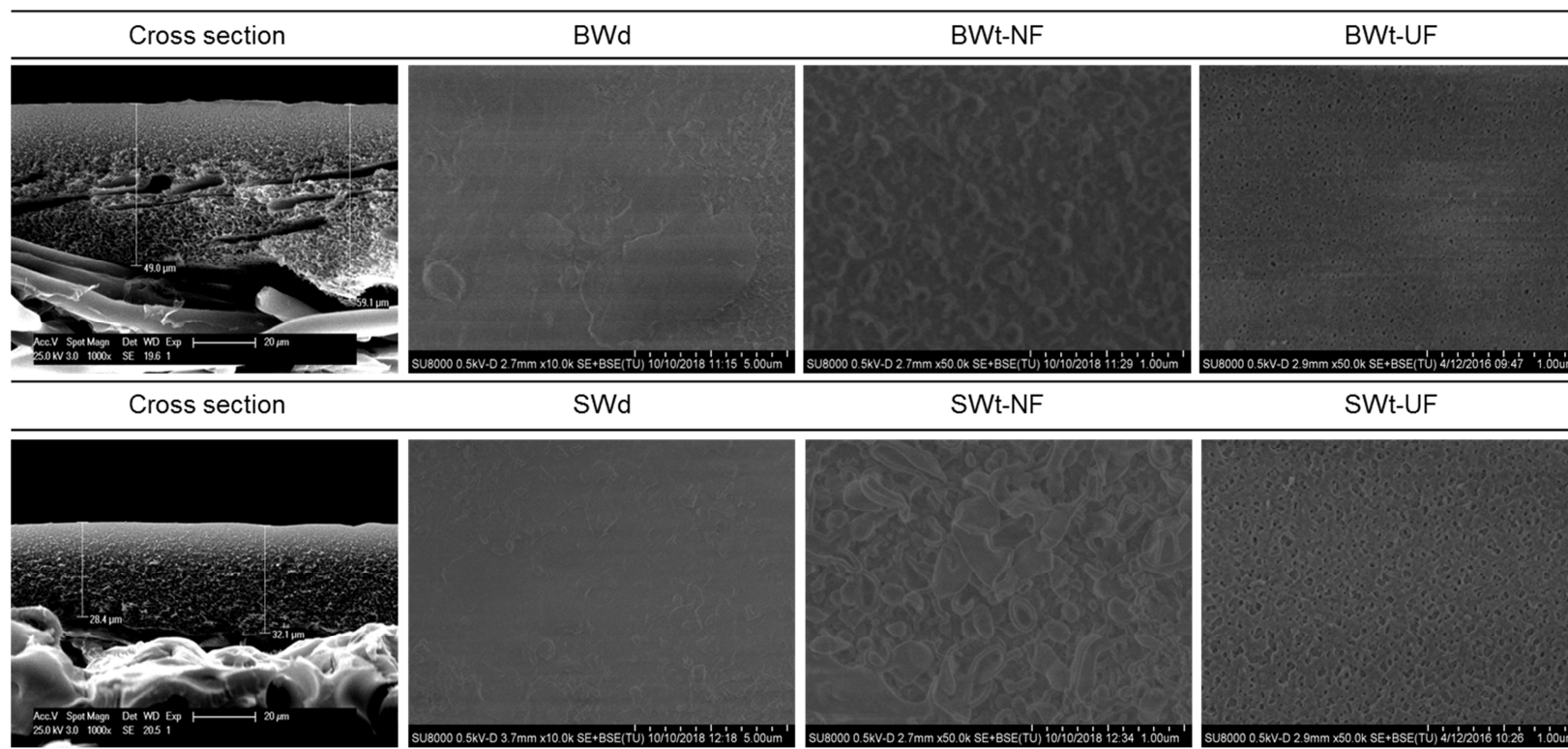


Fig.1. Cross-section and surface SEM images of the EoL desalination membranes before (BWd, SWd) and after chemical attack with NaOCl at doses of 6200 ppm·h (BWt-NF and SWt-NF) and 300000 ppm·h (BWt-UF and SWt-UF).

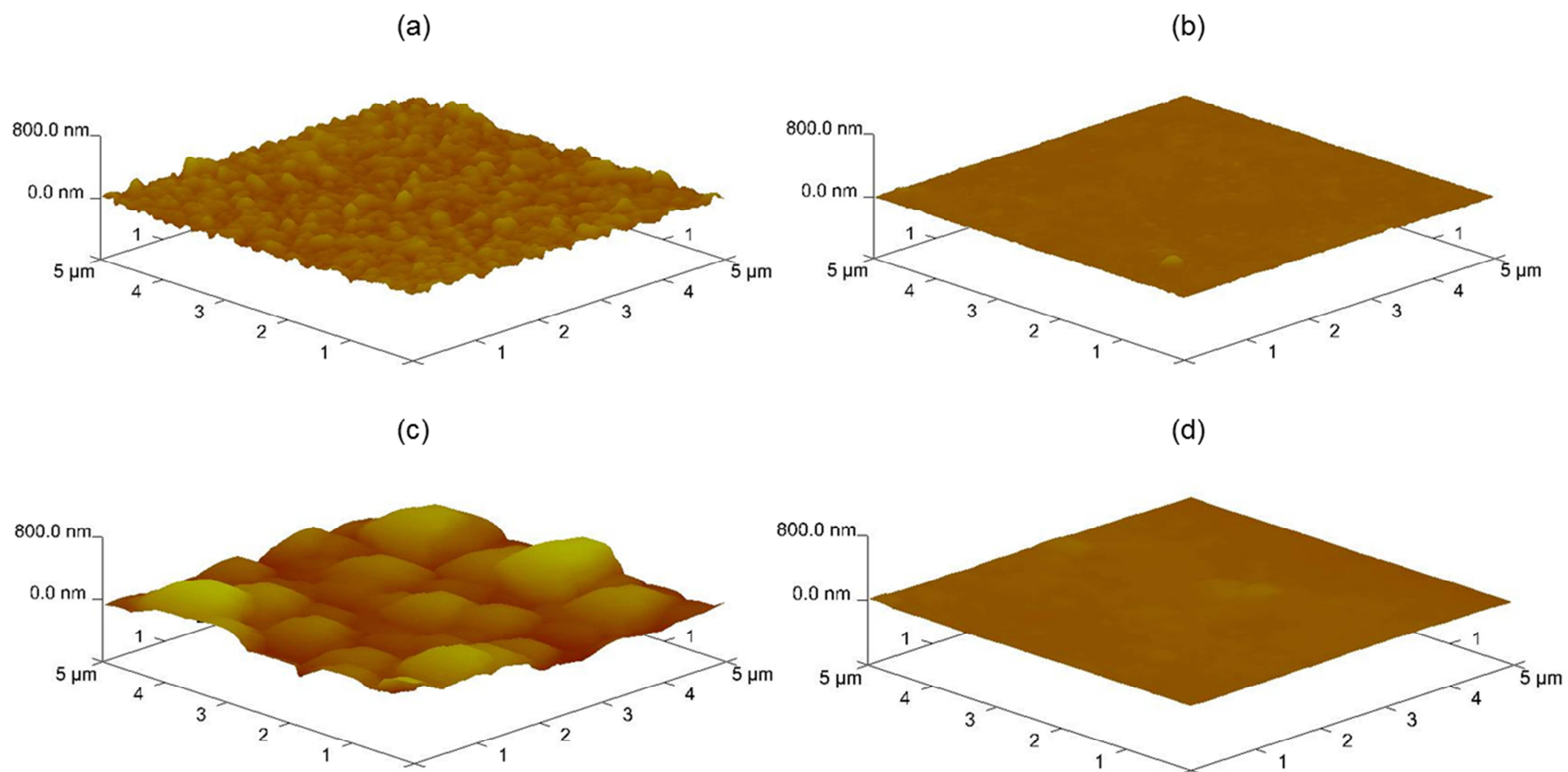


Fig.2. AFM images of membranes: (a) BWt-NF, (b) BWt-UF (c) SWt-NF and (d) SWt-UF

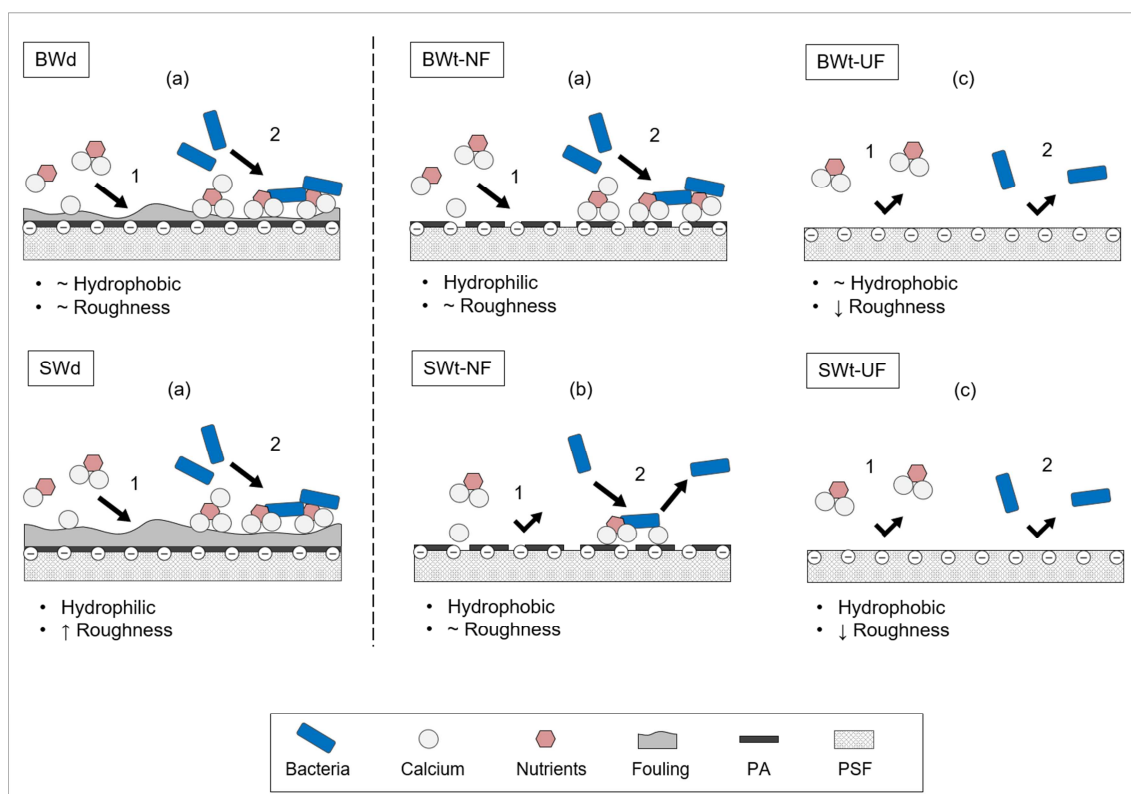
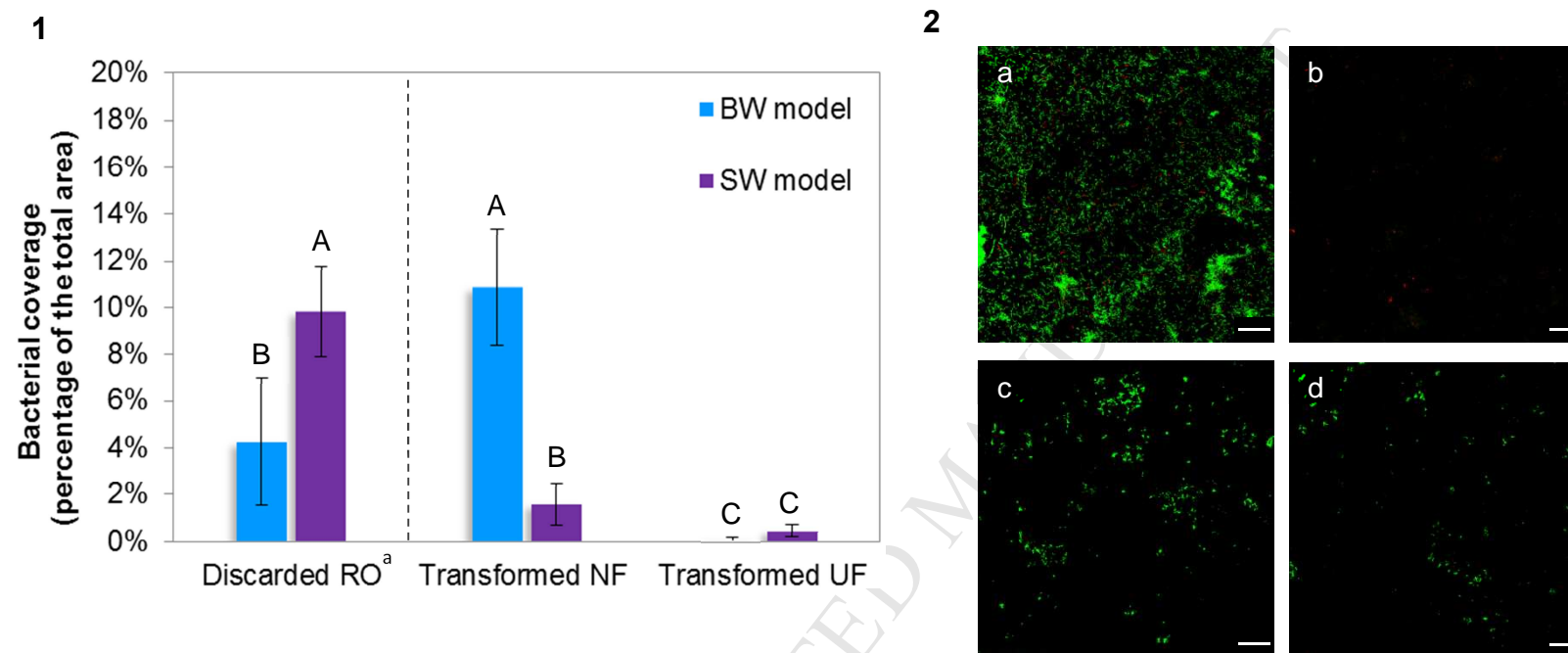
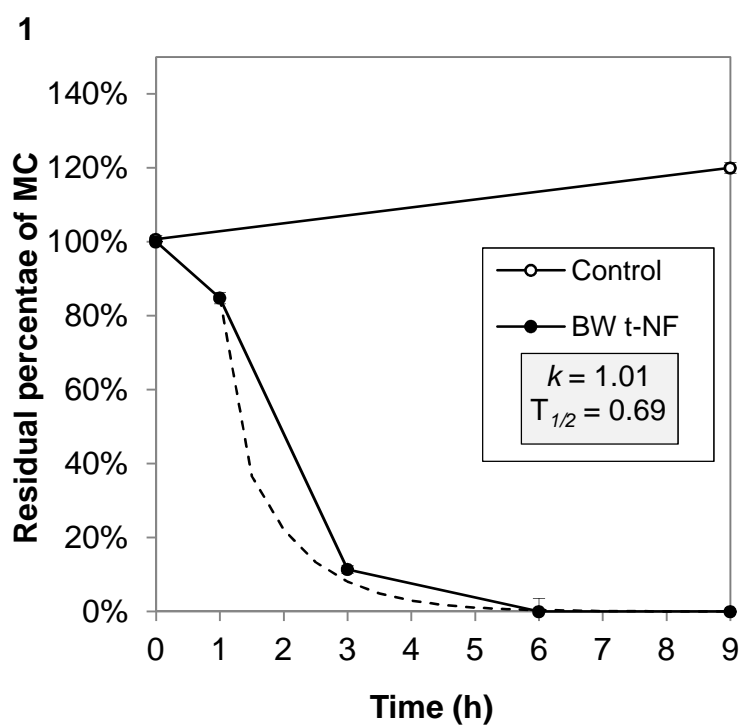


Fig. 3. Schematic illustration of early biofilm stages into tested EoL membranes. (1) Indicates the first conditioning stage, while (2) is bacterial deposition. Letters represent: (a) good conditioning and subsequent bacterial attachment; (b) bad conditioning and low bacterial deposition; (c) no conditioning and no bacterial attachment. The dotted line separates the untransformed discarded membranes from the transformed ones.



^a These results have been already reported (Morón-López et al., 2019).

Fig. 4. Bacterial deposition on different transformed membrane surfaces: **(1)** Percentage of bacterial coverage. Statistic analyses shows different capital letters when indicating significant differences between membranes at $p < 0.05$ after a one-way ANOVA; **(2)** CLSM images after staining transformed membranes with the life/dead bacterial kit. (a) BWt-NF, (b) BWt-UF and (c) SWt-NF and (d) SWt-UF.



2

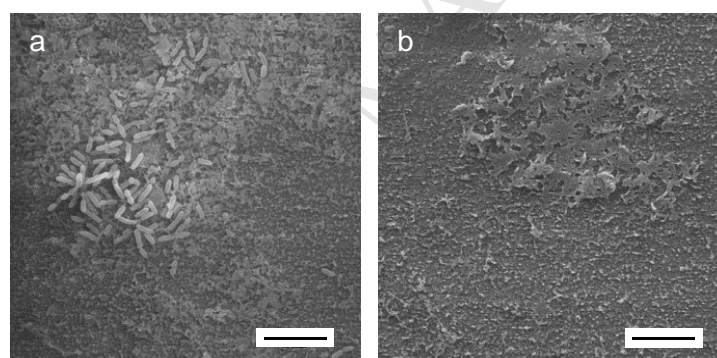


Fig. 5. MC degradation capability of strain IM-1 attached to the BWt-NF membrane. (1) MC degradation during the second MC exposure (after the acclimation time). The dotted line indicates the pseudo first-order model prediction. The MC error bars represent the standard deviation of two technical replicates. (2) The SEM images of the BWt-NF membrane surface with (a) strain IM-1 attached; (b) no bacterial deposition (control). Bar shows 5 μm .

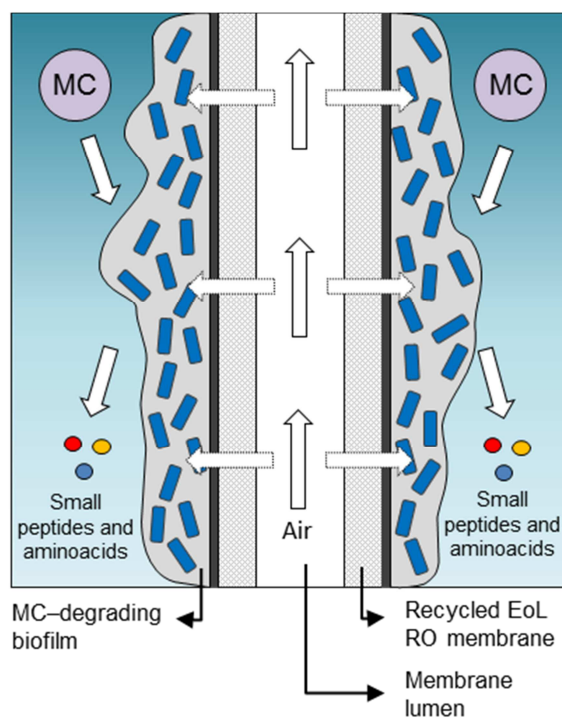


Fig.6. Schematic illustration of the aerobic MC-degrading biofilm attached onto recycled EoL RO membrane adapted to MBfRs.

Highlights

- Tested doses of chlorine trigger different changes on membrane surfaces depending on the polyamide nature and treated water sources during their lifespan (fouling).
- Chlorination process increase hydrophilicity and maintain roughness in some membranes, improving the surface conditioning and bacterial deposition.
- The chlorine attack does not affect the biological activity of the MC-degrading bacteria once attached into BWt-NF.
- Recycled thin film composite polyamide membranes allow competitive gas permeability at low pressure, which is a key characteristic for membrane biofilm reactors (MBfRs).